
KODAK Image Station 4000R

User's Guide

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rad PADD patent number 6,346,707 Issued 2/12/2002.

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Table of Contents

Introduction	1-1
About the User's Guide	1-2
Navigating through the User's Guide	1-3
What's in the IS4000R Package	1-4
System Requirements	1-5
Registering the IS4000R System	1-6
Obtaining Technical Support	1-7
Hardware Overview	2-1
Front View of the IS4000R	2-2
Rear View of the IS4000R	2-3
Sample Work Area	2-4
Closed Optical Path Image (COPI) Chamber	2-7
IS4000R Capture System Chamber	2-10
The KODAK rad PADD (Optional Accessory)	2-13
Software Overview	3-1
IS4000R Acquire Window	3-4
Preview Window	3-18
Capture Status Windows	3-19
Preferences Window	3-29
Setting Up for Capture	4-1
Sample Illumination/Detection Guidelines	4-1
Preparing the IS4000R for Imaging	4-5
Launching the Software	4-6
Orienting the Sample for Capture	4-8
Previewing the Image	4-10
Choosing the Filter and Illumination Conditions	4-12

Setting the <i>f</i> -Stop	4-16
Image Capture Guidelines Summary	4-17
Using the Focus Tool	4-18
Capturing Images	5-1
Using the Standard Exposure Option	5-2
Using the Time Lapse Exposure Option	5-8
Using the Progressive Exposure Option	5-13
Using the rad PADD Exposure Option	5-18
Applying an Automatic Illumination Reference File	5-23
Generating an Illumination Reference File Library	5-24
Custom Capture Settings	5-26
Maintaining the System	6-1
Regular Maintenance	6-2
Replacing or Changing an Emission Filter	6-3
rad PADD Care and Maintenance	6-6
Transilluminator Lid Bulb Replacement	6-7
Troubleshooting the System	7-1
Obtaining Technical Support	7-2
Common Instrument Questions	7-3
Common Image Questions	7-4
Common Software Questions	7-7
Installing the System	8-1
Environmental Requirements	8-2
Electrical Requirements	8-2
Space Requirements	8-2
Installing the IS4000R	8-3
Installing the KODAK Image Station Transilluminator Lid (Optional Accessory)	8-4
Installing KODAK Software–Windows	8-8
Installing KODAK Software–Macintosh	8-14
Launching KODAK Molecular Imaging Software for the First Time	8-23
Repackaging or Moving the IS4000	8-30

Warranty & Regulatory Information	9-1
IS4000R Limited Warranty	9-2
Software License Agreement	9-6
Regulatory Information	9-9
Appendix A: Digital Imaging Concepts	A-1
Appendix B: UV Epi-Illumination	B-1
Appendix C: Image Correction.	C-1
Glossary	G-1



Introduction

Thank you for purchasing the KODAK Image Station 4000R (IS4000R) system. We know you will be pleased with this multi-purpose system for imaging of chemiluminescence, radioactive, fluorescence, and chromogenic gels and blots. The IS4000R is as sensitive as autoradiography films for most chemiluminescent blots and detects fluorescent signals at picomole to femtomole levels. In addition, when used with the patented rad PADD (radioisotopic phosphor assisted detection device), you can capture high energy beta-emitting radioisotopes such as ^{32}P .

The affordable system combines a thermoelectrically cooled charged-coupled device (CCD) camera, an innovative closed optical path image (COPI) style chamber, and the powerful KODAK Molecular Imaging Software (KODAK MI) to provide a sensitive and quantitative benchtop imaging solution for your laboratory.

The cooled CCD camera, combined with the 10X zoom lens, captures true 16-bit images that can be accumulated to 32-bit files at a pixel density of 2048 x 2048. This generates an image with a resolution of 10 microns/pixel at full zoom. The COPI chamber provides a light-tight environment, so there is no need to use a darkroom. The IS4000R Acquire window, within KODAK MI, accumulates and displays multiple image captures so you can view and optimize the quality of your image.

The KODAK Transilluminator Lid Accessory provides built-in UV transillumination that is ideal for imaging fluorescent samples like ethidium bromide stained gels. The White Light PADD (Phosphor Assisted Detection Device) can be placed over the transilluminator to convert UV light to white light and is an excellent way to image Coomassie gels or for densitometry of films. The transilluminator lid is easily installed by replacing your standard IS4000R lid.

About the User's Guide

The KODAK Image Station 4000R User's Guide provides you with all of the information you need to capture images. It is designed to be used in conjunction with the KODAK Molecular Imaging Software User's Guide and manuals you received with your computer.

If you are like most users, you will want to get started right away; but before you do, you need to have basic computer skills. Refer to your computer manual to become familiar with the basic operation of your computer.

Conventions

This User's Guide uses the following conventions:

- ✓ Windows and dialog boxes are displayed as they appear in Windows XP. They may appear differently on your screen depending your computer's operating system.
- ✓ Menu commands, tool names, and window names are capitalized.
- ✓ Tips, examples, and notes appear in the text as follows:

 NOTE: Molecular Imaging Systems provides maximum performance products.

- ✓ Important safety warnings appear in the text as follows:



WARNING: This symbol is used in the User's Guide to designate a warning or caution statement.



WARNING: This symbol is used in the User's Guide to designate where electrical shock is possible.



WARNING: This symbol is used in the User's Guide to designate when there is a potential exposure to hazardous ultraviolet light.



WARNING: This symbol is used on the instrument to indicate protective earth.

Navigating through the User's Guide

The KODAK IS4000R User's Guide is divided into the chapters listed below. In each chapter, you will find helpful hints and troubleshooting information.

- ✓ Chapter 1: *Introduction* provides you with information on what's in your package, how to navigate through this User's Guide, and how to obtain technical support.
- ✓ Chapter 2: *Hardware Overview* details the instrument hardware related to the IS4000R.
- ✓ Chapter 3: *Software Overview* reviews the main windows specific to IS4000R image capture.
- ✓ Chapter 4: *Setting Up for Capture* describes the steps necessary to set up your sample and system for imaging.
- ✓ Chapter 5: *Capturing Images* walks you, step-by-step, through the process of capturing images using standard, time lapse, progressive and rad PADD exposures.
- ✓ Chapter 6: *Maintaining the System* provides guidelines for the care and maintenance of your system.
- ✓ Chapter 7: *Troubleshooting the System* provides an outline for solving problems you may encounter.
- ✓ Chapter 8: *Installing the System* describes the installation process of your IS4000R.
- ✓ Chapter 9: *Warranty & Regulatory Information* provides detailed information on the IS4000R warranty, software licenses, and regulatory information.
- ✓ Appendix A: *Digital Imaging Concepts* discusses the basics of resolution and image histograms, and how they apply to the IS4000R.
- ✓ Appendix B: *UV Epi-Illumination* describes the specifications of the epi-illumination used in the IS4000R.
- ✓ Appendix C: *Image Correction* reviews the details of CCD, lens, and illumination corrections relating to the IS4000R.
- ✓ *Glossary* defines terms that are used in the User's Guide.

What's in the IS4000R Package

Prior to using your new system, please take a moment to ensure that all necessary parts have been received. If the system you ordered includes a computer, the computer may be shipped separately.

KODAK IS4000R Chamber and Capture System (1)

- ✓ Image Station 4000R instrument (1)
- ✓ Compression Pad, located inside the instrument lid (1)
- ✓ 120 Volt Power Cord (1) and/or 230 Volt Power Cords (2)
- ✓ IEEE 1394 (FireWire) Camera/Computer Interface Cable (1)
- ✓ Light Diffuser (1)
- ✓ Camera Calibration CD (1)
- ✓ Quick Reference Guide (1)
- ✓ User's Guide (1)
- ✓ Registration Form (1)
- ✓ 600 nm WB Em Filter (Ethidium Bromide) (1)
- ✓ rad PADD (Optional Accessory) (1)
- ✓ Image Station Transilluminator Lid Accessory (Optional Accessory) (1)

KODAK Molecular Imaging Software

- ✓ Molecular Imaging Software 3 User Package (1)
 - Molecular Imaging Software CD (1)
 - User Registration Card (1)
 - Molecular Imaging Software User's Guide (3)
 - Copy Protection Device (3)
- (or)
- ✓ Molecular Imaging Software Network Edition (Custom Package)
 - Molecular Imaging Software Network Edition CD (1)
 - Molecular Imaging Software Network Edition Administrator's Manual (1)
 - Molecular Imaging Software User's Guides (3)

System Requirements

These are minimum specifications required, however, we cannot ensure that all hardware and software systems are compatible. For optimal performance, we strongly suggest dedicating a computer exclusively for use with IS4000R and that the IS4000R be the only device on the IEEE 1394 (FireWire) bus.

Minimum System Requirements—Windows

- ✓ Personal computer with an IEEE 1394 (FireWire) and a USB port
 - ✓ Pentium IV (or equivalent) processor greater than 1 GHz, greater than 2 GHz recommended
 - ✓ Windows 2000 (Service Pack 4 or greater) or Windows XP (Service Pack 2 or greater) operating system software
-  NOTE: Check your operating system version by right-clicking on the My Computer icon and then on Properties
- ✓ 17 in. display—1280 x 1024 resolution
 - ✓ Minimum 1 GB of available RAM, 2 GB recommended
 - ✓ Minimum 10 GB of available hard disk space
 - ✓ CD drive, CD-RW drive recommended

Minimum System Requirements—Macintosh

- ✓ Macintosh Power PC G4 processor or better
 - ✓ Personal computer with an IEEE 1394 (FireWire) and a USB port
 - ✓ Mac OS 10.3.8 or greater
-  NOTE: Check your operating system version by selecting About this Mac under the Apple menu item.
- ✓ 17 in. display—1280 x 1024 resolution
 - ✓ Minimum 1 GB of available RAM, 2 GB recommended
 - ✓ Minimum of 10 GB of available hard disk space
 - ✓ CD drive, CD-RW drive recommended

Registering the IS4000R System

To be eligible for maintenance releases, upgrade programs, and to receive new product information, you must register your system by completing and returning the enclosed registration cards. For best product support, please take the time to complete and return the enclosed cards.

Obtaining Technical Support

For technical support, contact Technical Support or your Molecular Imaging Systems dealer. For up to date dealer information, visit our WEB site at www.carestreamhealth.com/go/molecular. When contacting technical support, please have the following information available:

- ✓ The serial number of your IS4000R system located on the back of the unit and the camera serial number located on the camera.
- ✓ The serial number and version number of your KODAK MI software.
 - ☞ NOTE: With KODAK MI running, select About KODAK MI under the Help menu (Windows) or select About KODAK MI under the Apple menu item (Macintosh).
- ✓ The type of computer you are using (make, model).
- ✓ Operating system software version.
 - ☞ NOTE: Check your operating system version by right-clicking on the My Computer icon and then on Properties (Windows) or select About This Mac under the Apple menu item (Macintosh).
- ✓ The type of image you are capturing or analyzing.
- ✓ The problem you are having and what you were doing when the problem occurred. Please note the exact wording of any error messages, including any error numbers displayed.

Molecular Imaging Systems Technical Support

Contact Molecular Imaging Systems Technical Support by:

- ✓ Utilizing our World Wide Web support pages at:
<http://www.carestreamhealth.com/go/molecular>
- ✓ Calling Molecular Imaging Systems Technical Support at:
877-747-4357 or 203-786-5657, between the hours of 8:00 a.m. and 6:00 p.m. (Eastern Standard Time) Monday through Friday
- ✓ E-mailing Molecular Imaging Systems Technical Support at:
molecular-support@kodak.com
- ✓ Faxing Molecular Imaging Systems Technical Support at:
203-786-5656

Hardware Overview

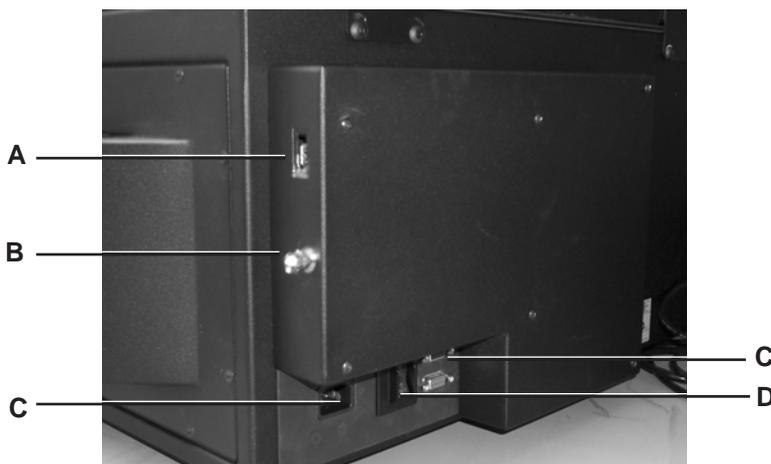
This section provides an overview of the KODAK Image Station 4000R (IS4000R) system. You will review the principles of operation and get a better understanding of the critical components—the Sample Work Area, the closed optical path image (COPI) chamber which also contains the UV illumination source, and the capture system.

Front View of the IS4000R



- A The *Lid* accesses the Sample Work Area and provides a light-tight environment for imaging and capturing fluorescent, luminescent, or radioactive images.
- B The *Sample Work Area* is a water-tight surface with overall dimensions of 52 x 33 cm.
- C The *Platen* is the imaging window that accommodates samples up to 20 x 20 cm.
- D The *Fan* ensures the heat from the radiator dissipates. The *Radiator* dissipates heat from the thermoelectric cooler.
- E The *Capture System Chamber* includes the four position Filter Wheel, a 10X zoom lens, and a cooled CCD camera.
- F The *Closed Optical Path Image (COPI) Chamber* isolates the optical path from the outside environment. It contains the 45° high efficiency mirrors which directs the light from the sample to the camera lens. The internal UV light sources are also contained in the COPI chamber.

Rear View of the IS4000R



- A The *IEEE 1394 (FireWire) Camera Cable Connector* (not shown) couples the camera in the IS4000R to the computer with a special Camera/Computer Interface Cable.



WARNING: Use only the provided IEEE 1394 (FireWire) Camera/Computer Cable to attach the IS4000R to your computer.



WARNING: Be sure to orient the IEEE 1394 (FireWire) Camera/Computer Cable properly with the narrow end pointing down to avoid damage to the circuitry and voiding the warranty.



WARNING: We recommend connecting the IS4000R as the only device on the IEEE 1394 (FireWire) bus.

- B The *BNC Connector* on the supervisor cover is designed for future accessories.
- C The *Power Cord Connector* is the receptacle for the power cord which provides electricity needed to power the instrument.
-  NOTE: If the optional Image Station Transilluminator Lid Accessory is used, the power outlet on the main chassis is connected to the lid and the lid is connected to the wall outlet.
- D The *Circuit Breaker* protects the unit in the unlikely event that there is an electrical short circuit.
- E *RS232 Serial Ports* provide output for future accessories.

Sample Work Area

The Sample Work Area is made up of two main components—the Lid and the Platen. Let's review their features.

Standard Lid

The Standard Lid design provides easy access to the Sample Work Area for positioning a sample in Preview Mode. When closed, the lid creates a light-tight environment for capturing fluorescent, radioisotopic, or luminescent images. When opened, external room light (with the Light Diffuser) provides even white light transillumination.



- A The *Clasp* secures the closed lid to produce a light-tight environment.
- B The *Clip* holds the Compression Pad in the correct position.
- C The *Compression Pad* is a double-sided pad (black side and a white side) with a foam interior. The Compression Pad acts as a good background for imaging and holds thin samples flat and in place when the lid is closed. It can be easily removed to accommodate thicker samples.
- D The *Gasket* surrounding the lid to ensure a light-tight seal for the Sample Work Area.

KODAK Image Station Transilluminator Lid Accessory (Optional)

The KODAK Image Station Transilluminator Lid Accessory provides built-in broadband UV transillumination and is ideal for imaging fluorescent samples like ethidium bromide stained gels. The White Light PADD (Phosphor Assisted Detection Device) can be placed over the transilluminator to convert UV light to white light and is an excellent way to image Coomassie gels or for densitometry of films. The transilluminator lid is easily installed by replacing your standard IS4000R lid.



- A The *UV Switch* powers the UV transilluminator only when the lid is closed and locked into place and the *Illumination Switch* is turned to the ON position. The *Auto Shut-Off Switch* activates the automatic timer which automatically turns off the transilluminator lights after 15 minutes.
- B The *Thumbscrews* (3) hold the White Light PADD in the correct position.
- C The *UV Transilluminator* provides a uniform intense (306 nm peak emission) source of ultraviolet radiation. Although the Sample Work Area is 52 x 33 cm, the UV imaging area is limited to 20 x 20 cm.



WARNING: DANGER—Ultraviolet radiation emitted from this product. Avoid exposure. **ALWAYS WEAR PROTECTIVE CLOTHING. EXPOSURE MAY CAUSE PREMATURE AGING OF THE SKIN AND CANCER. ALWAYS WEAR PROTECTIVE EYEWEAR; FAILURE TO DO SO MAY RESULT IN SEVERE BURNS OR LONG TERM INJURY TO EYE.** Never look directly into the lamp. In individuals with a photosensitivity disease, exposure can cause a wide variety of effects, not just allergenicity.



WARNING: Some medications, cosmetics or diseases may increase your sensitivity to ultraviolet radiation. Consult a physician before operating this product if you are using medications or have a history of skin problems or believe yourself especially sensitive to sunlight. For Laboratory Use Only.

The UV source does not operate when the lid is open. Do not attempt to override this feature.

Platen

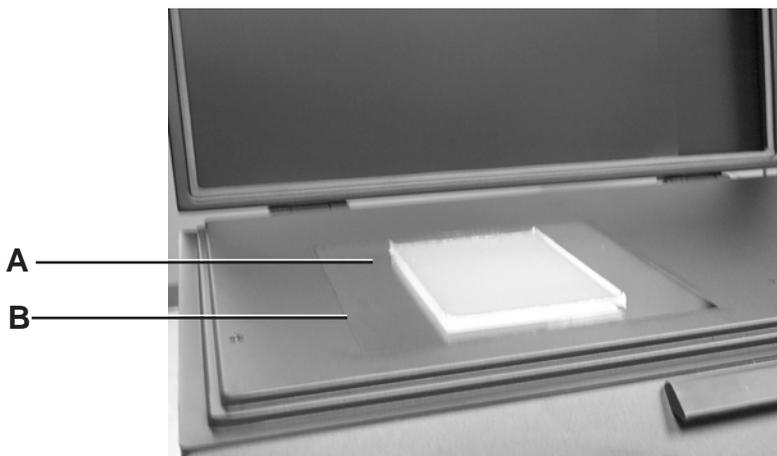
The imaging window is a scratch resistant optical grade material called the Platen. The Platen is framed in a coated steel panel mounted on the unit base. The Sample Work Area is water-tight, has been designed for easy cleanup and is well suited for both wet and dry samples.



WARNING: Do not use metal utensils or tweezers since they may scratch and permanently damage the Platen. This type of damage is not covered by the warranty.



WARNING: Do not expose the Platen to strong chemical solvents such as ketones, hexanes, acids and alkalis that may permanently damage the Platen. This type of damage is not covered by the warranty.

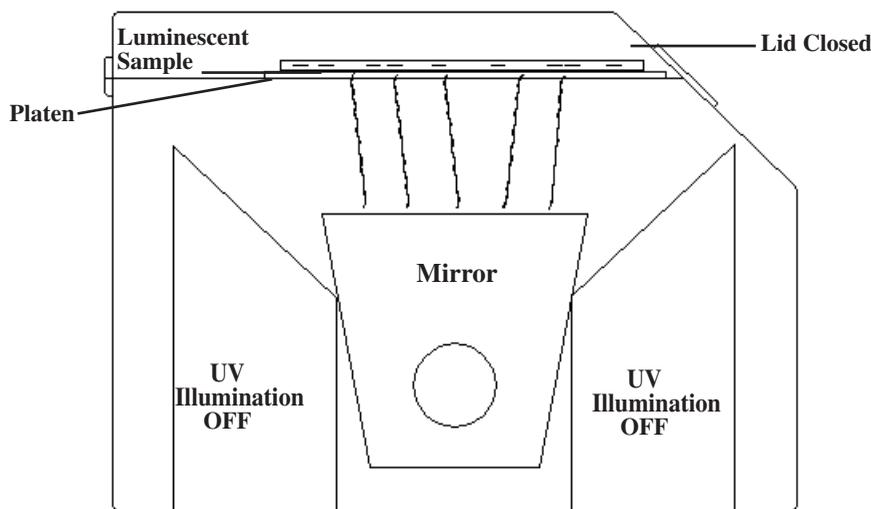


- A The *Platen* is a coated clear plate which accommodates up to a 20 x 20 cm sample for image capture.
- B The *Platen Gasket* lies under the Sample Work Area and creates a water-tight environment.

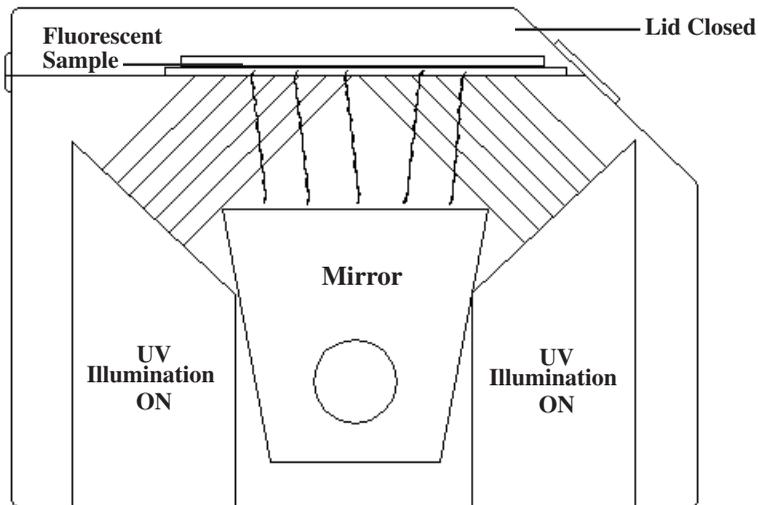
Closed Optical Path Image (COPI) Chamber

The IS4000R provides three standard forms of illumination including transillumination, epi-illumination, and luminescent (no-illumination) modes. An optional lid may be purchased for UV and white light transillumination. In the standard transillumination mode external (room) light (with a Light Diffuser) is utilized for absorbance imaging of samples such as Coomassie gels or autoradiography films. The Light Diffuser, a white plate of translucent acrylic, is placed over the sample to diffuse light uniformly across the sample area. The UV epi-illumination mode utilizes UV bulbs in the illumination chamber to provide 300–400 nm UV excitation of a wide range of fluorochromes. The UV epi-illumination mode can also be used for accurate fluorescence quenching measurements of samples such as TLC plates or chromogenic blots. The “no-illumination” mode is used for luminescent samples, such as chemiluminescent blots or radioactive blots using the rad PADD optional accessory.

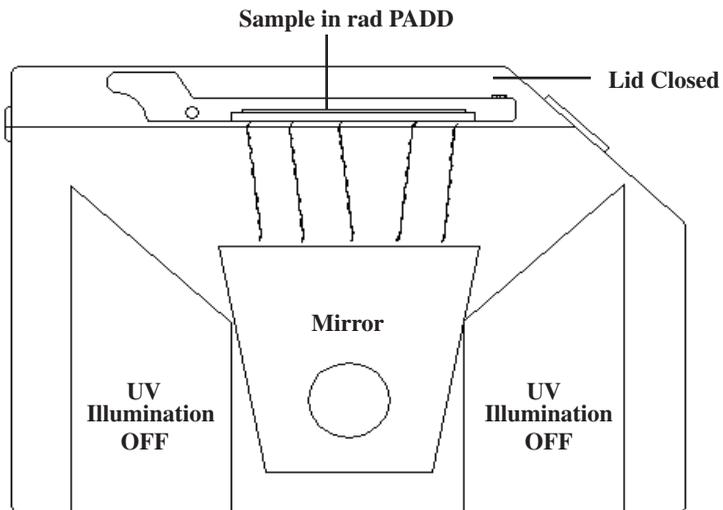
Schematic of IS4000R in “No-Illumination” Mode—Luminescence



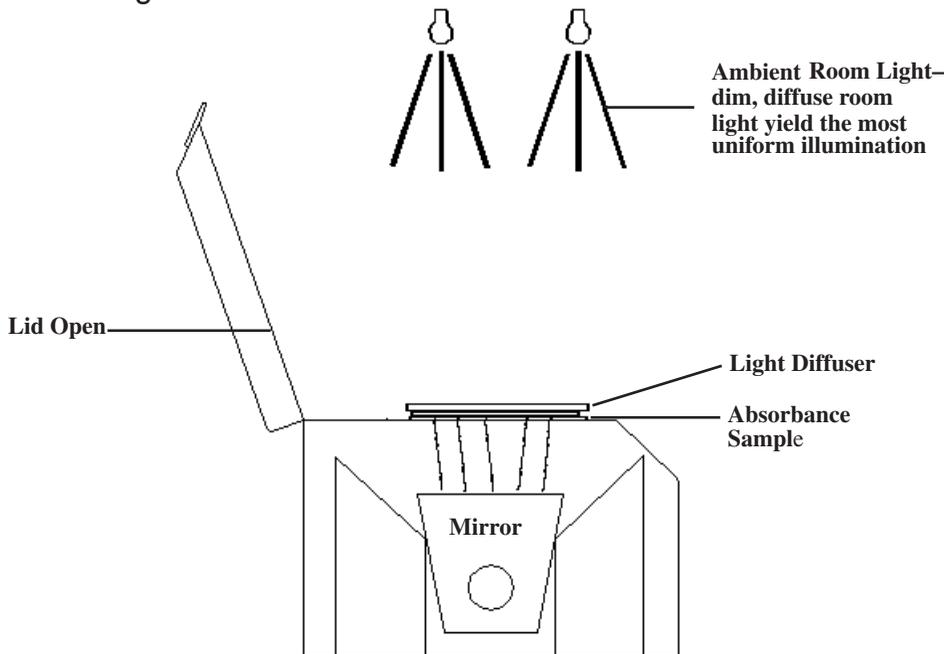
Schematic of IS4000R in UV Epi-Illumination Mode



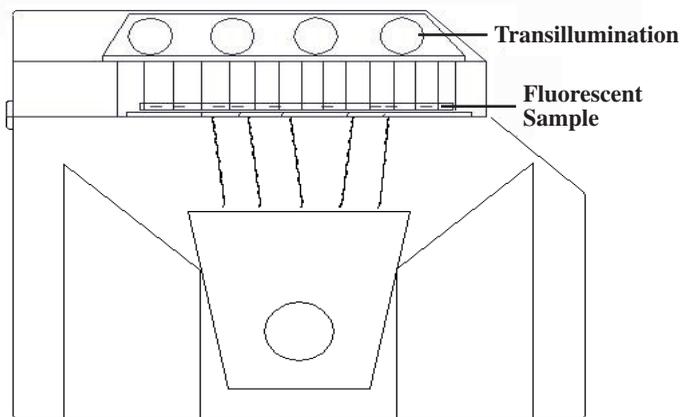
Schematic of IS4000R in “No-Illumination” Mode—Radioactive



Schematic of IS4000R in White Light Transillumination Mode with Ambient Light and Diffuser



Schematic of IS4000R Using the Image Station Transilluminator Lid Accessory



IS4000R Capture System Chamber

The cooled CCD camera, lens, and Filter Wheel are located in the Capture System Chamber.

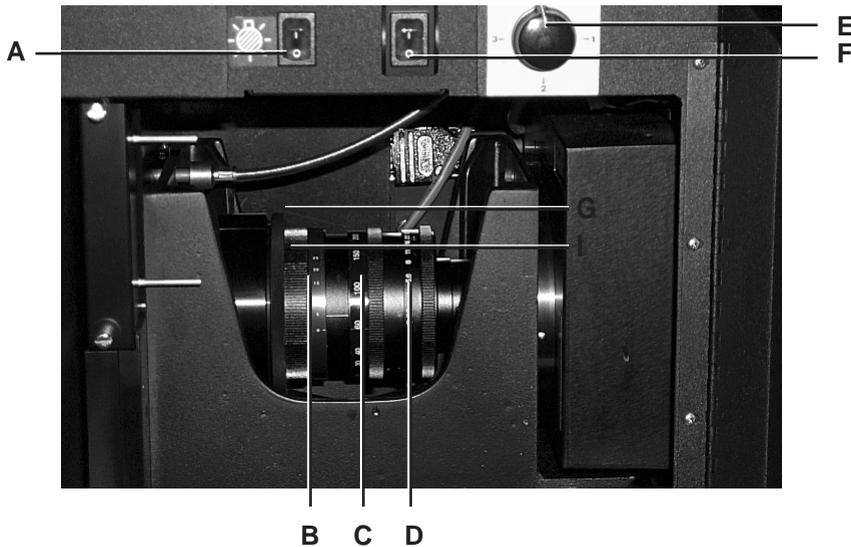
Cooled CCD Camera

The IS4000R camera has been custom designed to provide the highest sensitivity for molecular detection. The camera CCD is thermoelectrically cooled to maximize the sensitivity of the camera. The cooled camera collects the image data on a 2048 x 2048 pixel CCD. Single frame image data is digitized at 16-bits, and presented in software as a 16-bit floating point image with a maximum signal-to-noise ratio of greater than 5000. You can accumulate data from multiple exposures into a 32-bit image file. When you capture and accumulate multiple frames (up to 100 minutes per frame), each frame is added to the previous and the resultant image is available for analysis as a KODAK MI project. In multiple capture mode, you can save independent and/or accumulated images, providing a linear dynamic range signal-to-noise ratio of greater than 5000 times the square root of the number of accumulated images. For instance, a 16-image accumulation file has a maximum range of 20,000, exceeding 4 OD units.

The instrument utilizes a high efficiency mirror in the illumination chamber that projects the light from the sample into the lens system. This unique design minimizes focal length and thereby reduces loss of signal due to light scatter. The lens system is parfocal at all field of view settings. Therefore, once a plane of focus is determined, the lens focus should not need further adjustment.

You can manually adjust the focus, field of view, or aperture (f -stop) settings of the lens. The digital camera has a 10X zoom lens that will capture images from 20 x 20 cm (100 microns/pixel) to 2 x 2 cm (10 microns/pixel).

Capture System Chamber



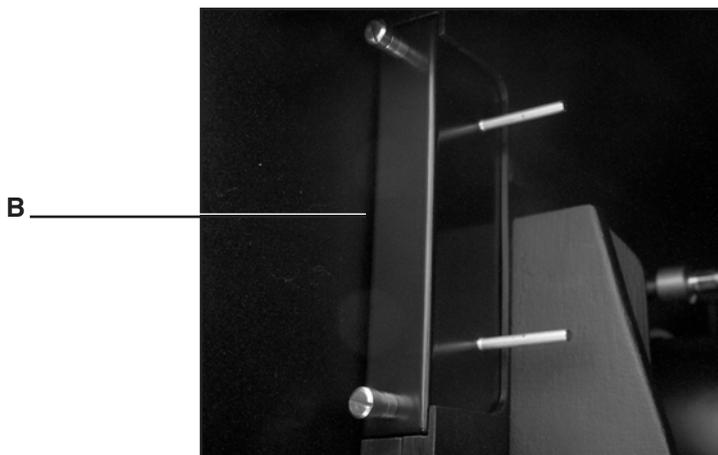
- A The *UV ON/OFF Switch* powers the UV light source. Always turn the switch to the OFF position when not in use to maximize the life of the UV bulbs. An indicator light is displayed when the UV light is powered. The Power Switch must also be powered to operate the UV light source.
- B The *Focus Ring* adjusts the focus of the lens. Once the lens is in focus (parfocal), no adjustment is required at any of the field of view settings. The Focus Ring is demarked to designate the distance in mm (-5–25) of the plane of focus above the Platen. Position 0 is in focus at the Platen.
- C The *Field of View (FOV) Ring* adjusts the field of view setting of the lens from 20 x 20 cm (100 microns/pixel) to 2 x 2 cm (10 microns/pixel).
- D The *f-Stop Ring* regulates the amount of light reaching the sensor for an exposure and influences the depth of focus.
- E The *Filter Selection Dial* changes the filter in front of the lens.
- F The *Power Switch* powers the IS4000R and activates the camera and communication hardware. An indicator is displayed when the camera is on.
- G The *Serial Number Label* identifies your unique camera.
- H The *Lens (labeled B, C and D)* is parfocal. At *f-stop O (Open)*, the maximum amount of light is passed through the lens.
- I The *Cooled CCD Camera* is mounted to the base of the instrument. The camera captures 2048 x 2048 pixel images. To use, turn the Power Switch on.

Filter Wheel

A Filter Wheel is located inside the Capture System Chamber in front of the lens assembly. This wheel has four filter cassette holders, each of which holds a custom 72 mm filter cassette. The IS4000R system includes the 600 WB 35 filter for imaging ethidium bromide stained gels. Molecular Imaging Systems offers additional filters that are suited for a variety of imaging needs. Position “0” is usually reserved as an open (no filter) position for luminescence, rad PADD, and transmission mode images. To change a filter, remove the Filter Access Panel inside the camera chamber. See *Replacing or Changing an Emission Filter* in Chapter 6: *Maintaining the System*. This light-tight door gives you direct access to the Filter Wheel. Use the pins on the Filter Access Panel to remove the filter by locating the pins in the two holes provided in the filter cassette and withdraw the filter. Replace the filter by aligning the tracks on the filter cassette with the Filter Wheel and then inserting the cassette.



WARNING: Remove the Filter Access Panel only when it is necessary to change a filter.



- A The *Filter Wheel* (not shown) holds the filters for detection of different fluorophors. There are four spaces in the wheel. Position “0” is preferably open, without a cassette. Based on your applications, select the appropriate filters for the remaining three positions.
- B The *Filter Access Panel* provides access to the Filter Wheel and is light-tight.

The KODAK rad PADD (Optional Accessory)

The patented rad PADD (radioisotopic phosphor assisted detection device) is an optional accessory, which converts emissions from radioisotope samples into light that can be imaged on the IS4000R. An optical plate beneath the phosphor efficiently couples the screen output to the lens system. Autoradiography capture times are comparable to that of KODAK X-OMAT AR film with a KODAK X-OMAT intensifying screen for exposures less than 4 hours.

 **NOTE:** To decontaminate your screen, refer to *Radiological Decontamination Procedure* in Chapter 6: *Maintaining the System*.

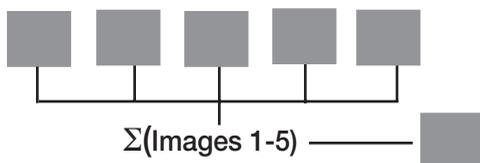


- A The *Cassette* provides a convenient isolated environment for your radioactive sample.
- B The *Phosphor Screen (cassette open)* is where you place your sample. If your object is opaque (blot), orient the sample side downward—if translucent/translucent (gel), orient the sample either way.
- C *IS4000R Platen*—position the rad PADD with the phosphor screen so that the underside (with the interference filter) is facing the Platen.
- D *rad PADD Cassette Screws* located on either end of the cassette may be removed to lift out the rad PADD Cassette Lid to accommodate thick samples such as microwell plates.

Software Overview

This section provides an overview of the KODAK Image Station 4000R (IS4000R) Image Acquire Software. This software is accessed from within KODAK Molecular Imaging Software (KODAK MI). The IS4000R supports a wide variety of image capture methods including single capture exposure, multiple capture exposure (accumulated into a single file or as separate files), time lapse exposures, progressive exposures, and rad PADD exposures. Using these exposure modes, the minimum and maximum exposure times are 0.05 second and 100 minutes, respectively. In addition, the rad PADD exposures allow for a single capture exposure or multiple exposures (final accumulated image).

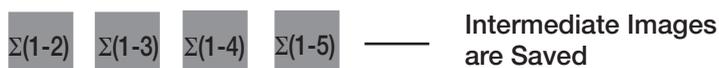
- ✓ *Standard Exposure—Final Accumulation (Single)* The IS4000R cooled CCD Camera with a 16-bit A/D converter provides a 16-bit data file. After capture, the image file automatically opens as a KODAK MI project.
- ✓ *Standard Exposure—Final Accumulation (Multi)* adds successive captures into a single accumulated n-bit image. The maximum number of images accumulated is 32. The final accumulated image is automatically opened as a KODAK MI project.



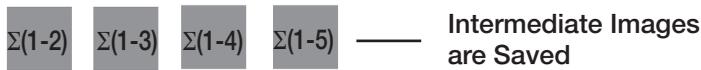
- ✓ *Standard Exposure—Separate Images* generates single captures in a multiple exposure as KODAK MI projects. The maximum number of exposures is 32. Each file is automatically saved to your hard drive.



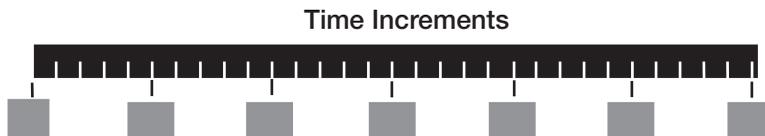
- ✓ *Standard Exposure—All Accumulations* saves an intermediate (incremental) series of captures as KODAK MI projects. The maximum number of exposures is 32. Each intermediate accumulation file is automatically saved to your hard drive.



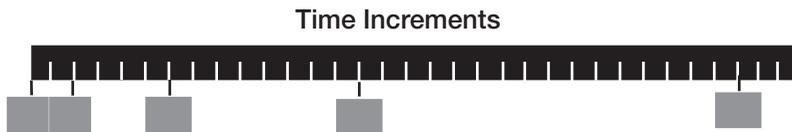
- ✓ *Standard Exposure—All Images* saves all intermediate and separate images. The maximum number of images that may be captured is 32 accumulated and 32 separate images. Each intermediate accumulation and separate file is automatically saved to your hard drive. The final accumulated image automatically opens as a KODAK MI project.



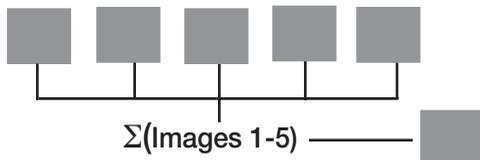
- ✓ *Time Lapse Exposure* provides exposures at set intervals. You can set the exposure time, the time lapse interval, the total exposure time, and the total number of images. The maximum duration time is 150 hours. The maximum number of images that may be captured is 32. Each file is automatically saved to your hard drive.



- ✓ *Progressive Exposures* is designed to take multiple exposures at increasingly different exposure times. This automates complex exposure options—thereby eliminating the need to manually set each exposure time. The maximum number of images that may be captured is 32. Each file is automatically saved to your hard drive.



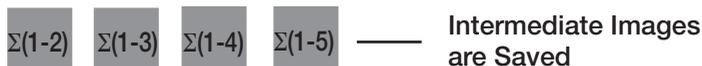
- ✓ *rad PADD—Final Accumulation (Single)* captures one optimized 16-bit image. After capture, the image file automatically opens as a KODAK MI project.
- ✓ *rad PADD—Final Accumulation (Multi)* adds successive captures into a single accumulated n-bit image. The maximum number of images accumulated is 32. The final accumulated image is automatically opened as a KODAK MI project.



- ✓ *rad PADD—Separate Images* generates single captures in a multiple exposure as KODAK MI projects. The maximum number of exposures is 32. Each file is automatically saved to your hard drive.

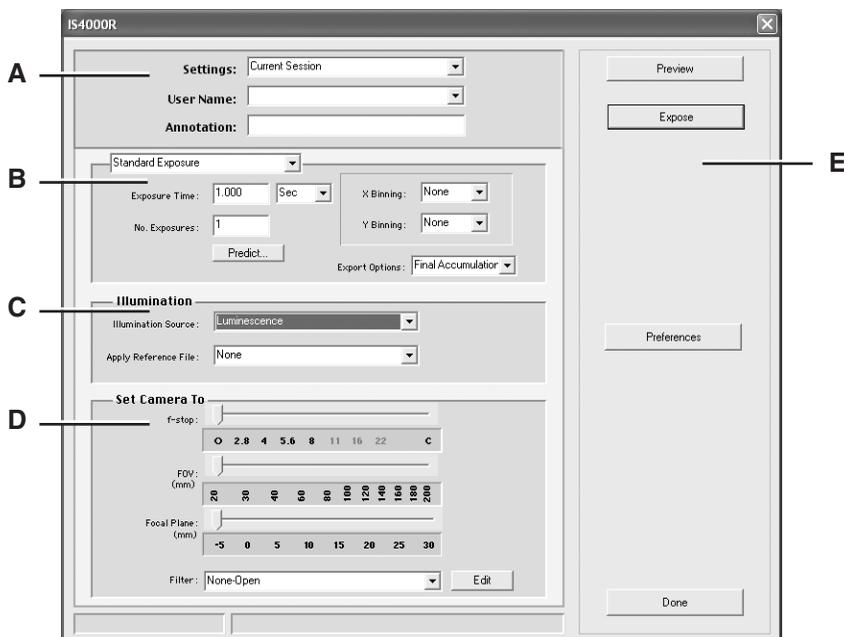


- ✓ *rad PADD—All Images* saves an intermediate (incremental) series of captures as KODAK MI projects. The maximum number of images accumulated is 32. Each intermediate accumulation file is automatically saved to your hard drive.



IS4000R Acquire Window

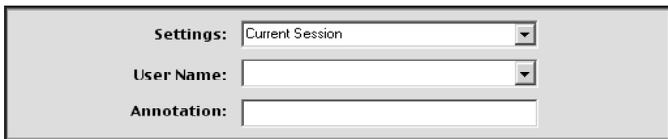
The IS4000R Acquire window is accessed from within KODAK MI Software. Choose Select Digital Camera from the KODAK MI File menu and IS4000R from the Digital Camera pop-up menu. After launching IS4000R, the IS4000R Acquire window appears. This window is divided into five sections. Let's review each section.



- A The *User* section contains options to select a predefined setting, record your name, and add annotations on the image for printing.
- B The *Exposure* section provides you with the various exposure methods and options associated with capturing images.
- C The *Illumination* section designates the type of illumination used to capture your image and applies the appropriate Illumination Reference File, when necessary.
- D The *Set Camera To* section allows you to record the position of the camera lens *f*-stop and field of view. In addition, it provides a pop-up menu for recording the filter used during capture. These parameters are important for documentation and calibration of your image.
- E The *Buttons* section contains the Preview, Expose, Preferences, and Done buttons used to navigate through an image capture.

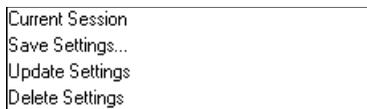
User Section of the IS4000R Acquire Window

The User section of the IS4000R Acquire window contains the Settings, User Name, and Annotation options. Let's review each option:



The screenshot shows a rectangular window with a light gray background. It contains three rows of controls. The first row is labeled 'Settings:' and has a dropdown menu with 'Current Session' selected. The second row is labeled 'User Name:' and has a dropdown menu. The third row is labeled 'Annotation:' and has a text input field.

Settings Option



The screenshot shows a vertical list of four menu items: 'Current Session', 'Save Settings...', 'Update Settings', and 'Delete Settings'.

You can speed up the capture process by creating custom capture settings for the various types of experiments or for specific personal preferences. Once saved, these settings can be conveniently applied to future image captures.

You can modify a saved settings (i.e., change exposure time, Illumination Reference File, etc.), update, edit, or delete a setting.

Once saved, settings remembered include:

- ✓ Exposure time
- ✓ Number of exposures
- ✓ Illumination settings
- ✓ *f*-Stop and field of view settings
- ✓ Camera filter (Filter location on Filter Wheel, but not filter name)
- ✓ Image orientation
- ✓ Preferences information
- ✓ Rotation
- ✓ Saturation levels
- ✓ Application of lens or Illumination Reference Files
- ✓ X and/or Y binning

In addition, only the filter location is retained in a user setting file. If you save a setting with one filter and later replace that filter and edit the name using the Filter Edit button, the new filter name appears in the Filter pop-up menu.

- ☞ NOTE: If your instrument is a shared resource, make sure that the appropriate filter name is in the Filter pop-up menu.
- ☞ NOTE: User defined settings are saved as individual files in the KODAK MI Extensions> IS4000R_Settings (Macintosh) or C:\KODAK MINS4000R\IS4000R_Settings (Windows) subfolders. Any settings files in this folder automatically appear in the Settings pop-up menu.

User Name Option

Use the User Name text edit box to type in your name. To save a new name for future experiments, choose Add User from the pop-up menu next to the User Name text edit box (down arrow). The name appears in the menu. Names can be deleted in the same manner by choosing Delete User.

Annotation Option

You can tag your image with an Annotation bar with up to 48 characters of text. In addition, the Annotation bar lists the time and date of the capture of the image, the exposure time, and the illumination source. The text appears on the long axis at the top of the printed annotations page.

All items in the Annotation bar are separate objects and are displayed in the Annotation window. This allows you to edit the text, change the font style, color, or size prior to printing. The contents are grouped, therefore, you must ungroup these objects prior to editing.

- ☞ NOTE: Each time you capture an image, the Annotation text edit box is cleared and can not be saved with the User Settings.
- ☞ NOTE: The Annotation bar is only available prior to image capture. Once you capture an image, you cannot add the Annotation bar.
- ☞ NOTE: Display or hide the Annotation bar by selecting the Show Annotation bar from within the Annotation panel. This is only available if one is present.

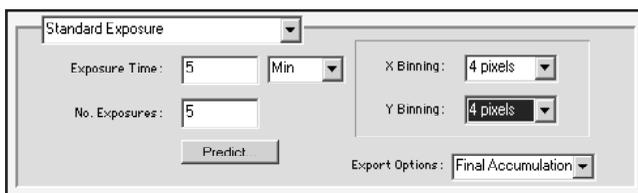
Exposure Section of the IS4000R Acquire Window

Use the IS4000R to take standard exposures (single and/or multiple exposure accumulations), time lapse exposures, progressive exposures, or rad PADD exposures. In addition, you can generate an Illumination Reference File. The pop-up menu at the top of this section sets the type of image captures to be performed.

To increase sensitivity, you may also choose to X and/or Y bin your image. Binning is the process by which adjacent CCD signal values are added together and then each pixel value is replaced with the new added value. For example, if two adjacent pixels have an intensity value of 100 and 150, when binning has been applied, both pixels are assigned an intensity value of 250. Binning can be applied on the X and/or Y axis of an image. While binning decreases the resolution of the image, it increases the intensity of the signal. Refer to Appendix A: *Digital Imaging Concepts* for a technical explanation of binning and other digital imaging concepts.

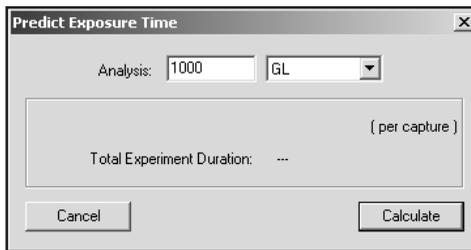
Standard Exposure

The Standard Exposure option lets you take a single image or capture multiple images. When you capture multiple images you can choose to save each capture as a separate file, accumulate all the captures in a single file, or generate a series of intermediate accumulations. To increase sensitivity you may also choose to X and/or Y bin your image.



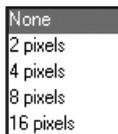
- ✓ The *Exposure Time* text edit box and *Units* pop-up menu defines the exposure. The minimum and maximum exposure times are 0.05 second per exposure and 100 minutes per exposure, respectively. The minimum exposure time for a binned image is 1 second. The time units can be set as seconds or minutes. The minimum increment is 0.1 second.
 - ✓ Use the *No. Exposures* text edit box to choose the total number of exposures. The maximum number of exposures is 32.
- 👉 NOTE: Generally multiple captures are not appropriate unless exposure times are greater than 20 minutes per capture or a single capture is close to saturation.

- ✓ The *Predict* button is ideal for estimating exposure times for luminescence and rad PADD images. Once selected, the Predict Exposure dialog box appears where you can enter the desired number of gray levels (GL). When the *Calculate* button is clicked, the camera takes a series of exposures to help predict the optimal exposure time. Once determined, the predicted exposure time and number of exposure are automatically updated in the IS4000R Acquire window for single capture determinations only. In a multiple frame exposure, the calculated times and number of exposures must be entered manually.



- The *Analysis* text edit box and GL pop-up menu allows you to enter the number of gray levels you want your exposure to achieve.
- The *Max Exposure Time* indicates when the image will become saturated. Increasing exposures beyond this time may adversely affect your analysis data.
- The *Total Time* equals the total time required to achieve the desired number of gray levels.

- ✓ To increase sensitivity with luminescence or rad PADD imaging, you may also choose to X and/or Y bin your image. *X Binning* and *Y Binning* options allow you to select from None (no binning), 2 pixel, 4 pixel, 8 pixel or 16 pixel binning in one or both the X and Y directions.



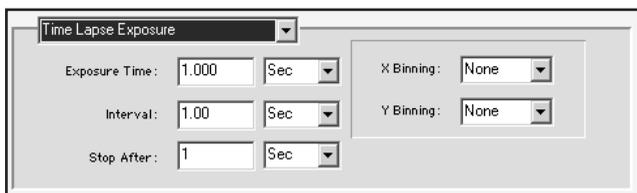
Binning is asymmetric in one direction. X-direction/horizontal on the Platen supports the long axis for asymmetric binning. Bin options are none, 1 x 2, 1 x 4, 1 x 8, 2 x 2, 2 x 4, 2 x 8, 4 x 4, 4 x 8, 8 x 8, and 16 x 16.

- ✓ *Export Options* defines the final image—choose from Final Accumulation, All Accumulations, All Images, or Separate Images.

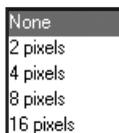


Time Lapse Exposure

The Time Lapse Exposure option is designed to take multiple exposures over fixed time intervals. For example, you may take a 10 second exposure every hour over the next 10 hours.



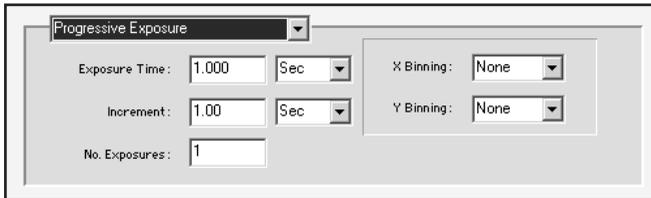
- ✓ The *Exposure Time* text edit box and *Units* pop-up menu defines the exposure. The minimum and maximum exposure times are 0.05 second per frame and 100 minutes per frame, respectively. The time units can be set as seconds or minutes. The maximum duration time is 150 hours. The minimal increment is 0.1 second.
 - ✓ The *Interval* is the time interval between the beginning of one exposure and the beginning of the next exposure. The minimum and maximum intervals are 1 second and 24 hours, respectively.
- ☞ NOTE: An interval must be at least 1 second greater than an exposure time.
- ✓ *Stop After* determines when to stop time lapse captures. You can set stop criteria based on experimental duration or by the number of frames.
 - ✓ To increase sensitivity with luminescence or rad PADD imaging, you may also choose to X and/or Y bin your image. *X Binning* and *Y Binning* options allow you to select from None (no binning), 2 pixel, 4 pixel, 8 pixel or 16 pixel binning in one or both the X and Y directions.



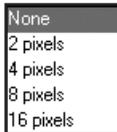
Binning is asymmetric in one direction. X-direction/horizontal on the Platen supports the long axis for asymmetric binning. Bin options are none, 1 x 2, 1 x 4, 1 x 8, 2 x 2, 2 x 4, 2 x 8, 4 x 4, 4 x 8, 8 x 8, and 16 x 16.

Progressive Exposure

The Progressive Exposure option is designed to take a continuous sequence of exposures at different exposure times. You can automate complex exposure options—rather than manually having to set each exposure time. For example, you may want to capture an image at each of the following exposure times: 10, 20, 40, and 80 seconds.



- ✓ The *Exposure Time* text edit box and *Units* pop-up menu defines the exposure. The minimum and maximum exposure times are 0.05 second per frame and 100 minutes per frame, respectively. The time units can be set as seconds or minutes.
- ✓ The *Increment* determines each new exposure time. The smallest increment for exposures is 1 second. For example, if you set 60 second increments—each subsequent exposure will be 60 seconds longer than the previous exposure. Another example would be to set an increment of 2X (Multiplier #X): If the first exposure was 1 minute, each successive exposure is double the previous exposure (2, 4, 8, 16 minutes, etc.). In addition, you can input the geometric order for generating exposure times. In this case the number entered will be the power to which successive exposure times will be raised.
- ✓ Use the *No. Exposures* text edit box to choose the total number of exposures. The maximum number of exposures is 32.
- ✓ To increase sensitivity with luminescence or rad PADD imaging, you may also choose to X and/or Y bin your image. *X Binning* and *Y Binning* options allow you to select from None (no binning), 2 pixel, 4 pixel, 8 pixel or 16 pixel binning in one or both the X and Y directions.

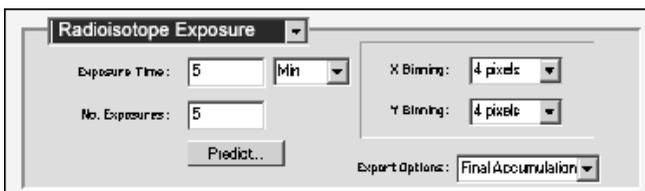


Binning is asymmetric in one direction. X direction/horizontal on the Platen supports the long axis for asymmetric binning. Bin options are none, 1 x 2, 1 x 4, 1 x 8, 2 x 2, 2 x 4, 2 x 8, 4 x 4, 4 x 8, 8 x 8, and 16 x 16.

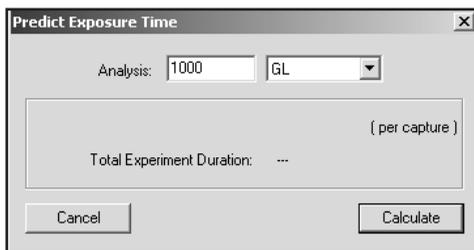
rad PADD

The rad PADD is designed to image samples that have equivalent signal to a 4 hour or less film exposure.

You can take a single image or capture multiple images. When you capture multiple images you can choose to save each capture as a separate file, accumulate all the captures in a single file, or generate a series of intermediate accumulations. To increase sensitivity you may also choose to X and/or Y bin your image.



- ✓ The *Exposure Time* text edit box and *Units* pop-up menu defines the exposure. The minimum and maximum exposure times are 0.05 second per exposure and 100 minutes per exposure, respectively. The time units can be set as seconds or minutes. The minimum increment is 0.1 second.
- ✓ Use the *No. Exposures* text edit box to choose the total number of exposures. The maximum number of exposures is 32.
- ✓ The *Predict* button opens a dialog box where you can calculate a prediction of the optimal exposure time based upon the desired number of gray levels (GL). When the *Calculate* button is clicked, the camera takes a series of exposures to help predict the optimal exposure time. Once determined, the predicted exposure time and number of exposure are automatically updated in the IS4000R Acquire window for single capture determinations only. In a multiple frame exposure, the calculated times and number of exposures must be entered manually.



- The *Analysis* text edit box and GL pop-up menu allows you to enter the number of gray levels you want your exposure to achieve.
- The *Max Exposure Time* indicates when the image will become saturated. Increasing exposures beyond this time may adversely affect your analysis data.
- The *Total Time* equals the total time required to achieve the desired number of gray levels.

- ✓ To increase sensitivity with luminescence or rad PADD imaging, you may also choose to X and/or Y bin your image. *X Binning* and *Y Binning* options allow you to select from None (no binning), 2 pixel, 4 pixel, 8 pixel or 16 pixel binning in one or both the X and Y directions.



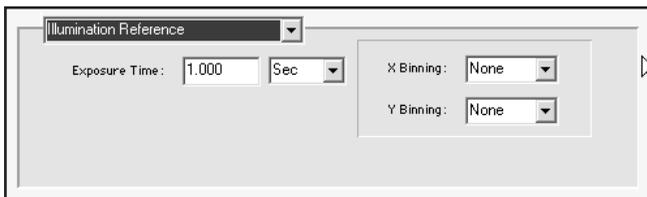
Binning is asymmetric in one direction. X direction/horizontal on the Platen supports the long axis for asymmetric binning. Bin options are none, 1 x 2, 1 x 4, 1 x 8, 2 x 2, 2 x 4, 2 x 8, 4 x 4, 4 x 8, 8 x 8, and 16 x 16.

- ✓ *Export Options* defines the final image—choose from Final Accumulation, All Accumulations, All Images, or Separate Images.



Illumination Reference

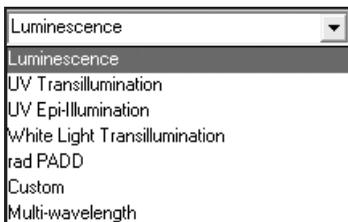
When a light source (UV or ambient light) is used, an illumination reference can be used to improve the quality of your data. The nonuniformity in the illumination generated by the UV bulbs or ambient white light is highly reproducible and may be corrected by simply normalizing the image by an illumination reference image. The f -stop and field of view settings used to capture the image must be the same as those used to generate the Illumination Reference File. See *Generating an Illumination Reference File Library* in Chapter 5: *Capturing Images*.



- ✓ The *Exposure Time* defines the exposure. The minimum and maximum exposure times are 0.05 second per exposure and 100 minutes per exposure, respectively. The time units can be set as seconds or minutes. The minimum increment is 0.1 second.

Illumination Section of the IS4000R Acquire Window

The Illumination section of the IS4000R Acquire window contains the options to select the Illumination Source and Illumination Reference File. The IS4000R captures images for many different applications including:



- ✓ *Luminescence*—a luminescent sample needs no additional illumination for capture—it emits light. The amount of light emitted by the sample is directly proportional to the limiting chemical component of the luminescent reaction, usually an enzyme-tagged molecular target. The CCD camera captures the emitted light and represents the signal in the image with very high fidelity (linear response at high precision).
- ✓ *UV Transillumination*—a fluorescent sample requires illumination to excite fluorescent molecules (called fluorochromes or fluorophores), which emit light. Ultraviolet transillumination is provided by the optional Image Station Transilluminator Lid Accessory. The spectrum of UV illumination has a peak emission at 306 nm. The UV light excites the sample and the camera detects the signal.

Fluorophores typically emit light in a manner that is proportional to their amount, and the CCD camera captures the fluorescent light and represents the signal in the image with very high fidelity (linear response at high precision).

- ✓ *UV Epi-illumination*—a fluorescent sample requires illumination to excite fluorescent molecules (called fluorochromes or fluorophores), which emit light. Internal ultraviolet illumination is provided by the IS4000R. The spectrum of UV illumination is between 300–400 nm. The UV illumination is designed to irradiate the sample at a 45° angle (called epi-illumination), and requires that the light interact with the sample in order to eventually enter the lens. Further, the CCD is relatively insensitive to the UV, and it is covered with a filter that blocks infrared radiation that emanates from any light source.

Fluorophores typically emit light in a manner that is proportional to their amount, and the CCD camera captures the fluorescent light and represents the signal in the image with very high fidelity (linear response at high precision).

UV/Fluorescence Quenching—detecting absorbance by reflected light, for samples such as chromogenic blots, may be accomplished using the UV epi-illumination provided by the IS4000R. In the case of a clear sample, the white backing of the Compression Pad may be used to assure light return. This method is an example of fluorescence quenching, which is actually an absorbance methodology.

- ✓ *White Light Transillumination*—a sample that absorbs light needs illumination to distinguish the absorbing molecules (called chromogens or chromophores). The CCD captures the light that is not absorbed in a highly linear manner. Generally, chromogens that absorb light do so in a manner that is proportional to the negative logarithm of the chromogen concentration. In terms of classical densitometry, this functional dependence is called absorbance or optical density (OD), and is related to the chromogen concentration by an extinction coefficient.

Absorbance can be measured by capturing the light that is transmitted through a sample. Transmission of light through a sample can be accomplished using the IS4000R, simply by opening the lid and permitting external light to be transmitted through a sample or using the optional Image Station Transilluminator Lid Accessory with the White Light PADD.

- ✓ *rad PADD*—If you purchased your IS4000R with the rad PADD accessory, you are able to capture high energy isotopes (^{32}P , ^{90}Y , ^{111}I) samples. Radioisotopes emit beta particles that interact with the phosphor screen in the rad PADD and convert energy to light. Capture time will be comparable to that of KODAK BioMax MS film with a KODAK HE intensifying screen, the fastest film/screen combination available for autoradiography.
- ✓ *Custom*—provides a custom setting option.

Applying an Illumination Reference File

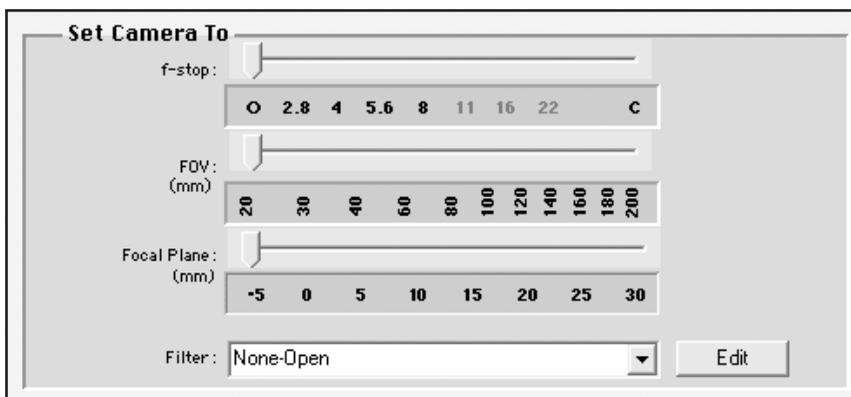
Improve the quality of your data by applying an illumination correction to images captured in the UV or white light imaging modes. The illumination non-uniformity is highly reproducible and may be corrected by normalizing the sample image to an illumination reference image.

The Apply Reference pop-up menu allows you to select a reference file. A reference file must have the same capture setting (*f*-stop, field of view) as the image. To create an Illumination Reference File, see *Generating an Illumination Reference File Library* in Chapter 5: *Capturing Images*.

There is also an automatic option that allows you to capture your image and automatically correct for illumination. When this option is selected, you capture your image and the software guides you through a second image capture as a reference file, see *Applying an Automatic Illumination Reference File* in Chapter 5: *Capturing Images*.

The Set Camera To Section of the IS4000R Acquire Window

The Set Camera To section of the IS4000R Acquire window allows you to enter information about the capture settings used. Documenting this information is critical, since KODAK MI uses this data to perform lens correction and illumination field flattening to optimize the image for display and analysis. In addition, these settings are recorded as part of the file history and can be used as a reference for similar type experiments.



- ✓ The *f-Stop* slider should match the aperture corresponding to the *f-stop* setting on your IS4000R camera. We do not recommend using *f-stops* over 11 for most applications.
 - ✓ The *FOV (mm)* slider is used to record the field of view to the corresponding settings on your IS4000R camera. For example, when you select 150 on the slider, your 150 x 150 mm image will be labelled as such.
 - ✓ The *Focal Plane (mm)* slider should match the current focus setting. The default value is 0 (focus to the Platen without interfering filters).
 - ✓ The *Filter* pop-up menu should match the filter selected on your IS4000R. You can also access the Available Filters dialog box by clicking the *Edit* button which allows you to edit the positions of the filters to correspond with the filter positions in the IS4000R Filter Wheel. The factory default is Position 0: None—Open. Position 1 is 600 nm WB Filter (Ethidium Bromide).
- 👉 NOTE: The Set Camera To sliders do not adjust the lens or filters, they only provide you a convenient place to record current settings.

Buttons Section of the IS4000R Acquire Windows



The Buttons section of the IS4000R Acquire window is used to navigate through an image capture.

- ✓ The *Preview* button previews the CCD image. The Preview window is described in more detail in the next section.
- ✓ The *Expose* button begins the image capture, opening a Capture Status window. The various Capture Status windows are described in more detail later in this chapter.
- ✓ The *Preferences* button opens the IS4000R Preferences window where you can customize certain capture parameters. The IS4000R Preferences window is described in more detail later in this chapter.
- ✓ The *Done* button closes the IS4000R Acquire window and returns you to KODAK MI.

Preview Window

The Preview button opens an IS4000R Preview window. The preview is used primarily for sample placement and is limited to a single frame capture. However, these frames are displayed in a continuous/successive sequence to allow for sample adjustment in a “live” mode. Set a shorter time to get a faster preview update.



- ✓ The *Exposure Time* text field shows current preview exposure time.
- ✓ The *Exposure Time* slider sets the exposure time for the preview image. The maximum exposure time is 5 seconds.
- ☞ NOTE: The Preview Exposure Time is remembered between sessions and is independent of the current exposure time in the IS4000R Acquire window.
- ✓ The *Exposure Time* progress bar gives you feedback on the current status of the preview image.
- ✓ The *Done* button stops the preview and returns you to the IS4000R Acquire window.
- ☞ NOTE: The Preview Exposure Time may not be an exact predictor of time for a full exposure. Use Predict for a more accurate exposure time estimate.
- ✓ The *Focus* check box accesses the software-assisted focusing aid.

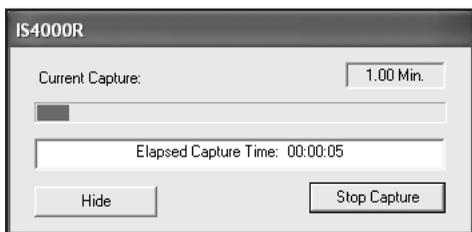
Capture Status Windows

Once an exposure is initiated a Capture Status window appears. There are two capture status windows—the Single Capture Status window (Standard) and the Multiple Capture Status window (Standard, Progressive, Time Lapse, rad PADD). Let's review.

Standard Exposure Capture Status Window

The Expose button starts the acquisition process. Immediately after the Expose button is selected, one of the capture status windows appears (depends on the exposure type).

For single frame captures, the following window appears:



- ✓ The *Current Capture* text field indicates the exposure time that was set and underway.
- ✓ The *Current Capture* progress bar displays the status of the capture.
- ✓ The *Elapsed Time* text field indicates the elapsed time since the beginning of the exposure.
- ✓ The *Hide* button closes the window without stopping the exposure. You can access the window by selecting IS4000R Capture from the KODAK MI File menu or the IS4000R Capture button on the Status bar.
- ✓ The *Stop Capture* button interrupts the capture and returns you to the IS4000R Acquire window. Stopping a single image capture will not save the current image. No image is opened or saved.

Standard Exposure (Multiple Exposure) Capture Status Window

The Expose button starts the acquisition process. If the acquisition you have selected is a standard exposure with multiple image captures, the following window appears:

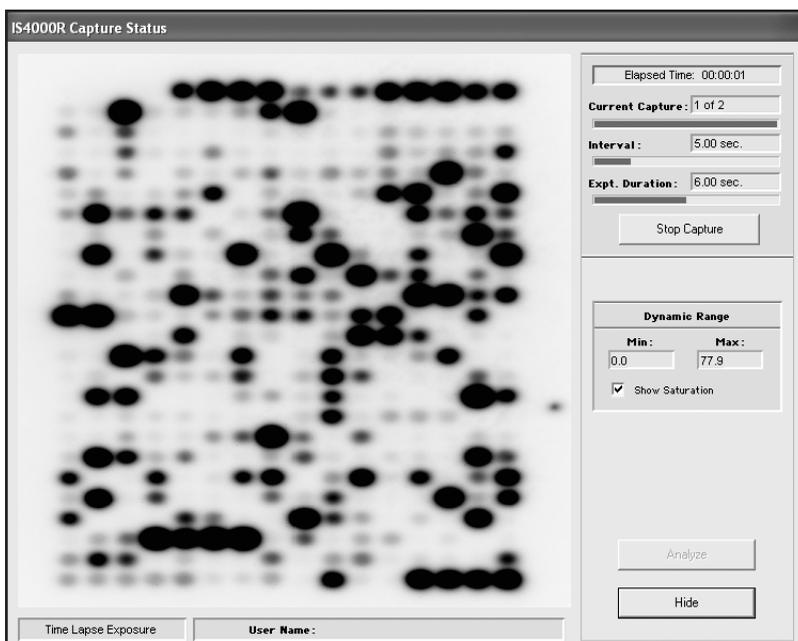


- ✓ The *Elapsed Time* text field shows the elapsed time since the beginning of the exposure.
- ✓ The *Current Capture* text field indicates the current exposure being captured.
- ✓ The *Current Capture* progress bar displays the status of the current capture.
- ✓ The *Expt Duration* text field displays the total time from the beginning of the first frame to the end of the last frame—regardless of the exposure type.
- ✓ The *Expt Duration* progress bar shows the status of the entire experiment (including all captures).
- ✓ The *Stop Capture* button interrupts the current capture and returns you to the IS4000R Acquire window. Any images saved to the hard drive prior to clicking this button are preserved. All the appropriate image filters/corrections are applied. If selected, a final accumulated image is opened in the Project window.

-
-
- ✓ *Dynamic Range* provides histogram information about the image being captured. *Min* and *Max* text fields display the current minimum and maximum signal values within the image. The *Show Saturation* check box shows any saturated pixels in the image in red. The *Auto Stop at* check box sets a dynamic range at which the capture process is interrupted. For example, you can set the auto stop value at 10,000 in the *Auto Stop at* text edit box. Whenever an accumulation is added and the difference between the maximum and minimum gray level values is greater than 10,000 then the capture process stops and the image is transferred to KODAK MI Software. The default value is 65,535 signal intensities.
 - ✓ The *Analyze* button transfers the last completed image or accumulation captured to KODAK Molecular Imaging Software for analysis. You can choose this option anytime during the capture process to open intermediate images. Selecting this option will not stop the capture in progress.
 - ✓ The *Hide* button closes the window without stopping the exposure. You can access the window by selecting IS4000R Capture from the KODAK MI File menu or by using the IS4000R Capture button on the Status bar.
 - ✓ The *Information* bar directly below the image contains the current exposure type and user name.

Time Lapse Exposure Capture Status Window

The Expose button starts the acquisition process. If the acquisition you have selected is a time lapse exposure, the following window appears:



- ✓ The *Elapsed Time* text field shows the elapsed time since the beginning of the exposure.
- ✓ The *Current Capture* text field indicates the number of captures in the time lapse series.
- ✓ The *Current Capture* progress bar displays the status of the capture.
- ✓ The *Interval* text field shows the elapsed time within the current time interval. The interval corresponds to the time between the beginning of one capture and the beginning of the next capture.
- ✓ The *Interval* progress bar displays the status of the current time interval.
- ✓ The *Expt Duration* text field displays total time from the beginning of the first frame to the end of the last frame—regardless of the exposure type.
- ✓ The *Expt Duration* progress bar shows the status of the entire experiment, including all time lapse exposures.

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- ✓ The *Stop Capture* button interrupts the current capture and returns you to the IS4000R Acquire window. Any images saved to the hard drive prior to clicking this button are preserved. All the appropriate image filters/corrections are applied, if selected a final accumulated image is opened.
 - ✓ *Dynamic Range* provides histogram information about the image being captured. *Min* and *Max* text fields display the current minimum and maximum signal values within the image. The *Show Saturation* check box shows any saturated pixels in the image in red.
 - ✓ The *Analyze* button transfers the last completed image or accumulation captured to KODAK MI for analysis. You can choose this option anytime during the capture process to open intermediate images. Selecting this option will not stop the capture in progress.
 - ✓ The *Hide* button closes the window without stopping the exposure. You can access the window by selecting IS4000R Capture from the File menu or by using the IS4000R Capture button on the Status bar.
 - ✓ The *Information* bar directly below the image contains the current exposure type and user name.

Progressive Exposure Capture Status Window

The Expose button starts the acquisition process. If the acquisition you have selected is a progressive exposure, the following window appears.

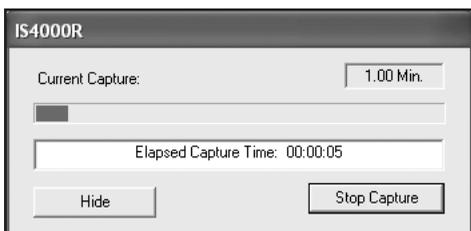


- ✓ The *Elapsed Time* text field shows the elapsed time since the beginning of the exposure.
- ✓ The *Current Capture* text field indicates the number of the current exposure in the progressive series being captured.
- ✓ The *Current Capture* progress bar displays the status of the capture.
- ✓ The *Expt Duration* text field displays total time from the beginning of the first frame to the end of the last frame—regardless of the exposure type.
- ✓ The *Expt Duration* progress bar shows the status of the entire experiment including all progressive exposures.
- ✓ The *Stop Capture* button interrupts the current capture and returns you to the KODAK MI main window. Any images saved to the hard drive prior to clicking this button are preserved. All the appropriate image filters/corrections will be applied. For example, you can stop the progressive capture at any current exposure without having to wait for the entire number of captures to be completed.

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- ✓ *Dynamic Range* provides histogram information about the image being captured. *Min* and *Max* text fields display the current minimum and maximum signal values within the image. The *Show Saturation* check box shows any saturated pixels in the image in red. The *Auto Stop at* check box sets a dynamic range at which the capture process is interrupted. For example, you can set the auto stop value at 10,000 in the *Auto Stop at* text edit box. Whenever an accumulation is added and the difference between the maximum and minimum gray level values is greater than 10,000 then the capture process stops and the image is transferred. The default value is 65,535 signal intensities.
 - ✓ The *Analyze* button transfers the last completed image or accumulation captured to for analysis. You can choose this option anytime during the capture process to open intermediate images. Selecting this option will not stop the capture in progress.
 - ✓ The *Hide* button closes the window without stopping the exposure. You can access the window by selecting IS4000R from the File menu or by using the IS4000R Capture button on the Status bar.
 - ✓ The *Information* bar directly below the image contains the current exposure type and user name.

rad PADD—Single Exposure Capture Status Window

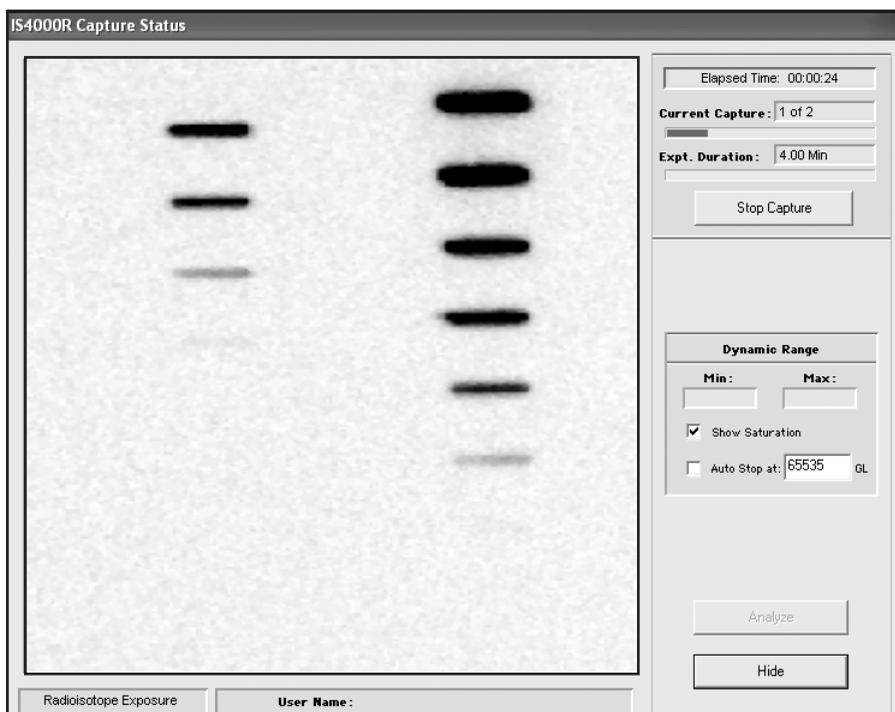
For single frame captures, the following window appears:



- ✓ The *Current Capture* text field indicates the exposure time that was set and underway.
- ✓ The *Current Capture* progress bar displays the status of the capture.
- ✓ The *Elapsed Time* text field indicates the elapsed time since the beginning of the exposure.
- ✓ The *Hide* button closes the window without stopping the exposure. You can access the window by selecting IS4000R from the File menu or by using the IS4000R Capture button on the Status bar.
- ✓ The *Stop Capture* button interrupts the capture and returns you to the IS4000R Acquire window. No image is opened or saved after stopping a single image capture.

rad PADD—Multiple Exposures Capture Status Window

The Expose button starts the acquisition process. If the acquisition you have selected is a rad PADD with multiple image captures, the following window appears:

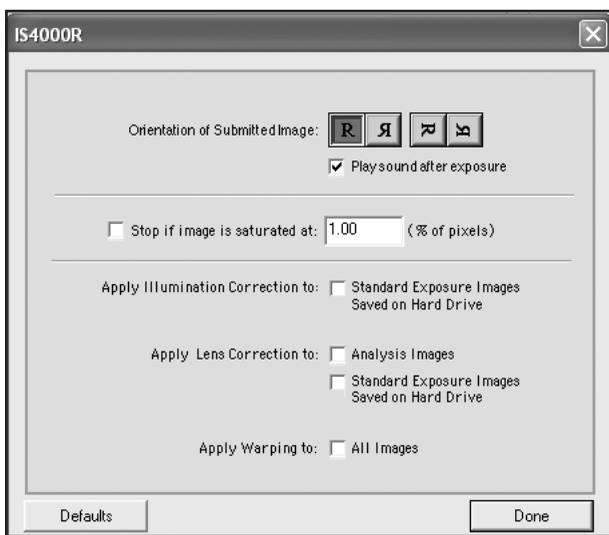


- ✓ The *Elapsed Time* text field shows the elapsed time since the beginning of the exposure.
- ✓ The *Current Capture* text field indicates the current exposure being captured.
- ✓ The *Current Capture* progress bar displays the status of the capture.
- ✓ The *Expt Duration* text field displays the total time from the beginning of the first frame to the end of the last frame—regardless of the exposure type.
- ✓ The *Expt Duration* progress bar shows the status of the entire rad PADD exposure.
- ✓ The *Stop Capture* button interrupts the current capture and returns you to the KODAK MI Main window. Any images saved to the hard drive prior to clicking this button will remain and not be deleted.

-
-
- ✓ *Dynamic Range* provides histogram information about the image being captured. *Min* and *Max* text fields display the current minimum and maximum signal values within the image. The *Show Saturation* check box shows any saturated pixels in the image in red. The *Auto Stop at* check box sets a dynamic range at which the capture process is interrupted. For example, you can set the auto stop value at 10,000 in the *Auto Stop at* text edit box. Whenever an accumulation is added and the difference between the maximum and minimum gray level values is greater than 10,000 then the capture process stops and the image is transferred. The default value is 65,535 signal intensities.
 - ✓ The *Analyze* button transfers the current image frame in the window for analysis.
 - ✓ The *Hide* button closes the window without stopping the exposure. You can access the window by selecting IS4000R from the File menu or by using the IS4000R Capture button on the Status bar.
 - ✓ The *Information* bar directly below the image contains the current exposure type and user name.

Preferences Window

The IS4000R Preferences window sets camera and software parameters including the image saturation level, orientation, and maximum exposure time. To access the IS4000R Preferences window, select the Preferences button from the IS4000R Acquire window.



✓ *Orientation of Submitted Image* option automatically rotates a newly captured image to a desired orientation while it is being downloaded.

-  The image is in the default orientation.
-  The image is flipped horizontally.
-  Rotates the image 90° clockwise.
-  Rotates the image 90° clockwise and flipped vertically.

 NOTE: The Rotation buttons do not change the orientation of the preview image. The rotation occurs during image submission.

 NOTE: Check the *Play sound after exposure* check box to be audibly notified of the completion of each frame capture.

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- ✓ The *Stop if image is saturated at* check box stops the image capture when the last completed capture becomes saturated to the extent noted in Preferences. For example, if 1% is designated, the capture will stop when 1% of the image is saturated. This setting applies to multiple captures only. Use the *Stop if image is saturated at* text edit field to select the percentage of pixels that can become saturated in the image. The image pixels turn red when the saturation level is reached if the *Show Saturation* check box is selected in the Capture Status window. The saturation levels for single and multiple frame capture can be set independently.
 - ✓ Use the *Apply Illumination Correction to* check box to apply illumination correction or select an Illumination Reference File in the IS4000R Acquire window to apply to standard exposure images saved to your hard drive. In Standard Exposure mode you can save all exposures, all intermediate accumulations, and the final accumulation. For the separate exposures and the intermediate accumulations, you may or may not want to apply illumination correction. For example, if you are interested in doing subsequent calculations, the application of illumination correction introduces errors.
 - ✓ The *Apply Lens Corrections to* option provides you with two options:
 - Analysis Images* check box to applies lens correction to a single frame capture, the final accumulation of multiple frame captures, progressive exposure captures, time lapse captures, and images submitted to KODAK MI from the Capture Status window which are opened as KODAK MI projects.
 - Intermediate Images Saved on Hard Drive* applies ONLY to the standard exposures. For separate exposures and intermediate accumulations (which are saved on the hard drive), you can select whether lens correction should be applied.
 - ✓ *Apply Warping to All Images* check box turns image warping on or off. Image warping is especially useful for correcting for lens distortion at low magnifications—i.e., 96 well plates, macroarrays.

To save the updated preferences, click Done. To reset the preferences to the factory default settings, click Defaults.

Setting Up for Capture

Now you will step through the KODAK Image Station 4000R (IS4000R) set up procedures for image capture including preparing the sample, setting illumination, and previewing the image.

Sample Illumination/Detection Guidelines

The IS4000R captures images for many different applications. When performing experiments, it is important that the appropriate capture conditions are used. The capture parameters relating to four major modes of detection are described below. Later in this chapter, you will find step-by-step procedures for preparing for image capture, setting illumination, and previewing your image.

Luminescence Imaging

A luminescent sample needs no additional illumination for capture—it emits light. The amount of light emitted by the sample is directly proportional to the limiting chemical component of the luminescent reaction, usually an enzyme-tagged molecular target. The CCD camera captures the emitted light and represents the signal in the image with very high fidelity (linear response at high precision).

Luminescent samples can vary greatly in signal intensity. Many luminescent signals are dim and may not fill all the signal levels in the Image Histogram in a reasonable period of time. Therefore, luminescent signals may require binning in both the X and Y directions. Minimally, greater than 500 gray levels is desired, particularly if multiple objects are to be measured. In general, the total exposure time for luminescent captures on the IS4000R will be close to that of autoradiography film. For higher signal levels and a wider dynamic range, increase the exposure time. If no significant signal is visible, select an exposure time of up to 20 minutes with a single capture. If no signal is apparent after 20 minutes of exposure, the IS4000R system is not detecting the signal. Verify that your sample is generating signal by exposing with film.

Filters can be used to discriminate and differentially display different kinds of luminescence. However, for maximum sensitivity (usually a priority in luminescence experiments), do not use a filter, open the camera aperture ($f/0$), choose the highest binning state, and an extended exposure time (multiple minutes).

UV Epi-Illumination and Transillumination

Internal ultraviolet epi-illumination is provided by the IS4000R. The spectrum of UV epi-illumination is between 300 and 400 nm; for details refer to Appendix B: *UV Illumination* of this User's Guide. A fluorescent sample requires illumination to excite fluorescent molecules (called fluorochromes or fluorophores), which emit light. The Image Station 4000R UV epi-illumination is designed to irradiate the sample at a 45° angle, and requires that the light interact with the sample in order to eventually enter the lens. The CCD is relatively insensitive to the UV, and any IR from the light source will be filtered by the UV emission filters.

Also available as an option, the KODAK Image Station Transilluminator Lid Accessory provides UV ideal for imaging fluorescent samples like ethidium bromide stained gels. The spectrum of UV illumination has a peak emission at 306 nm. The UV light excites the sample and the camera detects the signal.

Fluorochromes typically emit light in a manner that is proportional to their amount, and the CCD camera captures the fluorescent light and represents the signal in the image with very high fidelity (linear response at high precision).

Fluorescence detection usually generates enough signal within a reasonable period of time to preclude the need for binning. However, binning may be convenient if consistent with the spatial resolution of the sample. Asymmetric binning (in one direction) may enhance signal without significant loss of resolution.

For documentation purposes, achieving 1000 signal levels of a target feature is sufficient. For high quality quantitation, greater than 4,000 signal levels is recommended. In situations where high background fluorescence consumes much of the available gray scale, accumulating images to greater than 20,000 signal levels may be necessary for good quantitation.

Autofluorescence from surrounding materials often presents an interfering background in a fluorescent detection experiment. To diminish the autofluorescent background, a series of filters are available that can be inserted into the IS4000R Filter Wheel. Selecting a filter may optimize the fluorescent signal by discriminating against background and/or differentiating between multiple fluorochromes within the experimental sample.

White Light Transillumination

A sample that absorbs light needs illumination to distinguish the absorbing molecules (called chromogens or chromophores). The CCD captures the light that is not absorbed in a highly linear manner. Generally, chromogens that absorb light do so in a manner that is proportional to the negative logarithm of the amount of chromogen. In terms of classical densitometry, this functional dependence is called absorbance or optical density (OD), and is related to the chromogen concentration by an extinction coefficient.

Absorbance can be measured by capturing the light that is transmitted through a sample. Transmission of light through a sample can be accomplished using the IS4000R, simply by opening the lid and permitting external light to be transmitted through a sample. To minimize imaging artifacts, a translucent panel (Light Diffuser) is placed above the sample on the Platen, optimally scattering the incoming light. Because of the high sensitivity of the camera, dim ambient light usually suffices.

Similarly, you can use the optional Image Station Transilluminator Lid Accessory to generate white light. The White Light PADD (Phosphor Assisted Detection Device) can be placed over the transilluminator to convert UV transillumination to white light for imaging Coomassie gels or for densitometry of films.

Alternatively, detecting absorbance by reflected light, for samples such as chromogenic blots, may be accomplished using the UV epi-illumination provided by the IS4000R. In the case of a clear sample, the white backing of the Compression Pad may be used to assure light return. Generally, better absorbance data may be generated using filtered epi-illumination.

Using transillumination, ample signal is provided and short exposure times are expected. Binning is never necessary and exposure times per capture will be approximately 0.1 second or less. Increasing the f -stop up to $f/8$ extends exposure times. Care should be taken not to saturate the image (pixels turn red). Saturation precludes the appropriate calculation of optical density for absorbance measurements. Utilize the accumulation mode to approach 60,000 signal levels to maximize the range of the optical density.

For improved sample quantitation, it is recommended that filters be used for detection using absorbance methods. For example, a blue dye presents maximum contrast (signal difference) if a red or yellow filter is used.

rad PADD Imaging Guidelines

The rad PADD utilizes patented technology to convert high-energy beta particles (^{32}P , ^{90}Y , ^{111}In , ^{125}I) into light (photons) that are collected by the IS4000R lens. The rad PADD cassette provides containment of radioactively labelled samples, reducing cross contamination of sample which can cause imaging artifacts.

When using the rad PADD with the IS4000R for exposure times of 4 hours or less, you will detect signals equivalent to KODAK X-OMAT AR film and KODAK X-OMAT Screens.

Like luminescent samples, radiolabeled samples can vary greatly in signal intensity. Low signals may generate images that do not fill all the signal levels in the Image Histogram in a reasonable period of time. Therefore, low signal levels may require binning in both the X and Y directions. Generally, greater than 300 gray levels is desired, particularly if multiple objects are to be measured. The total exposure time for radiolabeled samples on the IS4000R is typically close to that of autoradiography film for short exposures (< 2 hours) For higher signal levels and wider dynamic range, increase the exposure time.

Preparing the IS4000R for Imaging

Your IS4000R should be installed, see Chapter 8: *Installing the System*. The camera should be turned on and cooled prior to capturing images. The cooling of the camera is critical for low-level signal detection. The software warns you if you try to capture an image before the camera has sufficiently cooled (typically 20 minutes is required to stabilize temperature).

Be sure that the Platen surface is clean prior to any sample placement. Dust, fingerprints, or scratches on the Platen may introduce artifacts in your data. Extra care should be taken when cleaning the Platen to avoid scratches.

- 1 Open the Capture Chamber door.
- 2 Turn the Camera Power Switch on. An indicator light is displayed when the camera is operational. For maximum sensitivity allow the camera to cool down for approximately 30 minutes before beginning your imaging.

 NOTE: We recommend that you keep the camera powered on continuously. Turning the camera on and off may cause condensation in the optics. Power consumption is minimal.

- 3 Open the IS4000R lid.
- 4 Use an ammonia based spray cleaner. Wipe the Platen and the Surface Work Area with a lint-free soft cloth or damp paper towel.

 NOTE: Special care should be taken if your sample is dry. Assure a dry surface by a quick ethanol wipe.



WARNING: Ammonia based spray cleaners may cause eye irritation. Consult the manufacturer's material safety data sheet for additional information prior to use.



WARNING: Do not use metal utensils or tweezers since they may scratch and permanently damage the Platen. This type of damage is not covered by the warranty.



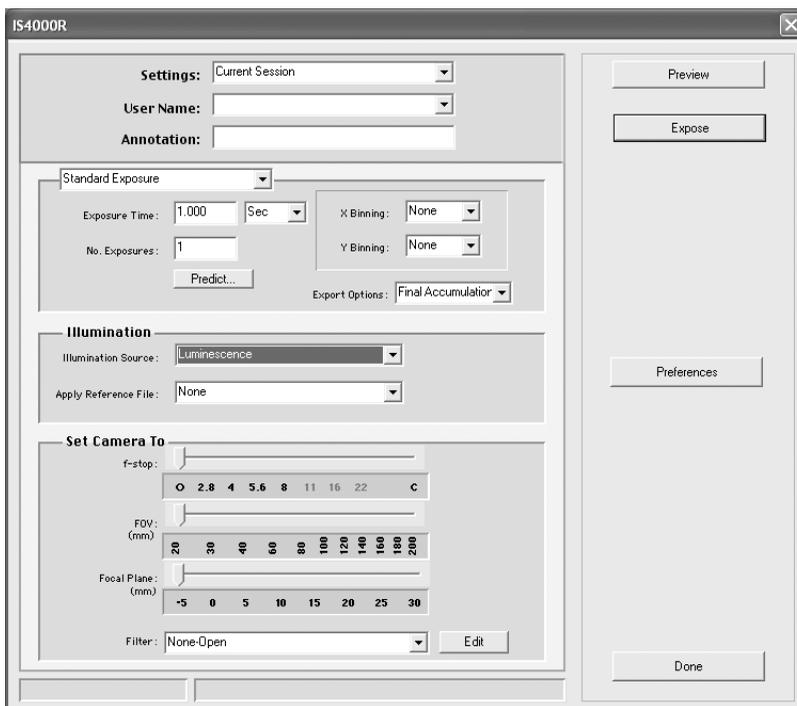
WARNING: Do not expose the Platen to strong chemical solvents such as ketones, hexanes, acids and alkalis that may permanently damage the Platen. This type of damage is not covered by the warranty.

Launching the Software

A few minutes before image capture, prepare the system by opening the IS4000R Acquire window.

- 1** Launch KODAK Molecular Imaging Software by clicking the KODAK MI icon found in the KODAK MI folder.
- 2** Choose Select Digital Camera from the File menu and IS4000R from the Digital Camera pop-up menu.
 - ✓ If you have previously used the IS4000R, click the IS4000R Capture button on the Status bar. The software searches for that camera in the port configuration last used.
 - ✓ If the IS4000R camera is not found, the Select Digital Camera dialog box appears allowing you to select the camera. Choose IS4000R from the Digital Camera pop-up menu and click OK.
 -  **NOTE:** The Port and Connection Speed pop-up menus will be disabled because these are not user-defined options for IEEE 1394 (FireWire) devices.
 - ✓ If a IS4000R camera is found, the software remembers the camera port and in the future automatically looks at this port first. The File menu and the Status bar Camera Access button updates to reflect the current camera selection.
- 3** You will be prompted upon first selection of the IS4000R to state whether you have purchased the optional Image Station Transilluminator Lid Accessory. Select Yes or No as appropriate.

4 The IS4000R Acquire window appears.



Orienting the Sample for Capture

Now that the Platen is clean and the IS4000R Acquire window is open, you are ready to orient your sample on the center of the Platen. Placement of your sample is similar to placing a document on a photocopier.

Non-Radioactive Sample Orientation

The Platen accommodates samples up to 20 x 20 cm. The Platen area is water-tight and inert, ideal for biochemical, luminescence, and other photochemical reactions.

- 1 Open the IS4000R lid.
- 2 Place the sample on the center of the Platen. Use plastic tweezers (included with the system) to avoid scratching the Platen.
 - ✓ For opaque samples (blots, small animals, plants)—orient the sample labelled side face down on the Platen.
 - ✓ For translucent samples (gels, autorads, culture plates, flasks and well plates)—orient the sample either way on the Platen.
 - ✓ If the sample is wet (gels, blots), puddle water or buffer on the Platen prior to placement of the sample to minimize bubble artifacts.

 NOTE: Leave the lid open to use ambient light for visualizing the sample.



WARNING: Do not use metal utensils or tweezers since they may scratch and permanently damage the Platen. This type of damage is not covered by the warranty.



WARNING: Do not expose the Platen to strong chemical solvents such as ketones, hexanes, acids and alkalis that may permanently damage the Platen. This type of damage is not covered by the warranty.

- 3 Proceed to *Previewing the Image*, later in this chapter.

Radioactive Sample Orientation Using the rad PADD Accessory

The rad PADD accommodates blots, membranes, or gels up to 16 x 16 cm.

- 1 Open the rad PADD cassette—the phosphor screen of the cassette should be placed facing on the Platen.
- 2 Place the sample on the center of the phosphor screen.



- ☞ NOTE: Leave the lid open to use ambient light for visualizing the sample. After previewing, close the cassette and close the IS4000R lid.
- ☞ NOTE: To decontaminate your screen, refer to *Radiological Decontamination Procedure* in Chapter 6: *Maintaining the System*.

Previewing the Image

The Preview mode offers you a way to position your sample and preview what the camera will image. It is important to maximize the number of pixels across the area you want to image and make sure that the image is in focus.

- 1 Click the Preview button to view the sample on the screen. The IS4000R Preview window opens.

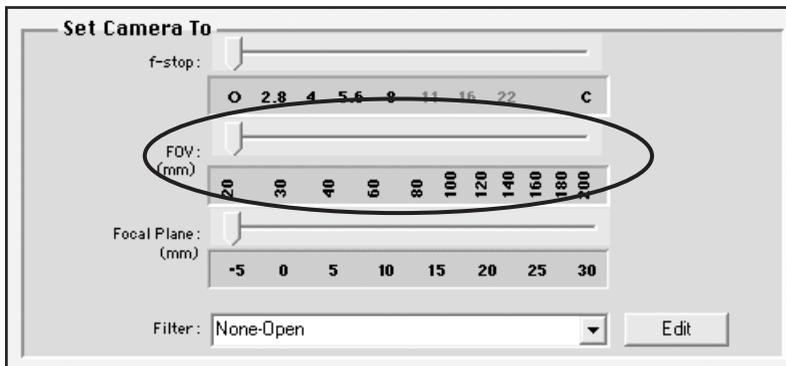


The Preview is designed to aid you in sample placement using ambient light.

- 2 Use the Exposure Time slider to set an appropriate preview exposure time. The maximum exposure time is 5 seconds.
-  **NOTE:** Focusing the camera should not be necessary for most samples placed on the Platen. To focus, adjust the Focus Ring on the camera lens. A special mark has been placed in the approximate parfocal location at the Platen.



- NOTE: Precision focus is available using the software-assisted focusing tool in the Preview mode. See *Using the Focusing Tool*, later in this chapter.
- NOTE: Focus adjustments are required when using the rad PADD.
- 3** Adjust the Field of View (FOV) Ring on the camera lens to fill the entire frame with the sample image or features of interest. This maximizes the number of pixels within your sample image.
- NOTE: The field of view setting relates directly to the image resolution. The IS4000R CCD has a pixel resolution of 2048 x 2048 pixels. At the smallest field of view (FOV 20), the number of pixels are spread over a 2 x 2 cm area producing a resolution of ~10 microns/pixel. At the largest field of view setting (FOV 200), the pixels are spread over a 20 x 20 cm area (~100 microns/pixel). An important consideration is the number of pixels that are applied to the smallest feature of interest.
- 4** Click Done when you are satisfied with your field of view.
- 5** Record the field of view setting using the FOV slider in the Set Camera To section of the IS4000R Acquire window.



Choosing the Filter and Illumination Conditions

This section contains the basic guidelines on how to set up the illumination and filters prior to image capture.

- 1 Select the filter using the Filter Selection Dial located at the top of the Capture System Chamber. The filters available for the IS4000R are described in Table 4.2: *Filter Selection*.

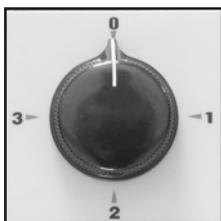


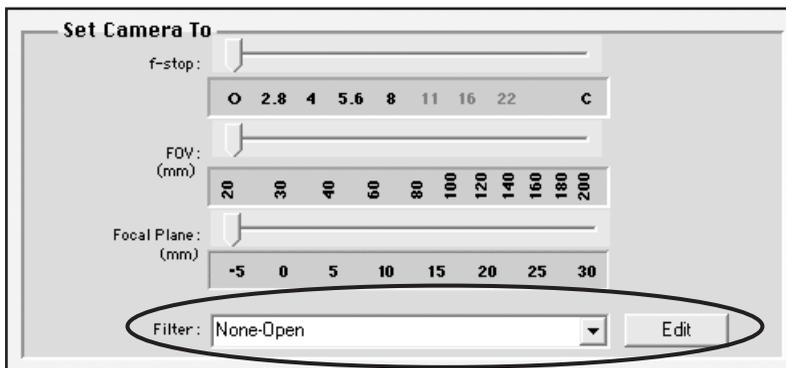
Table 4.1: Filter Selection

Filter	Wavelength	Applications	Comments
No Filter	—	film, Coomassie stained gels, chemiluminescence and radioisotopic	Transmission mode densitometry, rad PADD, chemiluminescence
600 nm WB	600 nm \pm 35	ethidium bromide, Ruby Red, Texas Red, Ruby Orange	Suppresses fluorescein and Cy-5
670 nm WB	670 nm \pm 35	Cy-5, TOT3, Acrydine Orange-RNA	Discriminates against fluoresceins and suppresses ethidium bromide and Cy-3
570 nm WB	570 nm \pm 35	Cy-3, Pyronin Y	Suppresses Cy-5
535 nm WB	535 nm \pm 35	Fluorescein, Rhodamine, Eosin, SYBR Green, SYBR Gold, Acrydine Orange-DNA	Suppresses ethidium bromide and discriminates against Cy-5
435 nm WB	435 nm \pm 35	DAPI, Hoescht, SYTO Blue, dansyl chloride	

 NOTE: IS4000R is supplied with the 600 nm WB filter (ethidium bromide). Specialized filters, as described above, are available through Molecular Imaging Systems. To change a filter, refer to *Replacing or Changing an Emission Filter*

in Chapter 6: *Maintaining the System*.

- Record the filter selection using the Filter pop-up menu in the Set Camera To section of the IS4000R Acquire window.



 **NOTE:** If your filter is not listed in the Filter pop-up menu, use the Edit button to define the filter using the Available Filter dialog box. See *Replacing or Changing an Emission Filter* in Chapter 6: *Maintaining the System*.

- Choose the appropriate sample background:

✓ Luminescence

Blot (flat)—use the black side of the Compression Pad to apply modest sample pressure.

✓ UV Epi-illumination

Thick samples (gel or other)—remove the Compression Pad from the lid.

Thin samples (blot or other)—use the black side of the Compression Pad to apply modest sample pressure.

UV fluorescence quenching with an autofluorescing sample (blot, TLC)—use the black side of the Compression Pad to apply modest sample pressure.

UV fluorescence quenching with a clear sample (thin gel, microscope slide)—use the white side of the Compression Pad to assure light return and to apply modest sample pressure. Assemble gel sandwich wet.

✓ UV Transillumination UV with the optional Image Station Transilluminator Lid Accessory—make sure the transilluminator is clean.

- ✓ White Light Transillumination with Light Diffuser

Thin samples (transmission)—place the Light Diffuser directly onto the sample to provide even distribution of external light and modest sample pressure.

Thick samples (transmission)—use the Light Diffuser Risers or add shims to space the diffuser from the sample.

- ✓ White Light Transillumination with the optional Image Station Transilluminator Lid Accessory—make sure the transilluminator is clean. Attach the White Light PADD.

- ✓ Radioisotopic—use the rad PADD accessory cassette. Place the sample on the phosphor screen and close the cassette.

4 If your sample requires illumination, power the illumination source.

- ✓ Luminescence—none

- ✓ UV Epi-illumination—use the UV ON/OFF switch on the front of the Capture System Chamber.

- ✓ UV fluorescence—use the UV ON/OFF switch on the front of the Capture System Chamber.

- ✓ UV Transillumination with the optional Image Station Transilluminator Lid Accessory—use the ON/OFF switch on the front of the lid.

- ✓ White Light Transillumination with Light Diffuser—none

- ✓ White Light Transillumination with the optional Image Station Transilluminator Lid Accessory—use the ON/OFF switch on the front of the lid.

- ✓ Radioisotopic—none



NOTE: Most UV and transmissions images will benefit from using illumination reference to correct non-uniform in sample illumination. If you have generated an Illumination Reference File, apply by selecting an Illumination Reference File with the corresponding f -stop and FOV from the pop-up menu. The f -stop and field of view settings used to capture the image must be the same as those used to generate the Illumination Reference File. If you need to create a new reference file, see *Generating an Illumination Reference File Library* in Chapter 5: *Capturing Images*.

5 Depending on the illumination mode, you may need to close and clasp the lid.

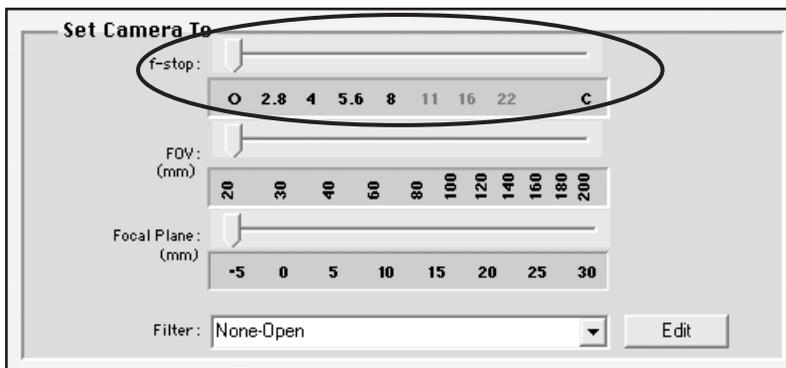
- ✓ Luminescence—closed
- ✓ UV Epi-illumination—closed
- ✓ UV fluorescence—closed
- ✓ UV Transillumination with the optional Image Station Transilluminator Lid Accessory—closed
- ✓ White Light Transillumination with Light Diffuser—open
- ✓ White Light Transillumination with the optional Image Station Transilluminator Lid Accessory—closed
- ✓ Radioisotopic—closed (both rad PADD cassette and lid)

Setting the f -Stop

The f -stop or lens aperture determines the amount of light that passes through the lens, while the exposure time controls how long the CCD is exposed. You will improve resolution using a higher f -stop (smaller apertures), however since signal is reduced, a longer exposure time may be required.

Generally, we recommend that you use f -stop O (Open) for speed and resolution. For flat formats, f -stops higher than f O will not substantially contribute to image resolution. However, you may need to increase the f -stop for unusually thick samples. Note that the exposure time is related to the inverse square of the f -stop.

- 1 Set the f -stop by using the f -Stop Ring on the lens. Use these guidelines:
 - ✓ Luminescence—use the largest aperture f O to maximize signal
 - ✓ UV epi-illumination—when exposure time is unusually long (dim fluorescence), use f 2.8
 - ✓ UV transillumination—use f O to f 2.8
 - ✓ White Light Transillumination—use f 2.8 to f 11 to extend exposure time so you have greater control of your dynamic range
 - ✓ Radioisotopic—use the largest aperture f O to maximize signal
- 2 Record the f -stop setting using the f -Stop slider in the Set Camera To section of the IS4000R Acquire window.



 NOTE: The f -stop on the IS4000R lens is adjustable. It is important that you record the setting for accurate lens correction.

Image Capture Guidelines Summary

The following table is a summary of these guidelines and helpful suggestions:

Table 4.2: Image Capture Guidelines

Detection Method	Examples	Exposure Type*	f-stop	Illumination	Typical Exposure Time	Comments
Luminescence	Chemi blots	Standard exposure, binning,	$f0$ (Open)	None	Equal to film exposure, < 15 minute for single exposure, > 15 min for multiple exposures accumulated	Lid closed. Use Black Pad.
UV epi-illumination	Gels and blots	Standard exposure, no binning,	$f0$ (Open)	UV epi-illumination , Lid closed	5 second single exposure	Lid closed. Use Black Pad.
UV Transillumination	Ethidium bromide gels and fluorescent blots	Standard exposure, no binning	$f0$ – $f8$	Transilluminator Lid Accessory	< 1 second single exposure	Optional Image Station Transilluminator Lid Accessory
White Light transillumination	Autorad film Coomassie gels plates	Standard exposure, no binning	$f2.8$ – $f11$	External or ambient overhead	0.05 second single exposure	Lid open. Use Light Diffuser
White Light transillumination	Autorad film Coomassie gels plates	Standard exposure, no binning	$f2.8$ – $f11$	External or ambient overhead	0.05 second single exposure	Optional Image Station Transilluminator Lid Accessory
UV epi-illumination Fluorescence Quenching (opaque sample)	Chromogenic blots TLC plates	Standard exposure, no binning	$f0$ (Open) - $f8$	UV epi-illumination	0.5–2 second(s) single exposure	Lid closed. Lid closed. Use Black Pad.
UV epi-illumination Fluorescence Quenching (clear sample)	Stained gels	Standard exposure, no binning,	$f0$ (Open) – $f8$	UV epi-illumination	0.5 second single exposure	Lid closed. Use White Pad
Radioisotopes	Northern/Southern blots, TLC, Kinetic experiments	rad PADD, 4 x 4 binning,	$f0$ (Open)	None	Equal to film exposure. < 3 minute for single high signal option. > 3 min for multiple exposures accumulated—low signal option	rad PADD accessory is required for this capture mode. Sample in closed cassette, IS4000R lid closed.

* Standard Single exposures provide excellent quality 16-bit images that are good for most applications, Standard multiple accumulated image options may also be used to improve quantitation and dynamic range—beyond 16-bit.

Using the Focusing Tool

The IS4000R has been designed to be parfocal at the Platen. If you should you need to adjust the focus, use the Focus Ring.

If your samples vary in thickness or are using the rad PADD, the software-assisted focusing tool is available in the Preview mode. The software determines camera focus by using the sharpness of edges within the image. To adjust focus:

- 1** Place your sample on the Platen.
- 2** Click Preview.
- 3** Adjust the Zoom to fill your field of view with the sample.
- 4** Place a ruler or any sharp edged flat object on top of your sample.
- 5** Click the Focus Camera checkbox. A selection rectangle will appear on screen, in the center of the image.
- 6** Adjust the Focus Ring to maximize the numerical value appearing in the text box.
 - ✓ Increasing values indicate focus is sharpening, whereas decreasing values indicate blurring of the image.
 - ✓ A contrast target placed at the same focal plane will improve your determination of focus.

To exit the focusing mode, uncheck the Focus Camera checkbox.

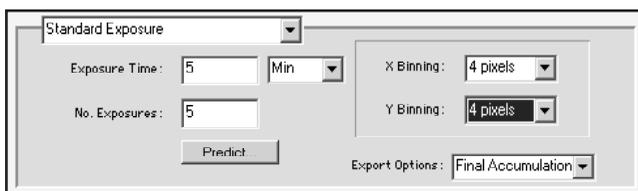
Capturing Images

Now you will step through the procedures for capturing and submitting KODAK Image Station 4000R (IS4000R) images. The IS4000R supports a wide variety of capture modes, providing you with a flexible, multi-purpose imaging system for your laboratory. Capture modes include single frame, multiple frame, time lapse, progressive, and rad PADD exposures.

Using the Standard Exposure Option

Use the Standard Exposure option to take a single image or capture multiple images. When you capture multiple images you can choose to save each capture as a separate file, accumulate all the captures in a single file, or generate a series of accumulations. To increase sensitivity you may also choose to X and/or Y bin your image.

- 1 Choose Standard Exposure from the Exposure pop-up menu. The Standard Exposure options appear.

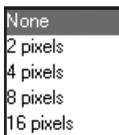


- 2 Enter the desired exposure time using the Exposure Time text edit box and the Units pop-up menu. Choose an exposure time to maximize the use of all available signal levels in the image without saturation.

The minimum and maximum exposure times are 0.05 second and 100 minutes per capture, respectively. The minimum time for binned exposures is 1 second. The time units can be set as seconds or minutes. The minimal increment is 0.1 second.

- ✓ Luminescence—use the exposure time you typically use with film or use the Predict button.
- ✓ UV Epi-illumination—start with an exposure time of 5 seconds per capture.
- ✓ UV Transillumination with the optional Image Station Transilluminator Lid Accessory—start with an exposure time of 0.5 second per capture.
- ✓ White Light Transillumination with Light Diffuser—start with an exposure time of 0.1 second per capture.
- ✓ White Light transillumination with the optional Image Station Transilluminator Lid Accessory and White Light PADD—start with an exposure time of 0.05 second per capture.
- ✓ rad PADD—use the exposure time you typically use with film or use the Predict button.

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- ☞ NOTE: For chemiluminescence and rad PADD, use the Predict button to open a dialog box to get a prediction of the optimal exposure time based upon the desired number of gray levels (GL). When the Calculate button is pressed, the camera takes a series of exposures to predict the optimal exposure time. Once determined, the predicted exposure time and number of exposures are automatically updated in the Exposure Time and No. Exposures text edit boxes.
 - ☞ NOTE: For multiple captures, use the IS4000R Preferences window to set a capture preference that automatically stops image capture when saturation occurs in a single frame. Additionally, in the Capture Status window, you can choose to display the saturated pixels on screen in red.
- 3 Enter the number of exposures using the No. Exposure text edit box. You can choose to take up to 32 exposures.
 - ✓ Multiple captures are recommended for weak signal samples where exposures of greater than 20 minutes are required. You may also choose to do multiple exposures if you want to increase dynamic range.
 - 4 To increase sensitivity, you may also choose to X and/or Y bin your image using the X Binning and Y Binning pop-up menus.



- ✓ Select binning options for low signal conditions, such as luminescent and rad PADD exposures. Experimental considerations can also influence the choice of binning settings, such as preferential spatial resolution in either the X or Y direction. Therefore, if you need to resolve closely spaced bands in the Y-direction, consider binning only in the X direction to preserve the integrity of band resolution in the Y-direction while enhancing signal. This is called asymmetric binning, enabling a maximum of 8 x 1 binning that yields an 8X signal increase. When necessary, use the pop-up menus to select X and/or Y binning and the number of pixels.
- ✓ Selecting X and/or Y binning combines the values from two, four, eight or sixteen adjacent pixel signals in the direction(s) selected. Expect up to a 256-fold increase in the rate of signal accumulation when 16 x 16 X and Y binning are selected.

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- 5 For multiple exposures, you can choose the types of images you want to generate from the exposure using the Export options pop-up menu.



- ✓ *Final Accumulation* opens only the final image as a project.
 - ✓ *All Accumulations* saves each accumulation to the hard drive and opens the final accumulation.
 - ✓ *All Images* displays the final accumulation as a project and saves the intermediate accumulations and each of the separate image files on your hard drive.
 - ✓ *Separate Images* saves each separate exposure on your hard disk.
- 6 Optional: Select your name from the User pop-up menu or enter your name in the User text edit box in the User section of the IS4000R Acquire window. Select Add User to save the user name.
- 7 Optional: Type text (up to 48 characters) in the Annotation text edit box. The Annotation bar is appended to the image upon printing and to the image when opened in the Annotation mode.
- ✓ When you choose to annotate your image, illumination type, time of capture, and exposure time and date are appended to your annotation.
 - ✓ Each time you take a picture the Annotation text edit box is cleared.
 - ✓ Display or hide the Annotation bar by selecting the Show Annotation bar from within the Annotation panel.
 - ✓ All items in the Annotation bar are separate objects in the Annotation window. This allows you to edit or change the font style, color, or size prior to printing. The contents, however are grouped and must be ungrouped prior to editing.
- 8 Choose the Illumination Source from the pop-up menu in the IS4000R Acquire window.
- 9 Choose a reference file (as appropriate) from the Apply Reference pop-up menu.
- ✓ Use a reference file to correct for uneven illumination when imaging using epi-illumination or Transillumination light sources.
 - ✓ A library of illumination corrections can be generated for your convenience, refer to *Generating an Illumination Reference File Library* section later in this chapter.

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- ✓ Reference files must match illumination and capture settings of the image you are capturing. A warning message appears if you try to apply a reference file with incorrect parameters.

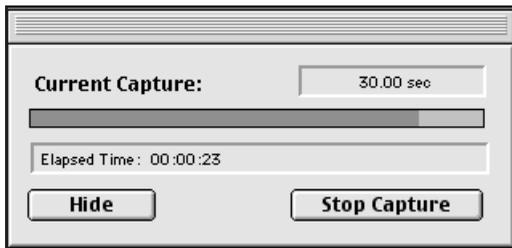
- ✓ Automatic can be chosen to create a reference file at the time of image capture.

10 Click Expose. If you have set multiple exposures with intermediate or separate files options, the Save dialog box appears prompting you to select a location to save the files. Choose the destination drive and file name (and folder for multiple image captures). Click Save to start the capture.

- 👉 **NOTE:** You can save your capture parameters using the Settings option in the User section of the IS4000R Acquire window. See *Custom Capture Settings* later in this chapter.

11 The Capture Status window appears.

For single image captures, the following window appears.



- ✓ The *Current Capture* text field indicates the exposure time that was set for the capture in progress.
- ✓ The *Current Capture* progress bar displays the status of the capture.
- ✓ The *Elapsed Time* text field indicates the elapsed time since the beginning of the exposure.
- ✓ Use the *Hide* button to close the window without stopping the exposure. You can access the window by selecting IS4000R from the File menu or the IS4000R Capture button on the Status bar.
- ✓ Use the *Stop Capture* button to interrupt the capture and return to the IS4000R Acquire window. No Image is saved.

12 Turn off the light source, if appropriate.

13 For multiple image captures, the following window appears.



- ✓ The *Elapsed Time* text field shows the elapsed time since the beginning of the exposure.
- ✓ The *Current Capture* text field indicates the current capture in the series.
- ✓ The *Current Capture* progress bar indicates the status of the current exposure.
- ✓ The *Expt Duration* text field displays the total time from the beginning of the first frame to the end of the last frame—regardless of the exposure type.
- ✓ The *Expt Duration* progress bar shows the status of the entire experiment (including all exposures).
- ✓ Use the *Stop Capture* button to interrupt the current capture and return to the Image Window. Any images saved to the hard drive prior to selecting this button are preserved. All the appropriate image filters/corrections are applied. If selected, a final accumulated image is opened.
- ✓ *Dynamic Range* provides signal intensity information about the image being captured. *Min* and *Max* text fields display the current minimum and maximum signal values within the image. The *Show Saturation* check box shows any

saturated pixels in the image in red. The *Auto Stop at* check box sets a dynamic range at which the capture process is interrupted. For example, you can set the auto stop at value to 10,000 in the *AutoStop at* text edit box. Whenever an accumulation is added and the difference between the maximum and minimum values is greater than 10,000 signal values, the capture process stops and the image is opened.

- ☞ NOTE: If the image reaches saturation immediately (image turns red), decrease the exposure time or use a higher *f*-stop setting.
- ☞ NOTE: If the image is not near saturation, estimate the exposure time by using the Predict button.
- ☞ NOTE: To achieve the maximum dynamic range, the exposure time estimate determined above (filling the 16-bit scale with a single capture) should be implemented using the accumulation of 32 captures (filling the floating point bit scale with accumulated signal). Intermediate levels of dynamic range may be achieved by accumulating fewer images.
- ✓ The *Analyze* button transfers the last completed image or accumulation captured for analysis. Use this option anytime during the capture process to open intermediate images. Selecting this option will not stop the capture in progress.
- ✓ Use the *Hide* button to close the window without stopping the exposure. You can access the window by selecting IS4000R from the File menu or by using the IS4000R Capture button on the Status bar.
- ✓ The *Information* bar directly below the image contains the current exposure type and user name.

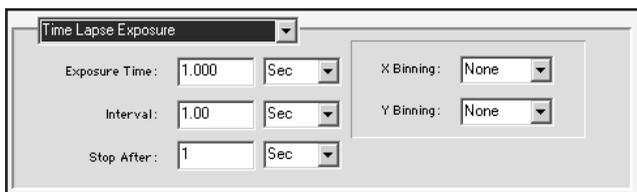
Upon completion a dialog box appears alerting you that the capture is complete.

- 14 Once the exposure(s) are completed, you are ready to print, analyze, or annotate your image.
 - ✓ Single captures are opened as KODAK MI projects on your desktop.
 - ✓ Final accumulation images are opened as projects on your desktop.
 - ✓ Intermediate accumulations are saved as projects to your hard drive.
 - ✓ Separate images from a multiple image capture are saved as projects to your hard drive.
- 15 Turn off the light source, if appropriate.

Using the Time Lapse Exposure Option

The Time Lapse exposure option is designed to take multiple exposures over fixed time intervals. For example, you may take a 10 second exposure every hour over the next 32 hours.

- 1 Choose Time Lapse Exposure from the Exposure pop-up menu. The Time Lapse Exposure options appear.



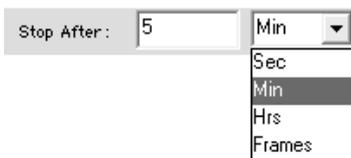
- 2 Enter the desired exposure time using the Exposure Time text edit box and the Units pop-up menu. Choose an exposure time to maximize the use of all available signal levels in the image without saturation.

The minimum and maximum exposure times are 0.05 second and 100 minutes per capture, respectively. The time units can be set as seconds or minutes. The minimal increment is 0.1 second.

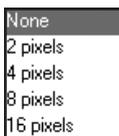
- ✓ Luminescence—use the exposure time you typically use with film.
- ✓ UV Epi-illumination—start with an exposure time of 5 seconds per capture.
- ✓ UV Transillumination with the optional Image Station Transilluminator Lid Accessory—start with an exposure time of 0.1 second per capture.
- ✓ White Light Transillumination with Light Diffuser—start with an exposure time of 0.2 second per capture.
- ✓ White Light transillumination with the optional Image Station Transilluminator Lid Accessory and White Light PADD—start with an exposure time of 0.05 second per capture.
- ✓ rad PADD—use the exposure time you typically use with film.

 **NOTE:** For multiple captures, use the IS4000R Preferences window to set a capture preference that automatically stops image capture when saturation occurs in a single frame. Additionally, in the Capture Status window, you can choose to display the saturated pixels on screen in red.

- 3 Enter the interval in the Interval text edit box and the Units pop-up menu. The time interval is the time between the beginning of one exposure and the beginning of the next exposure. Thus, it must exceed the entry for the exposure time. The maximum time interval is 24 hours.
- 4 Enter the stop criteria using the Stop After text edit box and the Units pop-up menu. Using this pop-up menu you can choose to set the stop criteria based on a total elapsed time or by the number of frames. The maximum duration is 150 hours or 32 captures.



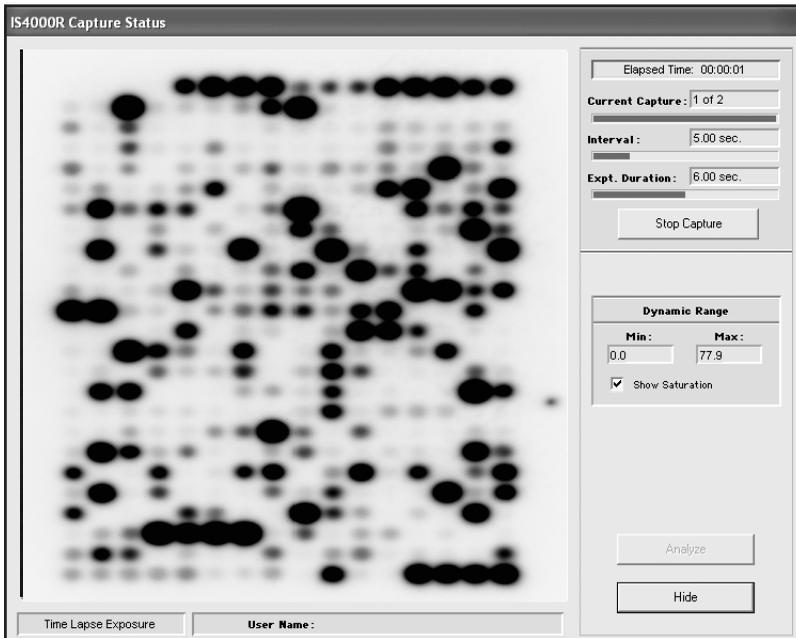
- 5 To increase sensitivity, you may also choose to X and/or Y bin your image using the X Binning and Y Binning pop-up menus.



- ✓ Select binning options for low signal conditions, such as luminescent and rad PADD exposures. Experimental considerations can also influence the choice of binning settings, such as preferential spatial resolution in either the X or Y direction. Therefore, if you need to resolve closely spaced bands in the Y-direction, consider binning only in the X direction to preserve the integrity of band resolution in the Y-direction while enhancing signal. This is called asymmetric binning, enabling a maximum of 8 x 1 binning that yields an 8X signal increase. When necessary, use the pop-up menus to select X and/or Y binning and the number of pixels.
 - ✓ Selecting X and/or Y binning combines the values from two, four, eight or sixteen adjacent pixel signals in the direction(s) selected. Expect up to a 256-fold increase in the rate of signal accumulation when 16 x 16 X and Y binning are selected.
- 6 Optional: Select your name from the User pop-up menu or enter your name in the User text edit box in the User section of the IS4000R Acquire window. Select Add User to save the user name.

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- 7** Optional: Type text (up to 48 characters) in the Annotation text edit box. The Annotation bar is appended to the image upon printing and to the image when opened in the Annotation mode.
- ✓ When you choose to annotate your image, illumination type, time of capture, and exposure time and date are appended to your annotation.
 - ✓ Each time you take a picture the Annotation text edit box is cleared.
 - ✓ Display or hide the Annotation bar by selecting the Show Annotation bar from within the Annotation panel.
 - ✓ All items in the Annotation bar are separate objects in the Annotation window. This allows you to edit or change the font style, color, or size prior to printing. The contents, however are grouped and must be ungrouped prior to editing.
- 8** Choose the Illumination Source from the pop-up menu in the IS4000R Acquire window.
- 9** Choose a reference file (as appropriate) from the Apply Reference pop-up menu.
- ✓ Use a reference file to correct for uneven illumination when imaging using epi-illumination or Transillumination light sources.
 - ✓ A library of illumination corrections can be generated for your convenience, refer to *Generating an Illumination Reference File Library* later in this chapter.
 - ✓ Reference files must match illumination and capture setting to the image you are applying them. A warning message appears if you try to apply a reference file with incorrect parameters.
- 10** Click Expose. A Save dialog box appears prompting you to select a location to save the files. Choose the destination drive and folder. Enter the file name and click Save.
-  **NOTE:** You can save your capture parameters using the Settings option in the User section of the IS4000R Acquire window. See *Custom Capture Settings* later in this chapter.

11 The Time Lapse Capture Status window appears.



- ✓ The *Elapsed Time* text field shows the elapsed time since the beginning of the exposure.
- ✓ The *Current Capture* text field indicates the current capture in the Time Lapse series.
- ✓ The *Current Capture* progress bar displays the status of the current capture.
- ✓ The *Interval* text field shows the elapsed time in the current time interval. The interval corresponds to the time between the beginning of one capture and the beginning of the next capture.
- ✓ The *Interval* progress bar displays the status of the capture during the current time interval.
- ✓ The *Expt Duration* text field displays the total time from the beginning of the first frame to the end of the last frame—regardless of the exposure type.
- ✓ The *Expt Duration* progress bar shows the status of the entire experiment (including all exposures).

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- ✓ Use the *Stop Capture* button to interrupt the current capture and return to the IS4000R Acquire window. Any images saved to the hard drive prior to clicking this button are preserved and all the appropriate image filters/corrections are applied.
 - ✓ *Dynamic Range* provides signal intensity information about the image being captured. *Min* and *Max* text fields display the current minimum and maximum signal values within the image. The *Show Saturation* check box shows any saturated pixels in the image in red.
 - ☞ NOTE: If the image reaches saturation immediately (image turns red), decrease the exposure time or set a higher *f*-stop setting.
 - ✓ The *Analyze* button transfers the last completed image or accumulation for analysis. You can choose this option anytime during the capture process to open intermediate images. Selecting this option will not stop the capture in progress.
 - ✓ The *Hide* button closes the window without stopping the exposure. Access the window by selecting IS4000R from the File menu or by using the IS4000R Capture button on the Status bar.
 - ✓ The *Information* bar directly below the image contains the current exposure type and user name.

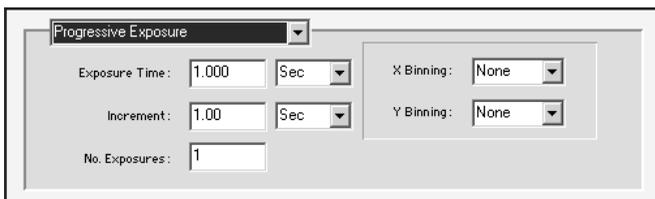
Upon completion a dialog box appears alerting you that the capture is complete.

- 12** Turn off the light source, if appropriate.
- 13** Once the exposure(s) are completed, you are ready to print, analyze, or annotate your image. Each separate image is saved as a project on your hard drive.

Using the Progressive Exposure Option

The Progressive Exposure option is designed to take a continuous sequence of exposures at different exposure times. You can automate complex exposure options—rather than manually having to set each exposure time. For example, you may want to capture an image at each of the following exposure times: 10, 20, 40, and 80 seconds.

- 1 Choose Progressive Exposure from the Exposure pop-up menu. The Progressive Exposure options appear.

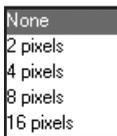


- 2 Enter the desired exposure time using the Exposure Time text edit box and the Units pop-up menu. Choose an exposure time to maximize the use of all available signal levels in the image without saturation.

The minimum and maximum exposure times are 0.05 second and 100 minutes per capture, respectively. The minimum exposure time for a binned image is 1 second. The time units can be set as seconds or minutes. The minimal increment is 0.1 second.

- ✓ Luminescence—use the exposure time you typically use with film.
 - ✓ UV Epi-illumination—start with an exposure time of 5 seconds per capture.
 - ✓ UV Transillumination with the optional Image Station Transilluminator Lid Accessory—start with an exposure time of 0.5 second per capture.
 - ✓ White Light Transillumination with Light Diffuser—start with an exposure time of 0.01 second per capture.
 - ✓ White Light transillumination with the optional Image Station Transilluminator Lid Accessory and White Light PADD—start with an exposure time of 0.05 second per capture.
 - ✓ rad PADD—use the exposure time you typically use with film.
-  **NOTE:** For multiple captures, use the IS4000R Preferences window to set a capture preference that automatically stops image capture when saturation occurs in a single frame. Additionally, in the Capture Status window, you can choose to display the saturated pixels on screen in red.

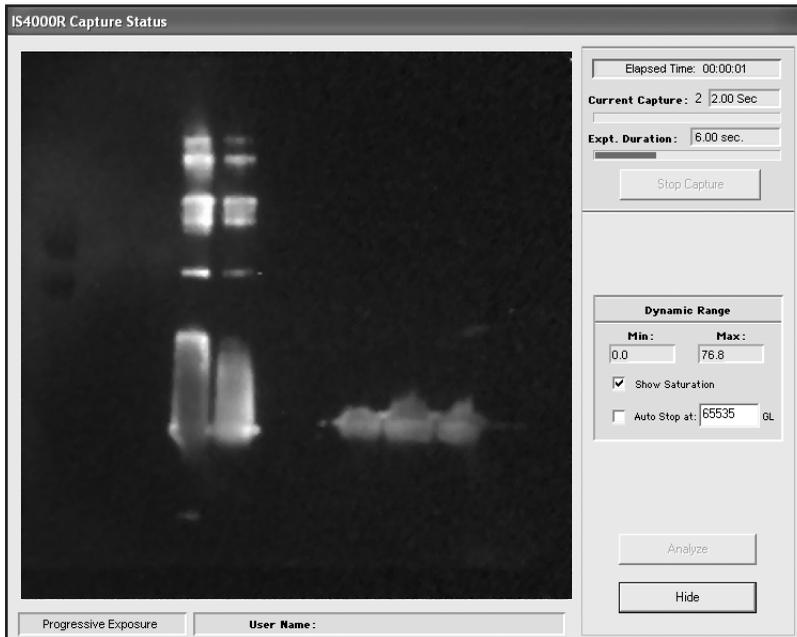
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- 3 Enter the increment value for each exposure, using the Increment text edit box and the Units pop-up menu. The minimum exposure time increment is 0.1 second.
 - ✓ The *Additive* option of *Sec* or *Min* adds the increment value to each exposure. For example: 2 sec + 2 sec = 4 sec, 4 sec + 2 sec = 6 sec, 6 sec + 2 sec = 8 sec.
 - ✓ The *Multiplier* option multiplies the increment to create a new exposure. For example: 2 sec x 2 sec = 4 sec, 4 sec x 2 sec = 8 sec, 8 sec x 2 sec = 16 sec.
 - ✓ The *Geometric Order* option raises each exposure time to the power of the increment value entered. For example: (2 sec)² = 4 sec, (4 sec)² = 16 sec, (16 sec)² = 256 sec. This option is particularly relevant to maximize the signal-to-noise ratio of a CCD image.
 - 4 Enter the number of exposures using the No. Exposure text edit box. You can choose to take up to 32 exposures.
 - 5 To increase sensitivity, you may also choose to X and/or Y bin your image using the X Binning and Y Binning pop-up menus.



- ✓ Select binning options for low signal conditions, such as luminescent and rad PADD exposures. Experimental considerations can also influence the choice of binning settings, such as preferential spatial resolution in either the X or Y direction. Therefore, if you need to resolve closely spaced bands in the Y-direction, consider binning only in the X direction to preserve the integrity of band resolution in the Y-direction while enhancing signal. This is called asymmetric binning, enabling a maximum of 8 x 1 binning that yields an 8X signal increase. When necessary, use the pop-up menus to select X and/or Y binning and the number of pixels.
 - ✓ Selecting X and/or Y binning combines the values from two, four, eight or sixteen adjacent pixel signals in the direction(s) selected. Expect up to a 256-fold increase in the rate of signal accumulation when 16 x 16 X and Y binning are selected.
- 6 Optional: Select your name from the User pop-up menu or enter your name in the User text edit box in the User section of the IS4000R Acquire window. Select Add User to save the user name.

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- 7** Optional: Type text (up to 48 characters) in the Annotation text edit box. The Annotation bar is appended to the image upon printing and to the image when opened in the Annotation mode.
 - ✓ When you choose to annotate your image, illumination type, time of capture, and exposure time and date are appended to your annotation.
 - ✓ Each time you take a picture the Annotation text edit box is cleared.
 - ✓ Display or hide the Annotation bar by selecting the Show Annotation bar from within the Annotation panel.
 - ✓ All items in the Annotation bar are separate objects in the Annotation window. This allows you to edit or change the font style, color, or size prior to printing. The contents, however, are grouped and must be ungrouped prior to editing.
 - 8** Choose the Illumination Source from the pop-up menu in the IS4000R Acquire window.
 - 9** Choose a reference file (as appropriate) from the Apply Reference pop-up menu.
 - ✓ Use a reference file to correct for uneven illumination when imaging using epi-illumination or Transillumination light sources.
 - ✓ A library of illumination corrections can be generated for your convenience, refer to *Generating an Illumination Reference File Library* later in this chapter.
 - ✓ Reference files must match illumination and capture setting to the image you are applying them. A warning message appears if you try to apply a reference file with incorrect parameters.
 - 10** Click Expose. A Save dialog box appears allowing you to select a destination drive and folder. Enter the file name and click Save.
 - 11** Turn off the light source, if appropriate.

12 The Progressive Capture Status window appears.



- ✓ The *Elapsed Time* text field shows the elapsed time since the beginning of the exposure.
- ✓ The *Current Capture* text field indicates the number of the current exposure in the progressive series being captured.
- ✓ The *Current Capture* progress bar displays the status of the current capture.
- ✓ The *Expt Duration* text field displays total time from the beginning of the first frame to the end of the last frame—regardless of the exposure type.
- ✓ The *Expt Duration* progress bar shows the status of the entire experiment including all progressive exposures.
- ✓ Use the *Stop Capture* button to interrupt the current capture and return to the IS4000R Acquire window. Any images saved to the hard drive prior to clicking this button are preserved and all the appropriate image filters/corrections are applied.
- ✓ *Dynamic Range* provides signal intensity information about the image being captured. *Min* and *Max* text fields display the current minimum and maximum signal values within the image. The *Show Saturation* check box shows any

saturated pixels in the image in red. The *Auto Stop at* check box sets a dynamic range at which the capture process is interrupted. For example, you can set the auto stop value at 10,000 in the *Auto Stop at* text edit box. Whenever an accumulation is added and the difference between the maximum and minimum value is greater than 10,000, the capture process stops and the image is transferred. The default setting is 65,535 signal intensities.

 NOTE: If the image reaches saturation immediately (image turns red), decrease the exposure time or set a higher *f*-stop setting.

- ✓ The *Analyze* button transfers the last completed image or accumulation for analysis. Use this option anytime during the capture process to open intermediate images. Selecting this option will not stop the capture in progress.
- ✓ Use the *Hide* button to close the window without stopping the exposure. You can access the window by selecting IS4000R from the File menu or by using the IS4000R Capture button on the Status bar.
- ✓ The *Information* bar directly below the image contains the current exposure type and user name.

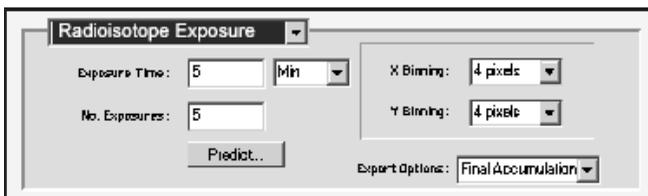
Upon completion a dialog box appears alerting you that the capture is complete.

- 13** Turn off the light source, if appropriate.
- 14** Once the exposure(s) are completed, you are ready to print, analyze, or annotate your image. Each separate image is saved as a project on your hard drive.

Using the rad PADD Exposure Option

The rad PADD cassette is an optional accessory available for your IS4000R, allowing detection of high energy radioisotopes. This option allows you to take a single image or capture multiple images. When you capture multiple images you can choose to save each capture as a separate file, accumulate all the captures in a single file, or generate a series of accumulations. To increase sensitivity you may also choose to X and/or Y bin your image.

- 1 Choose rad PADD from the Exposure pop-up menu. The rad PADD options appear.



- 2 Enter the desired exposure time using the Exposure Time text edit box and the Units pop-up menu. Choose an exposure time to maximize the use of all available signal levels in the image without saturation.

The minimum and maximum exposure times are 0.05 second and 100 minutes per capture, respectively. The minimum exposure time for binned images is 1 second. The time units can be set as seconds or minutes. The minimal increment is 0.1 second.

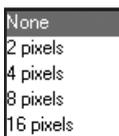
- ✓ rad PADD—use the exposure time you typically use with film or use the Predict button.

☞ NOTE: For rad PADD, use the Predict button to open a dialog box to get a prediction of the optimal exposure time based upon the desired number of gray levels (GL). When the Calculate button is pressed, the camera takes a series of exposures to predict the optimal exposure time. Once determined, the exposure time and number of exposures are automatically entered in the Exposure Time and No. Exposures text edit boxes.

☞ NOTE: For multiple captures, use the IS4000R Preferences window to set a capture preference that automatically stops image capture when saturation occurs in a single frame. Additionally, in the Capture Status window, you can choose to display the saturated pixels on screen in red.

- 3 Enter the number of exposures using the No. Exposure text edit box. You can choose to take up to 32 exposures.

- 4 To increase sensitivity, you may also choose to X and/or Y bin your image using the X Binning and Y Binning pop-up menus.



- ✓ Select binning options for low signal conditions, such as luminescent and rad PADD exposures. Experimental considerations can also influence the choice of binning settings, such as preferential spatial resolution in either the X or Y direction. Therefore, if you need to resolve closely spaced bands in the Y-direction, consider binning only in the X direction to preserve the integrity of band resolution in the Y-direction while enhancing signal. This is called asymmetric binning, enabling a maximum of 8 x 1 binning that yields an 8X signal increase. When necessary, use the pop-up menus to select X and/or Y binning and the number of pixels.
 - ✓ Selecting X and/or Y binning combines the values from two, four, eight or sixteen adjacent pixel signals in the direction(s) selected. Expect up to a 256-fold increase in the rate of signal accumulation when 16 x 16 X and Y binning are selected.
- 5 For multiple exposures, you can choose the type of file you want to generate from the exposure using the Export options pop-up menu.



- ✓ *Final Accumulation* opens only the final image as a project.
 - ✓ *All Accumulations* saves each accumulation to the hard drive and opens the final accumulation.
 - ✓ *All Images* displays the final accumulation as a project and saves the intermediate accumulations and each of the separate image files on your hard drive.
 - ✓ *Separate Images* displays each image as it is captured.
- 6 Optional: Select your name from the User pop-up menu or enter your name in the User text edit box in the User section of the IS4000R Acquire window. Select Add User to save the user name.

7 Optional: Type text (up to 48 characters) in the Annotation text edit box. The Annotation bar is appended to the image upon printing and to the image when opened in the Annotation mode.

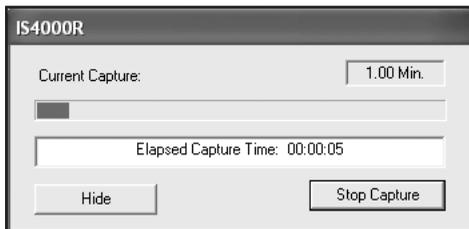
- ✓ When you choose to annotate your image, illumination type, time of capture, and exposure time and date are appended to your annotation.
- ✓ Each time you take a picture the Annotation text edit box is cleared.
- ✓ Display or hide the Annotation bar by selecting the Show Annotation bar from within the Annotation panel.
- ✓ All items in the Annotation bar are separate objects in the Annotation window. This allows you to edit or change the font style, color, or size prior to printing. The contents, however, are grouped and must be ungrouped prior to editing.

8 Click Expose. If you have set multiple exposures with intermediate or separate files options, the Save dialog box appears allowing you to select a location to save the files. Choose the destination drive and folder. Enter the filename and click Save.

 **NOTE:** You can save your capture parameters using the Settings option in the User section of the IS4000R Acquire window. See *Custom Capture Settings* later in this chapter.

9 The Capture Status window appears.

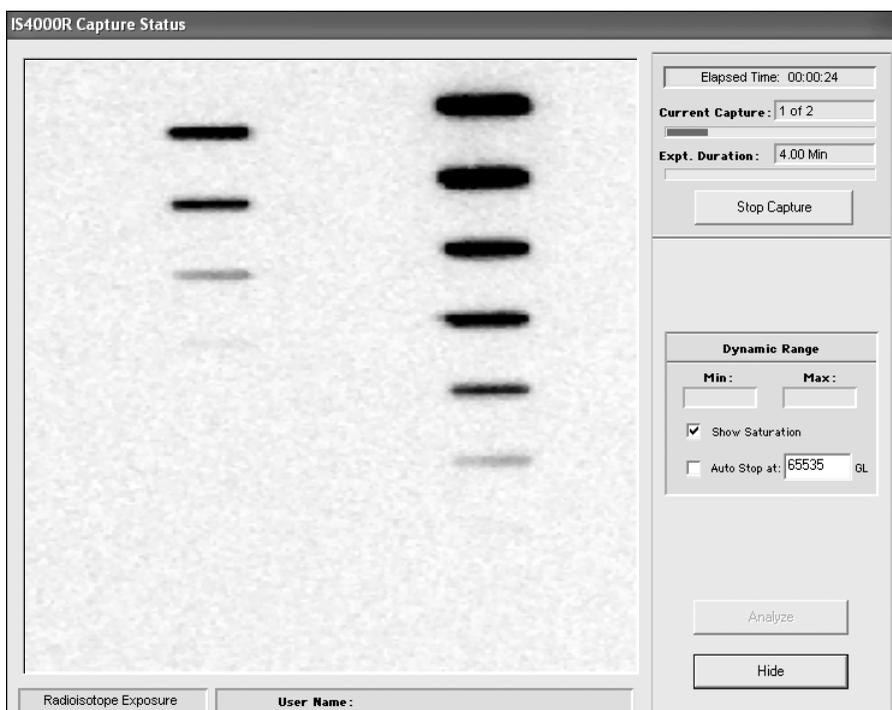
For single image captures, the following window appears.



- ✓ The *Current Capture* text field indicates the exposure time that was set for the capture in progress.
- ✓ The *Current Capture* progress bar displays the status of the capture.
- ✓ The *Elapsed Time* text field indicates the elapsed time since the beginning of the exposure.
- ✓ Use the *Hide* button to close the window without stopping the exposure. You can access the window by selecting IS4000R from the File menu or the IS4000R Capture button on the Status bar.

- ✓ Use the *Stop Capture* button to interrupt the capture and return to the IS4000R Acquire window. No image is saved.

For multiple image captures, the following window appears.



- ✓ The *Elapsed Time* text field shows the elapsed time since the beginning of the exposure.
- ✓ The *Current Capture* text field indicates the number of the current exposure in the progressive series being captured.
- ✓ The *Current Capture* progress bar displays the status of the current capture.
- ✓ The *Expt Duration* text field displays total time from the beginning of the first frame to the end of the last frame—regardless of the exposure type.
- ✓ The *Expt Duration* progress bar shows the status of the capture, including all exposures.

-
-
- ✓ Use the *Stop Capture* button to interrupt the current capture and return to the IS4000R Acquire window. Any images saved to the hard drive prior to clicking this button are preserved and all the appropriate image filters/corrections are applied. If selected, a final accumulated image is opened.
 - ✓ *Dynamic Range* provides signal intensity information about the image being captured. *Min* and *Max* text fields display the current minimum and maximum signal values within the image. The *Show Saturation* check box shows any saturated pixels in the image in red. The *Auto Stop at* check box sets a dynamic range at which the capture process is interrupted. For example, you can set the auto stop value at 10,000 in the *Auto Stop at* text edit box. Whenever an accumulation is added and the difference between the maximum and minimum value is greater than 10,000, the capture process stops and the image is transferred. The default setting is 65,535 signal intensities.

 NOTE: If the image reaches saturation immediately (image turns red), decrease the exposure time or set a higher *f*-stop setting.

- ✓ The *Analyze* button transfers the last completed image or accumulation for analysis. Use this option anytime during the capture process to open intermediate images. Selecting this option will not stop the capture in progress.
- ✓ Use the *Hide* button to close the window without stopping the exposure. You can access the window by selecting IS4000R from the File menu or by using the IS4000R Capture button on the Status bar.
- ✓ The *Information* bar directly below the image contains the current exposure type and user name.

Upon completion a dialog box appears alerting you that the capture is complete.

10 Once the exposure(s) are completed, you are ready to print, analyze, or annotate your image.

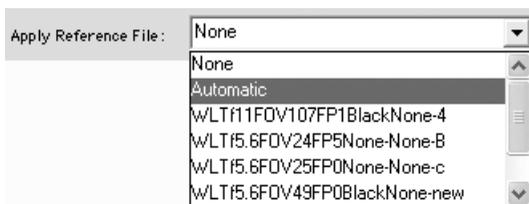
- ✓ Single captures are opened as projects.
- ✓ Final accumulation images are opened as projects.
- ✓ Intermediate accumulations are saved as projects to your hard drive.
- ✓ Separate images from a multiple image capture are saved as projects to your hard drive.

Applying an Automatic Illumination Reference File

Improve the quality of your data by applying field illumination correction to images captured. The illumination non-uniformity is highly reproducible and may be corrected by dividing the sample image by an illumination reference image. You can choose to create a library of illumination correction files (see *Generating an Illumination Reference File Library*) or generate an automatic illumination file following the procedure in this section.

To apply an automatic Illumination Reference file it is important to capture a reference image of the illumination field using the same camera settings (f -stop, zoom) used at the time you are capturing the image to be corrected.

- 1 Prior to capturing the image select the Automatic from the Apply Reference File pop-up window.



- 2 Capture an image as described in *Capturing Images*, earlier in this chapter.

👉 NOTE: Do not adjust the f -stop, zoom, exposure time, or position of lid. Upon completion of the capture the illumination correction dialog box appears.

- 3 Remove the sample from the imaging cabinet and clean the surface.
- 4 Enter a name in the Output name text edit box.
- 5 Click Continue. The illumination reference is applied and the output file opens as a new project.

👉 NOTE: You can not do an illumination reference correction if you have saturated pixels in your images.

Generating an Illumination Reference File Library

Improve the quality of your data by applying field illumination correction to images captured in the UV or white light imaging modes. The illumination non-uniformity is highly reproducible and may be corrected by dividing the sample image by an illumination reference image. It is important to capture a reference image of the illumination field using the same camera settings (f -stop, field of view) used when capturing the image to be corrected.

Preparing the Instrument for Creating Illumination Reference Files

- 1 Remove the sample from the Platen and clean the Platen surface. Prepare the surface of the imaging area as follows.
 - ✓ For Epi-illumination—Place the Compression Pad with the white side against the Platen. For best results, make sure that the Compression Pad is clean. Close the lid. Turn the Illuminator Power switch on.
 - 👉 NOTE: When using the optional Image Station Transilluminator Lid Accessory, a library of illumination correction files with varying f -stop and field of view combinations can be generated in advance. These files are saved in the Illumination Reference folder and are then available for application during image capture.
 - ✓ For White Light Transillumination with the Light Diffuser—place the Light Diffuser on the Platen. For best results, make sure that the Light Diffuser is clean. Open the lid to allow dim, ambient light to evenly illuminate the surface.
 - 👉 NOTE: Make sure that you maintain the same lid position when creating the Illumination Reference File and imaging your sample.
 - ✓ For White Light Transillumination with the optional Image Station Transilluminator Lid Accessory—make sure the transilluminator is clean. For white light attach the White Light PADD.
 - 👉 NOTE: For transillumination using the Image Station Transilluminator Lid Accessory, a library of illumination correction files with varying f -stop and field of view combinations can be generated in advance. These files are saved in the Illumination Reference folder and are then available for application during image capture.
- 2 Set the appropriate f -stop using the f -Stop Ring on the camera lens.
- 3 Set the field of view setting using the Field of View (FOV) Ring on the camera lens.

- 4 Move the Filter Selection Dial to position #1 to select the 600 nm WB filter (included with IS4000R).
- 5 Open the IS4000R Acquire window by choosing IS4000R from the File menu or by using the IS4000R Capture button on the Status bar.
- 6 Select Illumination Reference Exposure from the Exposure Type pop-up menu.



- 7 Set the exposure time.
 - ✓ For most transilluminated samples, set the exposure time to 0.05 second or adequate time to generate > 10,000 levels in a single exposure.
 - ✓ For most epi-illuminated (fluorescence) samples, set the exposure time between 10 to 30 seconds to generate > 10,000 gray levels.
- 8 Set X and Y binning to None.
- 9 Select the Illumination Source (UV/Fluorescence or Transmission/Absorbance).
- 10 Record the *f*-stop and FOV settings in the Set Camera To section to match the corresponding settings on the camera lens.
- 11 Record any filters you may be using with the Filter pop-up menus.
- 12 Click the Expose button. A dialog box appears asking you to enter a name for the file (a default name will be entered using the selected illumination source, *f*-stop, and FOV settings).
- 13 Click OK. The file is saved.
- 14 The Illumination Reference File opens as a file and the IS4000R Acquire window appears. The newly created Illumination Reference File is now available to apply to a new image.
 - 👉 NOTE: Illumination Reference Files may also be applied later in software using the Image Field Correction from the Edit menu.
 - 👉 NOTE: Upon application, the illumination file does not alter the mean value of the image to which it is applied.
- 15 Turn off the light source.

Custom Capture Settings

You can speed up the capture process by creating custom capture settings for the various types of experiments or specific personal preferences. Once saved, KODAK MI remembers the camera settings which include:

- ✓ Exposure time
- ✓ Number of exposures
- ✓ Illumination settings
- ✓ f -Stop, field of view (FOV)
- ✓ Emission filter
- ✓ Image orientation
- ✓ Preferences
- ✓ Rotation
- ✓ Saturation levels
- ✓ Lens, warping, and illumination correction
- ✓ X and/or Y binning

 **IMPORTANT:** Settings will NOT save the User Name or Annotations.

Saving Custom Settings

- 1** Make all your selections from within the IS4000R Acquire window.
- 2** Choose the Save Settings from the Settings pop-up menu. A File Name dialog box appears.
- 3** Enter a name to describe your custom settings.
- 4** Click OK. The settings are saved and will appear in the list of available settings.

 **NOTE:** If you choose a Custom Acquire Setting, and then make modifications, they will not be saved unless the user chooses to update them manually by selecting Update Settings from the settings pop-up menu.

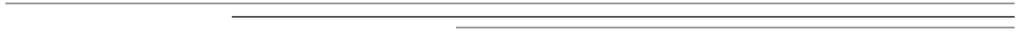
 **NOTE:** The Custom settings are files that are saved in the MI Extensions subfolder in the KODAK MI folder. To remove the Settings from the list, discard the file.

Updating Custom User Settings

- 1 Select a custom setting from the Settings pop-up menu.
- 2 Modify settings as desired.
- 3 Choose Update Settings in the Settings pop-up menu.

Deleting Custom User Settings

- 1 Select a custom setting from the Settings pop-up menu.
- 2 Choose Delete Settings.



Maintaining the System

The IS4000R Capture System Chamber is resistant to mild chemicals, water-tight, and therefore, easy to maintain. Periodically, some parts of the IS4000R may need to be replaced. This chapter reviews daily maintenance and user serviceable repairs.

Regular Maintenance

On a regular basis, the IS4000R requires little maintenance and is easy to clean. After each use, the Sample Work Area should be wiped down. Care is needed to avoid scratching the Platen. Clean the instrument with a soft lint-free cloth or lens paper.

Cleaning the Sample Work Area

- 1 Open the IS4000R lid.
- 2 Using an ammonia based spray cleaner, wipe the Platen, Sample Work Area and Compression Pad surfaces with a lint-free soft cloth or lens paper.
- 3 Be sure the Platen area is dry and without dust particles.

 NOTE: Always turn the UV light source off when not in use.



WARNING: Ammonia based spray cleaners may cause eye irritation. Consult the manufacturer's material safety data sheet for additional information prior to use.



WARNING: Do not use metal utensils or tweezers since they may scratch and permanently damage the Platen. This type of damage to the Platen is not covered by the warranty.



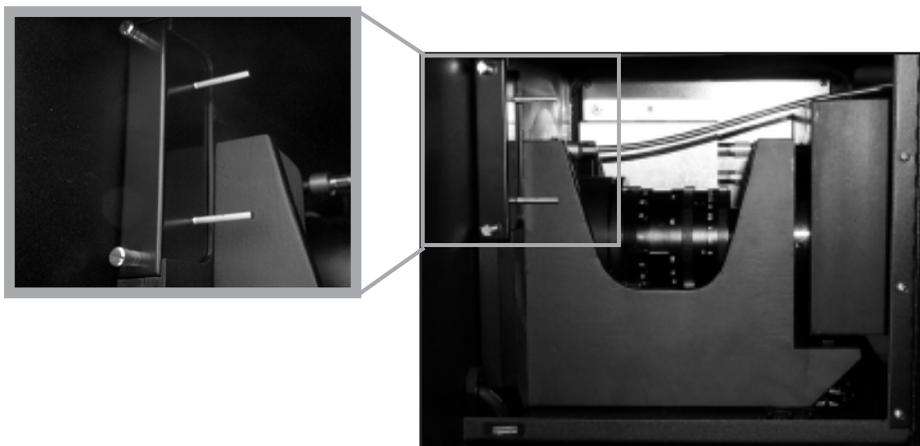
WARNING: Do not expose the Platen to strong chemical solvents such as ketones, hexanes, acids and alkalis that may permanently damage the Platen. This type of damage is not covered by the warranty.

Replacing or Changing an Emission Filter

Use the following procedure to replace or change an emission filter. The IS4000R is supplied with a 600 nm WB (ethidium bromide) filter. Specialized filters are available through Molecular Imaging Systems. Remember that the filter number that is indicated on the Filter Selection Dial does not correspond to the filter appearing at the Filter Access window as described in the chart below.

Changing or Replacing an Emission Filter

- 1 Open the Capture Chamber Door.
- 2 Loosen the two retaining screws and remove the Filter Access Panel.



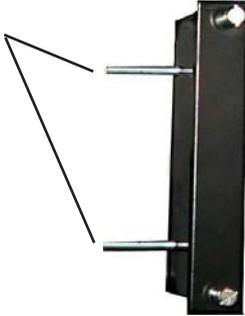
- 3 Rotate the Filter Selection Dial so that the desired filter slot is located in the Filter Access window.

 **NOTE:** The filter viewed in the Filter Access window is one position behind (counterclockwise) from the position indicated on the Filter Selection Dial.

Table 6.1: Filter Knob Position at Access Panel Relationship

Filter Knob Position	Filter at Access Window
0	3
1	0
2	1
3	2

-
-
- 4** To remove the Filter Cassette, insert the pins on the Filter Access Panel into the two holes in the Filter Cassette and withdraw the filter.



- 5** Carefully slide in the new Filter Cassette and align the “wings” on the Filter Cassette with the Filter Wheel. Be careful not to touch the filter element. The Filter Cassette will lock into place when it is fully inserted.

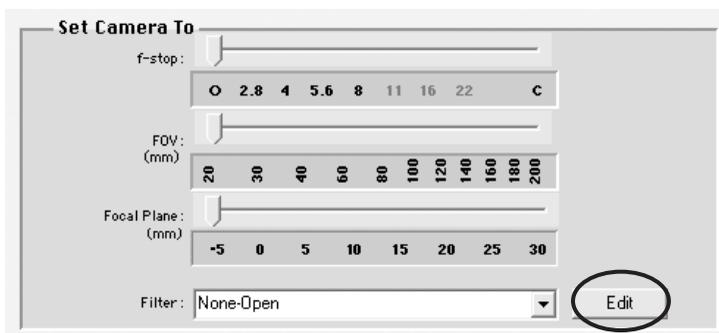
 **NOTE:** Do not force the filter. If the filter does not lock in place, remove the filter and realign before attempting to reinsert.

- 6** Replace the Filter Access Panel by aligning the pins in the holes and tightening the two (2) retaining screws.
- 7** Proceed to *Updating IS4000R Acquire Window*.

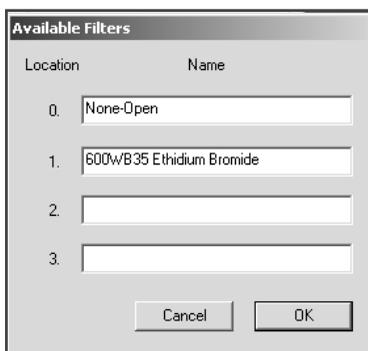
Updating the IS4000R Acquire Window

If you change or replace a filter, you will need to update the IS4000R Acquire window.

- 1 Launch KODAK Molecular Imaging Software by clicking on the KODAK MI icon found in the KODAK MI folder.
- 2 Choose the IS4000R Capture button or Select Digital Camera from the File menu and IS4000R from the Digital Camera pop-up menu.
- 3 The IS4000R Acquire window appears.
- 4 Choose the Edit button in the Set camera To section of the IS4000R window.



- 5 The Available Filters dialog box appears.



- 6 Edit the appropriate filter information in the Filter Location using the Filter text edit boxes.
- 7 Click OK to save the changes.

rad PADD Care and Maintenance

The rad PADD is a optional accessory. We recommend that you clean your rad PADD after each use. Care should be taken when handling the rad PADD cassette to avoid tearing the phosphor screen.

- 1 Open the rad PADD cassette and remove your sample.
- 2 Wearing latex or blue vinyl gloves, use a GM-type radioactivity meter calibrated in counts per minute (CPM) to determine the background readings on the rad PADD.
- 3 If any part of the rad PADD is found to show readings higher than background, wipe the area using Count Off™ (PerkinElmer Life Sciences) or another similar commercially available detergent and paper towels. If none are available, use Formula 409™ -like solution or a mild detergent.



 WARNING: Care should be taken to avoid tearing the phosphor screen.



 WARNING: If rad PADD has unremovable contamination after continued washing and the dpm/cm² remains constant above 100 of a short half-life isotope such as ³²P, the unit may be stored for 10 half-lives of both isotopic decay and the decontamination procedure repeated.

- 4 As you clean, discard liquid and solid waste (gloves and paper towels) according to your local and institutional regulations for radioactive material disposal. Continue washing until the GM meter reading for the contaminated area(s) is equal to or below background.



NOTE: It may be helpful to wrap your sample in plastic wrap when using wet samples on the rad PADD to minimize radioisotope contamination.

Transilluminator Lid Bulb Replacement

Tools Needed

- ✓ Image Station Transilluminator Lid Bulb Set
- ✓ 5/64 in. hex drive

- 1 Turn the lid illuminator power switch to the OFF position and unplug the power cord from the electrical outlet.
- 2 Open the lid to the full, upright position.
- 3 Using a 5/64 in. hex drive remove the 10 screws around the bulb cover.



- 4 Remove the bulb cover assembly.
- 5 The four bulbs are located at the perimeter of the light reflector. Rotate each bulb a 1/4 turn and remove. Always replace all 4 bulbs at the same time.



- 6 Replace the bulb cover, fasten the screws to secure.
- 7 Plug the power cord into a power receptacle.

Troubleshooting the System

In this chapter, common questions are addressed. The questions are divided into three sections: Instrument, Image, and Software. If you still have questions after reading this section and the corresponding information in the User's Guide, contact Technical Support. Please have your serial number and any technical information available.

Obtaining Technical Support

For technical support, contact Technical Support or your Molecular Imaging Systems dealer. For up to date dealer information, visit our WEB site at www.carestreamhealth.com/go/molecular. When contacting technical support, please have the following information available:

- ✓ The serial number of your IS4000R system located on the back of the unit and the camera serial number located on the camera.
- ✓ The serial number and version number of your software.
- ☞ NOTE: With KODAK MI running, select About KODAK MI under the Help menu (Windows) or select About KODAK MI under the Apple menu item (Macintosh).
- ✓ The type of computer you are using (make, model).
- ✓ Operating system software version.
- ☞ NOTE: Check your operating system version by right-clicking on the My Computer icon and then on Properties (Windows) or select About This Mac under the Apple menu item (Macintosh).
- ✓ The type of image you are capturing or analyzing.
- ✓ The problem you are having and what you were doing when the problem occurred. Please note the exact wording of any error messages, including any error numbers displayed.

Molecular Imaging Systems Technical Support

Contact Molecular Imaging Systems Technical Support by:

- ✓ Utilizing our World Wide Web support pages at:
<http://www.carestreamhealth.com/go/molecular>
- ✓ Calling Molecular Imaging Systems Technical Support at:
877-747-4357 or (203) 786-5657, between the hours of 8:00 a.m. and 6:00 p.m. (Eastern Standard Time) Monday through Friday
- ✓ E-mailing Molecular Imaging Systems Technical Support at:
molecular-support@kodak.com
- ✓ Faxing Molecular Imaging Systems Technical Support at:
203-786-5656

Common Instrument Questions

Problem	Probable Cause	Solution
Cannot access the IS4000R capture system.	FireWire Camera/Computer Interface Cable is not properly connected.	Check the FireWire connections on the back of the IS4000R and the computer.
	The FireWire cable that came with the IS4000R was not used.	Make sure that the cable being used is the one you received with your IS4000R.
		Check the IEEE 1394 (FireWire) Camera/Computer Interface Cable.
	No power to the camera or power supply failure.	Verify green LED on top of camera is lit. Check the Power Switch/Circuit Breaker.
	Computer was powered prior to powering the camera.	Shut down computer, turn the instrument's Power Switch to the ON position and then restart the computer.
	Camera drivers not loaded properly.	
Temperature Error displayed when trying to acquire images.	The camera is not cooled to temperature.	Wait 30 minutes after turning the camera on. We recommend always keeping the camera switch in the ON position.
		From the IS4000R Acquire window, press the Control or Ctrl and T keys simultaneously to view the CCD temperature. If the temperature is higher than minus 23° C, contact Technical Support.
		Check the environmental conditions required to operate the instrument. Requires 6 inches (15.2 cm) of clearance from wall or other objects. See Chapter 8: <i>Installing the System</i> .
UV light source not working.	No power to the UV light sources.	Check that the UV light source switch is turned on.
		Check that the power cord is properly attached and that the outlet is supplying power.
		Check that the circuit breaker switch on the back panel is turned to on.
	UV light source is faulty.	Contact Technical Support.

Common Image Questions

Problem	Probable Cause	Solution
Need a new filter.		Follow the procedure in <i>Replacing and Changing an Emission Filter</i> in Chapter 6: <i>Maintaining the System</i> .
Image is black or too dark.	Not enough light for a good contrast exposure.	Open the f -stop to O (Open) or increase the exposure time. See Chapter 4: <i>Setting up for Capture</i> .
Image is white or too light.	Too much light during the exposure.	Close down the f -stop (larger number for less light) and/or use the Light Diffuser for transmitted light samples. Do not use a f -stop larger than $f8$. Rotate Attenuator dial/knob to decrease output. See Chapter 4: <i>Setting up for Capture</i> .
Image is blurred.	Image is not in focus.	For thin samples, set the focus on or around the white mark on the Focus Ring. For thicker samples, adjust the Focus Ring and/or increase the depth of field by increasing the f -stop setting. Use the Focusing tool in the Preview mode to optimize.
	Dirty or cracked filter.	Capture an image with a different filter selected to verify cause; replace filter.
	Contaminated optics.	Contact Technical Support.
	Condensation on the CCD window from running at low temperatures or high dew point.	Leave the camera power on at all times. The condensation forms from fast cool down in less than ideal environmental conditions. If camera is left on, the condensation will dissipate and does not reoccur unless the camera is cycled on and off.
	Lens came loose.	Call Technical Support. See <i>Obtaining Technical Support</i> in Chapter 1: <i>Introduction</i> .

Problem	Probable Cause	Solution
Bright imaging artifacts not coming from the sample.	Light leak.	Check that the Filter Access Panel is properly in place.
		Ensure the lid has been closed and latched in place.
		Ensure the Lid Gasket is clean and free of debris.
		Verify by performing a 5 minute exposure with the lid closed, X and Y binning (8 x 8) selected, no illumination, and no sample. Save the image.
		Expose an additional image with the same settings, but this time place the Light Diffuser over the Platen, remove the Compression Pad and close the lid. If significant increase in artifact signal is seen, the light leak is due to a faulty Lid Gasket. Call Technical Support.
If a signal artifact continues and is not changed in the above procedure, call Technical Support to diagnose the leak and to obtain further instructions.		
Image appears with many speckles in an UV illuminated image.	Dust or dirt on the Platen.	See <i>Preparing the IS4000R for Imaging</i> in Chapter 4: <i>Setting up for Capture</i> .
	Camera correction files are incorrect or not installed.	Verify that the camera CD matches your camera serial number.
Image partially blocked.	Filter not centered over the lens properly.	Check that the Filter Selection Dial is lined up with the filter selection properly. View the Filter Wheel alignment by looking down into the Platen.

Problem	Probable Cause	Solution
Lines in the image.	Scratched Platen.	Contact Technical Support for further instructions.
	Image transfer was interrupted	Check cables and connections.
Upon opening image, the background illumination in the image is uneven.	Improper or no Illumination Reference File was used for correction.	Be sure the proper Illumination Reference File is used.
The image is not the correct size.	<i>f</i> -stop and FOV settings recorded in Set Camera To dialog box are not the same as those used to capture the image.	Be sure the proper <i>f</i> -stop and FOV inputs are used.
Dark center (circle) on brightfield image.	Condensation in window.	Leave the camera power on at all times. The condensation forms from fast cool down in less than ideal environmental conditions. If camera is left on, the condensation will dissipate and does not reoccur unless the camera is cycled on and off.
Smear in image not related to sample that does not change with settings.	Contaminated optics.	Requires service. Contact Technical Support.
Cannot detect radioactive signal.	Not using the rad PADD accessory.	To image radioactive signal, you will need to use the optional rad PADD accessory, contact Molecular Imaging Systems for ordering information.
	Sample is not in the proper orientation.	If your object is opaque (blot), orient the sample side downward—if translucent (gel), orient the sample either way.
	No radioactive label on sample.	Use a GM-type radioactivity meter calibrated in counts per minute (CPM) to check for activity.
	Wrong radioisotope.	The rad PADD and IS4000R are design to detect high-energy beta-emitters such as ³² P, or ¹²⁵ I.

Common Software Questions

Problem	Probable Cause	Solution
Copy protection error.	The copy protection device is not responding.	Check the connections and restart the software and computer. Refer to the KODAK Molecular Imaging Software User's Guide.
Error communicating with camera.	System/Display sleep mode set incorrectly.	Windows: right-click on the desktop and select Properties. Select the Screen Saver tab. Click on the Power tab. Click on the Power Scheme tab (Power Schemes should be Always On and settings should be set to Never.
No Illumination Reference File available in the pop-up menu.	Capture settings do not match those in the reference file.	Create Illumination Reference File to match current capture settings.

Installing the System

This section details the installation of the IS4000R system. This information will be used by your local representative when your equipment is initially installed. Should you need to install the IS4000R system, this section will tell you how to unpack the equipment and what to consider in selecting a new location for the equipment. In addition, it contains instrument performance checks, necessary after installation, and how to repackage and prepare the equipment for a move.

Environmental Requirements

The IS4000R system is designed to operate effectively within the temperature and humidity ranges typically found in laboratories. For effective operation, the temperature and relative humidity should be:

- ✓ Temperature: 17° to 27° C
- ✓ Relative Humidity: < 75%, non-condensing
- ✓ Altitude: This product is designed for use at altitudes up to 2000 meters.

This product is designed for indoor use only. This product meets Pollution Degree 2 standards in accordance with IEC 664.

Electrical Requirements

- ✓ 100–240 VAC*, 50/60Hz, 1A

* Supply voltage fluctuations should not to exceed $\pm 10\%$.

Space Requirements

The IS4000R and computer must be placed on a bench or table that is level and capable of supporting 250 pounds (118 kg), located not more than 3 feet (91 cm) from an electrical outlet.

The IS4000R system, including computer CPU, monitor and keyboard, requires a minimum space 69 inches wide, 24 inches deep and 26 inches high (175 x 61 x 66 cm). These dimensions do not allow for a printer or other peripheral device.



WARNING: Maintain a minimum of 6 inches (~15 cm) clearance around the Radiator to allow for proper cooling.



NOTE: Additional space adjacent to, above or behind the IS4000R will allow for easier operation.

This product is designed to withstand transient overvoltage according to Installation Category II.

Refer to your computer operators's manual for its electrical requirements.

Installing the IS4000R

Setting Up the IS4000R Hardware

- 1 Remove the instrument from the shipping container. Remove packing materials.
 -  NOTE: Retain the box and packing material in case you need to repackage and move the system or return the system for repair.
- 2 Verify components using the list in *What's in the IS4000R Package* in Chapter 1: *Introduction* of this User's Guide.
- 3 Position the IS4000R next to the computer.
 -  IMPORTANT: Do not attach the IS4000R to the computer until after KODAK Molecular Imaging Software has been installed.
- 4 Install your Emission filters. Refer to *Replacing or Changing an Emission Filter* in Chapter 6: *Maintaining the System*.
- 5 Proceed to *Installing KODAK Software—Windows or Installing KODAK Software—Macintosh*

Installing the IEEE 1394 (FireWire) Card

If your computer is not equipped with IEEE 1394 (FireWire) ports, contact your computer manufacturer for assistance in obtaining an add-on IEEE 1394 (FireWire) card. The IEEE 1394 (FireWire) card enables your computer to communicate with your camera. Follow the manufacturer's directions for installation of the IEEE 1394 (FireWire) card.



WARNING: The card is stored in an antistatic bag because it is sensitive to static electricity. Be careful when you handle the card.



WARNING: Refer to your computer user's manual for information on opening your computer case. Follow the installation instructions for the IEEE 1394 (FireWire) card and software drivers that are enclosed with your IEEE 1394 (FireWire) Add-on card.

Proceed to *Installing KODAK Software—Windows or Installing KODAK Software—Macintosh*.

Installing the KODAK Image Station Transilluminator Lid (Optional Accessory)

The Image Station Transilluminator Lid Accessory is an optional accessory. The lid accessory is easily installed by replacing your standard IS4000R Lid.



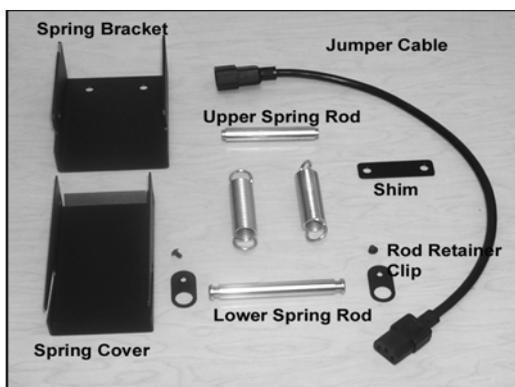
WARNING:DANGER—Ultraviolet radiation emitted from this product. Avoid exposure. **ALWAYS WEAR PROTECTIVE CLOTHING. EXPOSURE MAY CAUSE PREMATURE AGING OF THE SKIN AND CANCER. ALWAYS WEAR PROTECTIVE EYEWEAR; FAILURE TO DO SO MAY RESULT IN SEVERE BURNS OR LONG TERM INJURY TO EYE.** Never look directly into the lamp. In individuals with a photosensitivity disease, exposure can cause a wide variety of effects, not just allergenicity. Some medications, cosmetics or diseases may increase your sensitivity to ultraviolet radiation. Consult a physician before operating this product if you are using medications or have a history of skin problems or believe yourself especially sensitive to sunlight. For Laboratory Use Only.

Package Contents

When unpacking your Image Station Transilluminator Lid Accessory, please take a moment to ensure that all the necessary parts have been received.

Unpack and carefully examine all parts. A slight rattling sound in the unit is normal from the movement of small particles that are present inside the fluorescent bulbs. Your package should contain:

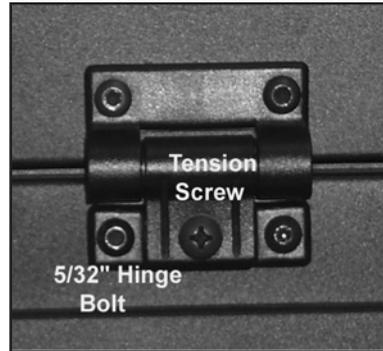
- ✓ Transilluminator Lid (1)
- ✓ Rod Retainer Clip and Screw (2)
- ✓ Spring Bracket (1)
- ✓ Spring Cover (1)
- ✓ Upper Spring Rod (1)
- ✓ Springs (2) (not shown)
- ✓ Lower Spring Rod (1)
- ✓ Jumper Cable (1)
- ✓ Shim (1)
- ✓ White Light PADD (not shown)
- ✓ Lid Liner (not shown)



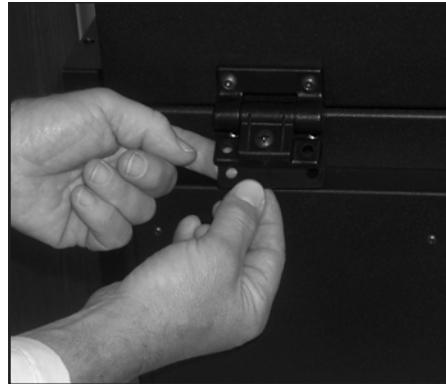
Tools Needed

- ✓ #2 Phillips Screwdriver
- ✓ 5/32 in. hex drive

- 1 Shut down the computer that is attached to your IS4000R.
- 2 Turn the power to the IS4000R off and remove the power cord from the wall and from the unit.
- 3 Disconnect the IS4000R from your computer.
- 4 Position the IS4000R to allow access to front and rear of the unit, and so the lid may be fully opened.
- 5 Facing the rear, remove the lower bolts on each hinge using a 5/32 in. hex drive. Save the bolts for installation of the new lid.



- 6 Remove the lid from the IS4000R cabinet.
- 7 Place the Lid Illuminator on the Image Station.
- 8 Place a Shim under the left hinge and fasten loosely with 2 bolts using a 5/32 in. hex drive.



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- 9** Attach the Spring Bracket under the right-side hinge and fasten tightly to the cabinet with 2 hinge bolts using a 5/32 in. hex drive.

 **NOTE:** To ensure proper alignment, tilt the spring bracket so the top left edge fits snug under the safety switch

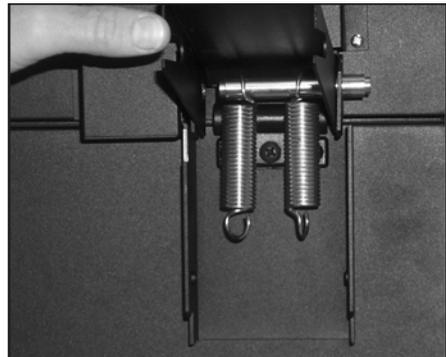
- 10** With a #2 Phillips screwdriver tighten the hinge tension screws on both hinges.
- 11** Slide the Upper Spring Rod through the right holes of the Spring Bracket and Spring Cover.



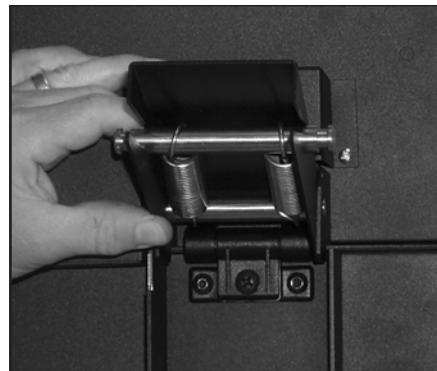
- 12** Slide the Upper Spring Rod through the left holes in the Spring Bracket and Spring Cover.
- 13** Slide the Springs over the Upper Spring Rod.

- 14** Adjust the Upper Spring Rod to fit flush on each side of the Spring Bracket.

 **NOTE:** Upper Spring Rod must be centered in the Spring Bracket to provide proper balance and tension.



- 15** Slide the Lower Spring Rod through the Spring Cover and Springs.
- 16** Raise the lid so it is perpendicular to the Sample Work Area on the cabinet allowing the Lower Spring Rod to reach the Lower Spring Bracket.

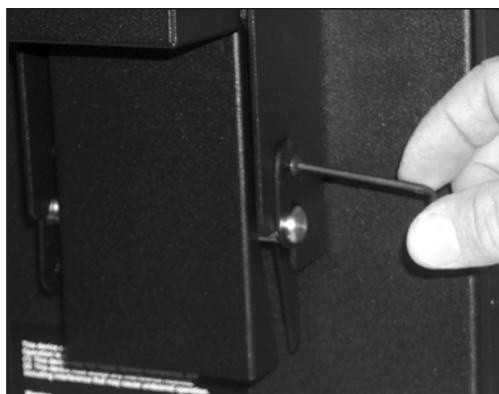


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- 17** Position the Spring slightly forcing the Lower Spring Rod under the Spring Bracket, seating the groove at each end of the rod in the slot on both sides of the Spring Bracket.



- 18** Position the Rod Retainer Clips over the lower spring rod and fasten with a #6 screw through the upper hole into the Spring Bracket with a hex drive.

-  **NOTE:** After installation of the hinge according to these directions, the Safety Shut-Off lever on the hinge should function in a way as to allow the lights to go on when the lid is closed and then off when the lid opens.



- 19** Using the Jumper Cable, plug the Lid Illuminator into the IS4000R cabinet.
- 20** Plug the Lid Illuminator in using the Image Station line cord.
- 21** Verify proper activation of UV lights by taking a 2-second exposure with the lid closed and no filter in place. The image should demonstrate light intensity significantly above the image without lights powered.
-  **NOTE:** The Auto Shut-Off Switch activates the automatic UV light shut-off. When the switch is in the on position, the UV Transilluminator will automatically turn off after 15 minutes.
- 22** Close and latch the lid. With the UV lights off, the lens aperture to f_0 , take a 5-minute, binning 8 x 8 exposure and verify that the image is dark.
- 23** Your installation is complete, proceed to *Launching KODAK Molecular Imaging Software for the First Time*.

Installing KODAK Software—Windows

Any prior versions of KODAK and/or camera software that are loaded on the computer must be uninstalled prior to installation. If you are a new user, proceed to *KODAK Molecular Imaging Software Installation—Windows*. Windows 2000 and Windows XP installations may require administrator privileges.

If you have purchased the KODAK Molecular Imaging Software Network Edition (KODAK MI NE) to use with your Image Station 4000R:

- ✓ Follow the instructions in the KODAK Molecular Imaging Software Network Edition Administrator's Manual to install KODAK MI NE Software.
- ✓ Then proceed to *KODAK Image Station 4000R Camera Files Installation—Windows* and *Connecting the KODAK Image Station 4000R to a IEEE 1394 (FireWire) Port—Windows*, later in this chapter.

If you have purchased the KODAK Molecular Imaging Software, Regulatory Edition with Network Licensing (KODAK MI RE) to use with your Image Station 4000R:

- ✓ Follow the instructions in the KODAK Molecular Imaging Software, Regulatory Edition Network Administrator's Manual to install KODAK MI RE Software.
- ✓ Then proceed to *KODAK Image Station 4000R Camera Files Installation—Windows* and *Connecting the KODAK Image Station 4000R to a IEEE 1394 (FireWire) Port—Windows*, later in this chapter.

Uninstalling a Previous Version of KODAK MI or KODAK 1D Software—Windows

1 Remove your KODAK MI or 1D copy protection device from your computer.

2 Inactivate any virus protection software.

 NOTE: Norton Utilities and Norton Antivirus software must be deactivated before you uninstall the KODAK Software. The software might not uninstall properly with the virus protection left running. After installation, you can restart your virus protection software.

3 Close all software applications that may be running on your computer.

 NOTE: KODAK cameras should not be connected to the computer while uninstalling the software.

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- 4 Move any customized standards or templates and any projects from their respective subfolders in your existing KODAK MI or KODAK 1D X.X folder to a temporary folder outside the Kodak folder. If you are currently using the KODAK MI database, also move the database folder. The database folder labeled GMPDB is located in the C:\Programs Files\KODAK MI\KodakAdminPortal folder.
 - 5 Uninstall previous version(s) of KODAK MI or KODAK 1D Software.
 - ✓ Uninstall KODAK MI by choosing Control Panel from the Start menu and Add/Remove Programs. Scroll to locate Kodak MI 4.X and click the Remove button. You can also uninstall the application using your KODAK MI CD. The installer shield automatically detects that you have the software loaded and offers an uninstall option.
 - ✓ Uninstall KODAK 1D by choosing Programs from the Start menu and selecting Remove KODAK 1D X.X from the KODAK 1D X.X submenu.
 - ✓ Uninstall KODAK MI NE or KODAK 1D NE following the instructions provided in your Network Administrator's Manual.
 - 6 Uninstall additional KODAK Software by choosing Control Panel from the Start menu and Add/Remove Programs. Scroll to locate Sentinel System Driver. Click to select.
 - 7 Click the Add/Remove button and follow the on-screen instructions to uninstall.
 -  NOTE: Automated uninstall features on some systems may not remove all previous program elements. Check drives for residual folders and files. Manual deletion of these folders and files may be necessary.
 - 8 Restart your computer.
 - 9 Proceed to *KODAK Molecular Imaging Software Installation—Windows*.

KODAK Molecular Imaging Software Installation—Windows

KODAK Molecular Imaging Software is installed like most Windows application programs and requires administrator privileges.

- 1** Inactivate any virus protection software.
 -  **NOTE:** Norton Utilities and Norton Antivirus Software must be deactivated before you install the KODAK Software. The installation might not run properly with the virus protection left running. After installation, you can restart your virus protection software.
- 2** Close all software applications that may be running on your computer.
 -  **NOTE:** Kodak cameras should not be connected to the computer while installing the software.
 -  **NOTE:** Remove any copy protection devices attached to your computer during installation.
- 3** Insert the KODAK Molecular Imaging Software Version 4.X CD into your CD drive and double-click on the KODAK MI Installer.exe icon to launch the installer.
- 4** The installer leads you through the installation process. Make sure to select the Complete Setup to install the software in the default directory to ensure full functionality of the system.
 -  **NOTE:** The Select Components window allows you to install MI without the database. Only install the database in the location that you want to maintain images acquired from the system.
- 5** While the software installation is occurring, complete your KODAK Software Registration Card and mail the card. This ensures that you receive information on new software releases, periodic maintenance releases, and technical bulletins.
- 6** A dialog box appears when the installation is complete. Select the option to restart the computer and click Finish.
- 7** Proceed to *Copy Protection Device Installation*.

Copy Protection Device Installation

KODAK MI Software is copy protected using a device that plugs into the USB port of your computer. It will not launch unless this device is attached to your computer. If you are installing the system for the first time, locate and install the copy protection device according to the instructions below. If you are upgrading your system, your package may or may not contain a new copy protection device.

To install the copy protection device:

- 1 Plug the copy protection device into a USB port of your computer. Please make sure that the connection is secure.
 - NOTE: Windows XP Users: A Welcome to the Found New Found Hardware Wizard dialog box appears on screen. You will be asked if you want the Windows update to search for software. Click No not at this time and advance through the installation wizard.
 - NOTE: If you are upgrading from a previous version of the KODAK MI or 1D Software and received a new copy protection device, you must attach both the old and new copy protection devices to your computer. After you launch the newest version for the first time, your old copy protection device is deactivated. Remove and discard the old key.
 - NOTE: If your computer has multiple USB ports, you can plug the copy protection device into any of them. The software, when launched, checks all available USB ports.
- 2 Proceed to *Setting Windows Power Settings*.

Setting Windows Power Settings

- 1 KODAK Imaging Systems require that your computer Energy Saver features be disabled.
- 2 Click the Power Options icon from the Start menu and the Control Panel.
- 3 Click Power Schemes tab and select Always On from the Power Schemes pop-up menu.
- 4 The settings for Turn off monitor, Turn off hard disks, System standby and System hibernates functions become *Never*.
- 5 *Proceed to Image Station 4000R Camera Files Installation—Windows.*

Image Station 4000R Camera Files Installation—Windows

- 1** Inactivate any virus protection software.

 **NOTE:** Norton Utilities and Norton Antivirus Software must be deactivated before you install the KODAK Software. The installation might not run properly with the virus protection left active. After installation, you can restart your virus protection software.

- 2** Insert the KODAK Image Station Camera Files CD into your CD drive and double-click the IS4000 Support Setup.exe icon to launch the Windows installer application.

 **NOTE:** KODAK MI Software must be installed prior to the camera files installation.

 **NOTE:** If you received your Image Station with KODAK 1D Image Analysis Software you will have to move the IS4000_HMPS and IS4000_DCRS folders from the C:\Program Files\KODAK 1D 3.6\IS4000 to C:\Program Files\KODAK MI\MI Application\IS4000. If you no longer have the files installed, copy the files from your KODAK IS4000 Camera Files CD.

- 3** The installer will proceed and display a dialog box when installation is complete.
- 4** Verify that the appropriate KODAK IS4000_HMPS and IS4000_DCRS files have been installed in the proper location. Navigate to C:\Program Files\KODAK MI\MI Application IS4000.
- 5** Click OK and proceed to *Connecting the KODAK Image Station 4000R to Your IEEE 1394 (FireWire) Port—Windows*.

Connecting the Image Station 4000R to Your IEEE 1394 (FireWire) Port—Windows

Transferring images via IEEE 1394 (FireWire) port is much faster than either serial or USB port connections. IS4000R requires that your computer is equipped with an IEEE 1394 port or an IEEE 1394 PCMCIA adapter (notebook users).

Once you have installed KODAK MI Software and your copy protection device, you can connect the IS4000R to your computer.



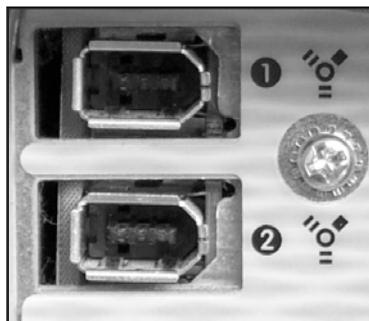
WARNING: Your computer should be turned off when connecting your camera to your computer.

- 1 Connect the IEEE 1394 (FireWire) Camera/Computer Interface Cable to the connector at the rear of the IS4000R. The smaller end of the connector should be facing down when connecting to the instruments port.



WARNING: Be sure to insert the IEEE 1394 (FireWire) Camera/Cable Interface Cable in the correct orientation. Incorrect placement will permanently damage your camera circuitry and will void the warranty.

- 2 Connect the other end of the IEEE 1394 (FireWire) Camera/Computer Interface Cable to the IEEE 1394 (FireWire) port of your computer. Refer to the figure to properly align the plug with the port.



- 3 Attach the AC line cord to the power input module at rear of the instrument and plug into an appropriate AC (MAINS) outlet.



WARNING: Select the appropriate modular plug to match your local AC (MAINS) outlet.

4 Turn the circuit breaker (located on the back of the unit) on.

5 Turn the power switch (located on the front of the unit) on.

6 Start up your computer.

 **NOTE:** WINDOWS: A Found New Hardware Wizard dialog box appears on screen. Click Next to advance through the installation wizard.

 **NOTE:** A Welcome to the Found New Found Hardware Wizard dialog box may appear on screen. You will be asked if you want the windows update to search for software. Click No not at this time and advance through the installation wizard.

 **NOTE:** During the installation, a dialog box will appear stating *Digital Signature Not Found*. Click Yes to continue the installation.

7 Click Finish when the wizard is complete.

8 Proceed to *Launching the Kodak Molecular Imaging Software for the First Time*, later in this chapter.

Installing KODAK Software—Macintosh

Any prior versions of KODAK and/or camera software that are loaded on the computer must be uninstalled prior to installation. If you are a new user, proceed to *KODAK Molecular Imaging Software Installation—Macintosh*. Macintosh OS X installations may require authentication of permissions.

If you have purchased the KODAK Molecular Imaging Software Network Edition (KODAK MI NE) to use with your Image Station 4000R:

- ✓ Follow the instructions in the KODAK Molecular Imaging Software Network Edition Administrator's Manual to install KODAK MI NE Software.
- ✓ Then proceed to *KODAK IS4000R Camera Files Installation—Macintosh* and *Connecting the KODAK Image Station 4000R to a IEEE 1394 (FireWire) Port—Macintosh*, later in this chapter.

If you have purchased the KODAK Molecular Imaging Software, Regulatory Edition with Network Licensing (KODAK MI RE) to use with your Image Station 4000R:

- ✓ Follow the instructions in the KODAK Molecular Imaging Software, Regulatory Edition Network Administrator's Manual to install KODAK MI RE Software.
- ✓ Then proceed to *KODAK IS4000R Camera Files Installation—Macintosh* and *Connecting the KODAK Image Station 4000R to a IEEE 1394 (FireWire) Port—Macintosh*, later in this chapter.

Uninstalling a Previous Version of KODAK MI or KODAK 1D Software—Macintosh

- 1 Remove your KODAK MI or 1D copy protection device from your computer.
- 2 Inactivate any virus protection software.
 - ☞ NOTE: Norton Utilities and Norton Antivirus software must be deactivated before you uninstall the KODAK Software. The software might not uninstall properly with the virus protection left running. After installation, you can restart your virus protection software.
- 3 Close all software applications that may be running on your computer.
 - ☞ NOTE: KODAK cameras should not be connected to the computer while uninstalling the software.

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- 4** Move any customized standards or templates and any projects from their respective subfolders in your existing Kodak MI 4.X or KODAK 1D X.X folder to a temporary folder outside the Kodak folder. If you are currently using the KODAK MI database, also move the database folder. Click the Applications folder and locate the database folder labeled GMPDB located in the KODAK MI folder KodakAdminPortal subfolder.
 - 5** Place the Kodak MI 4.X or KODAK 1D X.X folder in the Trash.
 - 6** Restart your computer.
 - 7** Empty the Trash.
 - 8** Proceed to *KODAK Molecular Imaging Software Installation—Macintosh*.

KODAK Molecular Imaging Software Installation—Macintosh

1 Inactivate any virus protection software.

 NOTE: Norton Utilities and Norton Antivirus Software must be deactivated before you install the KODAK Software. The installation might not run properly with the virus protection left running. After installation, you can restart your virus protection software.

2 Close all software applications that might be running on your computer.

 NOTE: KODAK cameras should not be connected to the computer while installing the software.

 NOTE: Remove any copy protection devices attached to your computer during installation.

3 Insert the KODAK Molecular Imaging Software Version 4.X CD into your CD drive and double-click the KODAK MI.mpkg icon to launch the installer application.

4 The installer leads you through the installation process. Make sure to select the Easy Install to install the software in the default directory and ensure full functionality of the system.

 NOTE: The Select Components window allows you to install MI without the database. Only install the database in the location that you want to maintain images acquired from the system.

5 While the software installation is occurring, complete your KODAK MI Software Registration Card and return the card by mail. This process takes only a few minutes to complete and ensures that you receive information regarding new software releases, periodic maintenance releases, and technical bulletins.

6 The installer notifies you when the installation is complete. Click Restart.

7 Proceed to *Copy Protection Device Installation*

Copy Protection Device Installation

KODAK MI Software is copy protected using a device that plugs into the USB port of your computer. KODAK MI will not launch unless this device is attached to your computer. If you are installing the system for the first time, locate and install the copy protection device according to the instructions below. If you are upgrading your system, your package may or may not contain a new copy protection device.

To install the copy protection device:

- 1** Plug the copy protection device into a USB port of your computer. Please make sure that the connection is secure.

 **NOTE:** If you are upgrading from a previous version of the KODAK MI or KODAK 1D Software and received a new copy protection device, you must attach both the old and new copy protection devices to your computer. After you launch the newest version for the first time, your old copy protection device is deactivated. Remove and discard the old key.

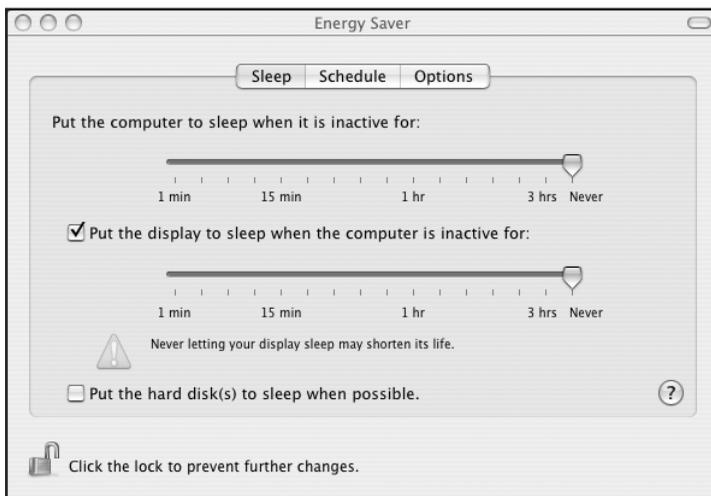
- 2** Proceed to *Macintosh Power Settings*.

Macintosh Power Settings

KODAK Imaging Systems require that the Energy Saver settings on your computer be disabled.

To disable the system sleep features

- 1 Choose Energy Saver from the File menu and the System Preferences submenu or from System Preferences on the Dock. The Energy Saver window opens.



- 2 Click the Sleep tab and set the Computer and Display sleep sliders to Never. Ensure the Put the hard disk(s) to sleep when possible checkbox is unchecked.
- 3 Proceed to *Image Station 4000R Camera File Installation—Macintosh*.

Image Station 4000R Camera File Installation—Macintosh

- 1 Insert the KODAK IS4000 Camera Files CD into your CD drive and double-click the Camera Files Installer to launch the installer application.

 NOTE: KODAK MI Software must be installed prior to the camera files installation.

 NOTE: If you received your Image Station with KODAK 1D Image Analysis Software you will have to move the IS4000_HPMS and IS4000_DCRS folders manually. Click the Application Folder and navigate to the KODAK 1D 3.6 and the IS4000 folder. Copy the files to the KODAK MI\MI Extensions\IS4000 folder. If you no longer have the files installed, copy the files from your KODAK IS4000 Camera Files CD.

 NOTE: In a manual copy operation, the HPMS and DCRS folders must have their Ownership & Permissions reset in OS X to allow access by other user accounts. After copying, select the folders one at a time and click Get Info from the File menu. Set the Others pop-up menu in the Ownership & Permissions section to Read & Write. Click Apply.

- 2 The installer will proceed and display a dialog boxes when installation is complete.
- 3 Verify that the the IS4000_HPMS and IS4000_DCRS files have been installed in the proper location Applications\KODAK MI\MI Extensions\IS4000.
- 4 Your installation is complete, proceed to *Connecting the Image Station 4000R to Your IEEE 1394 (FireWire) Port—Macintosh*.

Connecting the Image Station 4000R to Your IEEE 1394 (FireWire) Port—Macintosh

Transferring images via IEEE 1394 (FireWire) port is much faster than either serial or USB port connections. IS4000R requires that your computer is equipped with an IEEE 1394 port or an IEEE 1394 PCMCIA adapter (notebook users).

Once you have installed KODAK MI Software and your copy protection device, you can connect the IS4000R to your computer.



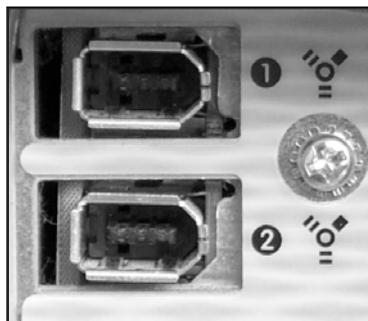
WARNING: Your computer should be turned off when connecting your camera to your computer.

- 1 Connect the IEEE 1394 (FireWire) Camera/Computer Interface Cable to the connector at the rear of the IS4000R. The smaller end of the connector should be facing down when connecting to the instruments port.



WARNING: Be sure to insert the IEEE 1394 (FireWire) Camera/Cable Interface Cable in the correct orientation. Incorrect placement will permanently damage your camera circuitry and will void the warranty.

- 2 Connect the other end of the IEEE 1394 (FireWire) Camera/Computer Interface Cable to the IEEE 1394 (FireWire) port of your computer. Refer to the figure to properly align the plug with the port.



- 3 Attach the AC line cord to the power input module at rear of the instrument and plug into an appropriate AC (MAINS) outlet.



WARNING: Select the appropriate modular plug to match your local AC (MAINS) outlet.

- 4** Turn the circuit breaker on (located on the back of the unit).
- 5** Turn the power switch (located on the front of the unit) on.
- 6** Start up your computer.
- 7** Proceed to *Launching the KODAK Molecular Imaging Software for the First Time*, later in this chapter.

Launching KODAK Molecular Imaging Software for the First Time

1 Launch KODAK Molecular Imaging Software.

- ✓ Windows Users—Click Programs from the Start menu and select KODAK MI.exe from the Kodak MI submenu or by double-clicking on the KODAK MI.exe icon found in the C:\Program Files\KODAK MI\MI Application folder.

☞ NOTE: Consider placing a Shortcut to the KODAK MI.exe file on your desktop. Navigate to the KODAK MI.exe file as shown above. Right-click on the KODAK MI.exe icon and choose Create Shortcut. A Shortcut is created in the MI Application folder. Drag the shortcut to the desktop.

- ✓ Macintosh Users—Select Applications from the Go menu and click the KODAK MI icon found in the KODAK MI folder and the MI Application subfolder.

☞ NOTE: Consider placing an Alias to the KODAK MI executable program file on your desktop. To create an Alias, single-click the KODAK MI icon and choose Make Alias from the File menu. An Alias is created in the MI Application folder. Drag the Alias onto the desktop. Alternately, drag the KODAK MI icon to the Dock to create an Alias on the Dock.

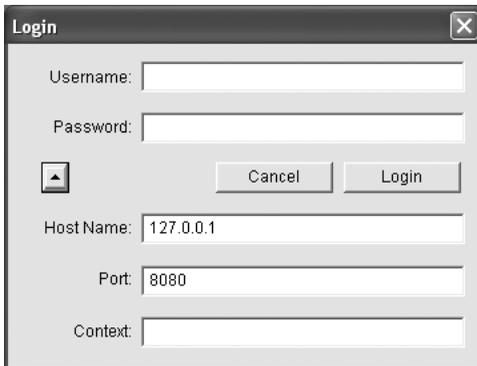
- ✓ KODAK MI NE Users—Click Programs from the Start menu and select KODAK MI 4.X Network.exe from the KODAK MI submenu or by double-clicking on the KODAK MI 4.X Network.exe icon found in the C:/Program Files/KODAK MI 4.X Network/MI Application folder (Windows) or Select Applications from the Go menu and click the Kodak MI 4.X Network icon found in the Kodak MI 4.X folder and the MI Application subfolder (Macintosh).

2 The KODAK MI Security Setup dialog box appears. You must select either *No Login* or *Login Required* from the pop-up menu. Click Continue.



- ☞ NOTE: The selected KODAK MI Security feature applies to your new KODAK MI installation for all users.

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- 3** The Login window appears. Enter “Admin” as the Username and “password” as the Password. The Host Name default value will cause KODAK MI to search your local drive for the KODAK MI database. If the KODAK MI database is located on another machine or file server, enter the IP address of that machine in the Host Name field. Click Login.



-  **NOTE:** The initial installation of KODAK MI Software comes with a single user with the User Name “Admin” and the Password “password”. Use this User name to gain access to KODAK MI Security Manager to set up users.
-  **NOTE:** If no KODAK MI database is detected at the Host Name location, you will be asked if you would like to continue without connection to a database.
- 4** If you selected No Login in the KODAK MI Security Setup dialog box, future launches of KODAK MI will skip the login windows and open to a MI Project window. Proceed to Step 6 to enter your User Name, Company and KODAK MI Software serial number.
- 5** If you selected Login Required in the KODAK MI Security Setup dialog box, you will be prompted to contact database administrator. Click OK to open KODAK MI. Future launches of KODAK MI will require both Username and Password to gain access to KODAK MI software and the KODAK MI database.
- 6** Type your Name and Organization in the boxes provided.
- 7** Enter your Serial Number exactly (including dashes) as provided on your registration card or serial number card included in your KODAK MI package.
-  **NOTE:** The serial number is required when installing KODAK MI or when contacting Technical Support. Keep the serial number in a safe location.

8 Click OK. The MI Project window appears. Your installation is complete.

 **NOTE:** Previous KODAK MI or 1D users may move any customized standards or templates, any projects, and database folders back into the new KODAK MI folder.

 **NOTE:** If you selected Login Required in the KODAK MI Security Setup dialog box when KODAK MI Software was first launched, you can set up User Names and Passwords for each KODAK MI Software user. Proceed to *KODAK MI Security Manager*, later in this chapter.

9 Your hardware setup and software installation are now complete. To learn about capturing images with your Image Station 4000R, proceed to Chapter 4: *Setting Up for Capture*.

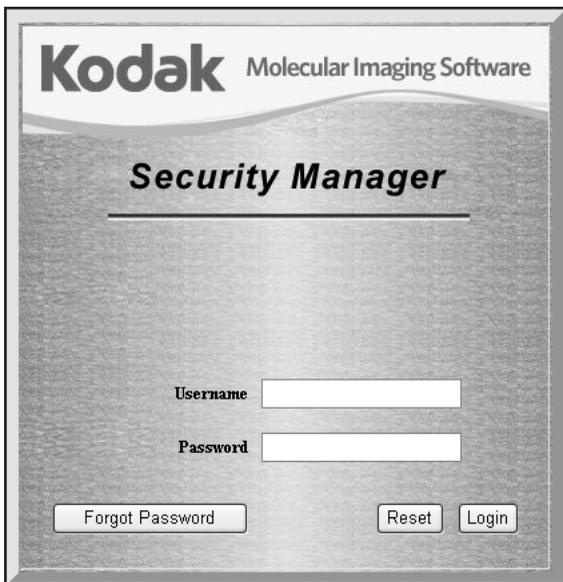
KODAK MI Security Manager

KODAK Molecular Imaging Software has two security modes: No Login (No Security) and Login Required (Password Security). When you launch KODAK MI Software for the first time, you are asked which mode you would like to use for this installation.

- ✓ In *No Login Mode*—all users gain access to KODAK MI Software bypassing the login screen.
- ✓ In *Password Mode*—requires you to enter your User Name and Password every time KODAK MI Software starts up.

👉 NOTE: The initial installation of KODAK MI Software comes with a single user with the User Name “Admin” and the Password “password”. Use this User name to gain access to KODAK MI Security Manager to set up users.

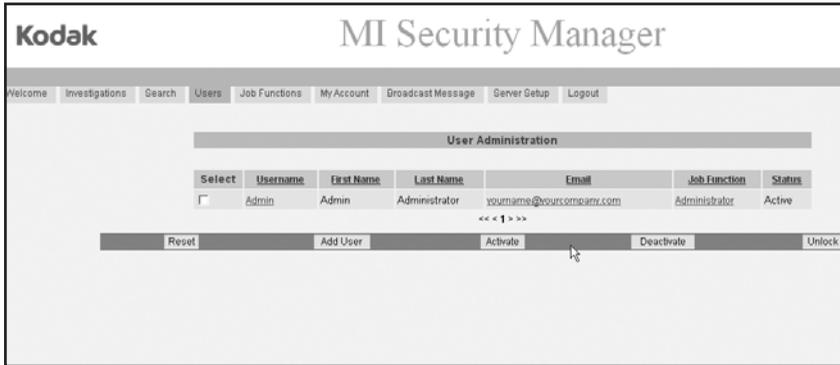
- 1 Select Database from the Navigation panel.
- 2 Your web browser launches and displays the KODAK Security Manager window opens.



The screenshot shows a web browser window titled "Kodak Molecular Imaging Software Security Manager". The window has a dark background with a light-colored header. The header contains the "Kodak" logo and the text "Molecular Imaging Software". Below the header, the title "Security Manager" is displayed in a large, bold font. Underneath the title, there are two input fields: "Username" and "Password". At the bottom of the window, there are three buttons: "Forgot Password", "Reset", and "Login".

- 3 Select the default Username, “Admin”, type the Password “password” (No quotation marks) and click Login.

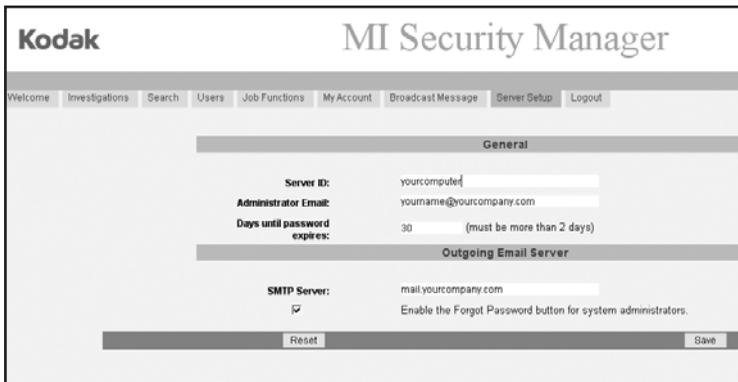
- 4 Click the Users tab to access the User Administration page, where you can add users. Select Add Users.



- 5 The User Information page appears. Fill out the information including First Name, Last Name, Username (enter a temporary password), Password, Job Function (Administrator, Principal Investigator, Research Scientist), an E-mail address. Click Create

 **NOTE:** New Users are automatically added to the default Investigation and default Study.

- 6 You may want to take some time to click on the Server Setup button to give your database server a name and specify a SMTP mail server.

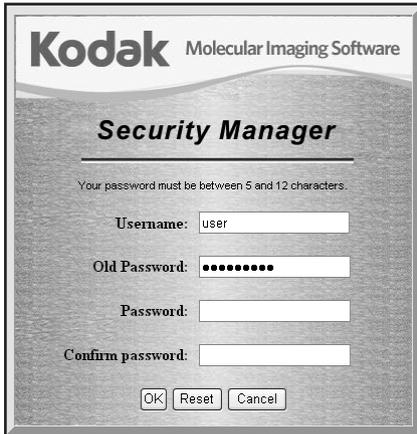


- ✓ *Server ID*—is a unique name for your machine. The Server ID can either be a name or number.

-
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- ✓ Administrator E-mail—this indicates the e-mail address of the Administrators's.
 - ✓ *Days until password expire*—allows administrators to determine how frequently passwords expire. When a User password expires, you will be prompted to enter a new password. the minimum is 2 days.
 - ✓ *SMTP Server*—This mail server can be used for sending out E-mails and broadcasting messages. This is a mandatory field that must be entered. The SMTP mail server is required when users click the Forgot Password button.
 - ✓ If the checkbox *Enable the forget password button* for administrator is checked. A user with system administrator privileges can use the forget password function.
- 7 An additional feature of KODAK MI Security Manager is the ability to broadcast e-mail messages to users. Administrators can enter a subject and message in the fields provided and click Send. All users receive the broadcast message. To send/receive a message, the user must have a valid e-mail address entered.

The screenshot shows the 'Kodak MI Security Manager' web interface. At the top, the 'Kodak' logo is on the left and 'MI Security Manager' is on the right. Below the logo is a navigation menu with tabs: 'Welcome', 'Investigations', 'Search', 'Users', 'Job Functions', 'My Account', 'Broadcast Message', 'Server Setup', and 'Logout'. The 'Broadcast Message' tab is selected. The main content area is titled 'Message Details' and contains two input fields: 'Subject:' and 'Message:'. The 'Message:' field is a large text area with a vertical scrollbar. At the bottom of the form, there are two buttons: 'Reset' and 'Send'.

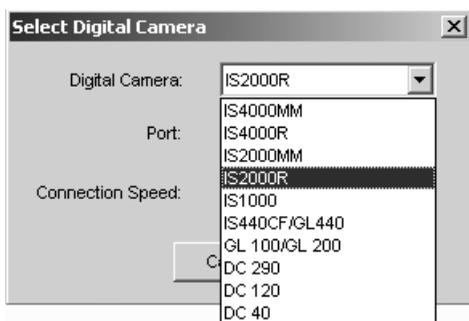
-
-
- 8 Click Logout to exit KODAK MI Security Manager. On the next restart of KODAK MI software, new users must enter their Username and password. The user will then be prompted to access the web portal to change their password.



The image shows a dialog box titled "Kodak Molecular Imaging Software Security Manager". At the top, it says "Kodak Molecular Imaging Software". Below that, the title "Security Manager" is centered. A note states: "Your password must be between 5 and 12 characters." There are four input fields: "Username:" with the text "user", "Old Password:" with ten black dots, "Password:" which is empty, and "Confirm password:" which is empty. At the bottom, there are three buttons: "OK", "Reset", and "Cancel".

Launching KODAK Molecular Imaging Software with the KODAK Image Station 4000R

- 1 Launch the software by clicking Programs from the Start menu and selecting KODAK MI.exe from the KODAK MI menu (Windows) or by clicking on the KODAK MI icon found in the KODAK MI folder (Macintosh). The project window appears.
- 2 Choose Select Digital Camera from the File menu. Choose the IS4000R from the Digital Camera pop-up menu. The software systematically searches the appropriate computer port(s). If the selected imaging device is not found, one of several messages will appear on-screen depending upon the imaging system selected.
 - ✓ If a KODAK Image Station 4000 System is not found, a message appears stating *Error Connecting with the Camera*. Ensure that IS4000R is properly connected and powered on, and that the IS4000R was selected in the Digital Camera pop-up menu.
- 3 Once IS4000R is found, the File menu and the Status bar Camera Access buttons reflect the current selection.
- 4 If no camera is found, the Select Digital Camera dialog box appears, allowing you to reselect the camera. Use the Digital Camera pop-up menu to select the IS4000R.



- 5 Click OK.

Repackaging or Moving the IS4000R

If you plan to ship your IS4000R, packing the components correctly is essential to protecting the imaging chamber and computer. Inappropriate packing will void the IS4000R warranty. Follow these directions when packing the IS4000R for shipment.

All returned material must be cleaned and decontaminated prior to shipping. To meet Federal and State Regulatory and Safety standards, please follow the decontamination procedure given here if radioactive materials are used with this product or are used in the vicinity of where this apparatus has been used or stored. If you are sending the unit for repair, contact Technical Support.

General Cleaning Procedure

For materials not contaminated with biological or radiological substances, components may be gently washed using water or an ammonia based spray cleaner using soft lint-free cloth or lens paper.



WARNING: Do not expose the Platen to strong chemical solvents such as ketones, hexanes, acids and alkalis that may permanently damage the Platen. This type of damage is not covered by the warranty.

Radiological Decontamination Procedure

For beta or gamma-emitting isotopes such as ^{32}P , use a GM-type radioactivity meter calibrated in counts per minute (CPM) to determine the background readings for your work area. Wearing latex gloves, survey the unit to be returned with the GM meter. If any part of the unit is found to show readings higher than background, wash the area using Count Off™ (PerkinElmer Life Sciences) or another similar commercially available detergent and paper towels. If none are available, a Formula 409™-like solution or a mild detergent will do. As you clean, discard liquid and solid waste (gloves and paper towels) according to your local and institutional regulations for radioactive material disposal. Continue washing until the GM meter reading for the contaminated area(s) is equal to or below background.

To decontaminate units where a GM-meter is not useful for detection, such as with ^3H or ^{35}S , it will be necessary to perform swipes of the unit and detect using a scintillation counter. The unit should be dry. Wipe surfaces with dry paper circles (these are commercially available or you can make your own). Areas can be charted, so that individual swipes can be done on different surfaces to better isolate areas of

contamination. These swipes are then placed into individual scintillation vials with an appropriate fluor and then analyzed on a properly programmed scintillation counter. If contamination above 100 disintegrations per minute dpm/100 cm² (dpm = CPM/efficiency) is found, wash the area as described above for beta emitting isotope decontamination. After cleaning the area, swipe it a second time to determine the amount of contamination remaining. If the area still has a greater than 100 dpm/cm², continue the cycle of swipes and washing until you achieve a reading of less than 100 dpm/cm².

Once the unit has been determined to be radiation free (< 100 dpm/cm²), remove all the hazardous and radioactive labels from the unit. If the labels cannot be removed, deface them. Failure to do so may result in significant delay or a refusal of repair.

If your unit has unremovable contamination (detectable with a GM-meter and not with paper swipes, or detectable with paper swipes but after continued washing the dpm/cm² remains constant above 100) of a short half-life isotope such as ³²P, the unit may be stored for 10 half-lives of isotopic decay and the decontamination procedure repeated.

 NOTE: Units contaminated with unremovable long half-life isotopes should not be shipped without proper procedures.

Repacking the IS4000R

Inappropriate packing will void the IS4000R warranty. Follow these directions when packing the IS4000R for shipment.

- 1** Decontaminate the IS4000R as described above.
- 2** Close and latch the lid.
- 3** Remove the power cord and IEEE 1394 (FireWire) Camera/ Computer Interface Cable from the IS4000R.
- 4** Remove any filters from the Filter Wheel and cradle the camera using foam padding.
- 5** Secure the Filter Access Panel to the filter housing with masking tape.
- 6** Put the IS4000R in its original plastic bag.
- 7** Pack the IS4000R in its original box. If you have discarded the original packing material, call your Molecular Imaging Systems dealer or Technical Support to arrange for delivery of new packing material.

Warranty & Regulatory Information

This section contains the IS4000R warranty and software license agreements information.

IS4000R Limited Warranty

Warranty Time Period

Carestream Health, Inc. warrants the KODAK Image Station 4000R (IS4000R) to function properly for one year from the date of purchase. Specific system components warranted by Carestream Health, Inc. are the IS4000R chamber, the hardware components installed within the chamber, the IEEE 1394 (FireWire) Camera/Computer Interface Cable, power cord, the KODAK Molecular Imaging Software, and optional Image Station Transilluminator Lid Accessory. Other components included with the IS4000R system, including the computer, monitor and illuminator are under separate warranty provided by their respective manufacturer.

Days and Hours of Coverage

Arrangements for service through Technical Support can be made Monday through Friday 8:00 a.m. to 6:00 p.m. EST in the United States, except for locally observed holidays. Hours of coverage outside the United States may vary. Contact your local Molecular Imaging Systems dealer for hours of coverage.

Warranty Repair Coverage

If the equipment does not function properly during the warranty period due to defects in either materials or workmanship, Carestream Health, Inc. will, at its option, either repair or replace the equipment without charge, subject to the conditions and limitations stated herein. Such repair service will include all labor as well as any necessary adjustments and/or replacement parts.

If replacement parts are used in making repairs, these parts may be remanufactured, or may contain remanufactured materials. If it is necessary to replace the entire system, it may be replaced with a remanufactured system.

Carestream Health, Inc. will also provide telephone assistance during the warranty period.

Limitations

WARRANTY SERVICE WILL NOT BE PROVIDED WITHOUT DATED PROOF OF PURCHASE. PLEASE RETURN THE WARRANTY REGISTRATION CARD WITHIN 30 DAYS OF PURCHASE.

THIS WARRANTY BECOMES NULL AND VOID IF YOU FAIL TO PACK YOUR INSTRUMENT IN A MANNER CONSISTENT WITH THE ORIGINAL PRODUCT PACKAGING AND DAMAGE OCCURS DURING PRODUCT SHIPMENT.

THIS WARRANTY DOES NOT COVER: CIRCUMSTANCES BEYOND CARESTREAM HEALTH, INC.'S CONTROL; SERVICE OR PARTS TO CORRECT PROBLEMS RESULTING FROM THE USE OF ATTACHMENTS, ACCESSORIES OR ALTERATIONS NOT MARKETED BY CARESTREAM HEALTH, INC.; SERVICE REQUIRED AS THE RESULT OF UNAUTHORIZED MODIFICATIONS OR SERVICE; MISUSE, ABUSE; FAILURE TO FOLLOW CARESTREAM HEALTH, INC.'S OPERATING, MAINTENANCE OR REPACKAGING INSTRUCTIONS; OR FAILURE TO USE ITEMS SUPPLIED BY CARESTREAM HEALTH, INC. (SUCH AS ADAPTERS AND CABLES).

CARESTREAM HEALTH, INC. MAKES NO OTHER WARRANTIES, EXPRESS, IMPLIED, OR OF MERCHANTABILITY OR FITNESS FOR A PARTICULAR PURPOSE FOR THIS EQUIPMENT OR SOFTWARE.

REPAIR OR REPLACEMENT WITHOUT CHARGE ARE CARESTREAM HEALTH, INC.'S ONLY OBLIGATION UNDER THIS WARRANTY. CARESTREAM HEALTH, INC. WILL NOT BE RESPONSIBLE FOR ANY SPECIAL, CONSEQUENTIAL OR INCIDENTAL DAMAGES RESULTING FROM THE PURCHASE, USE, OR IMPROPER FUNCTIONING OF THIS EQUIPMENT REGARDLESS OF THE CAUSE. SUCH DAMAGES FOR WHICH CARESTREAM HEALTH, INC. WILL NOT BE RESPONSIBLE INCLUDE, BUT ARE NOT LIMITED TO, LOSS OF REVENUE OR PROFIT, DOWNTIME COSTS, LOSS OF USE OF THE EQUIPMENT, COST OF ANY SUBSTITUTE EQUIPMENT, FACILITIES OR SERVICES, OR CLAIMS OF YOUR CUSTOMERS FOR SUCH DAMAGES.

Depending on your geographical location, some limitations and exclusions may not apply.

How to Obtain Service

If the instrument does not function properly during the warranty period, contact Carestream Health, Inc. or your local Carestream Health, Inc. dealer to arrange for service. If your unit needs to be returned for any reason, please contact your local Carestream Health, Inc. dealer to obtain a return authorization. All returned units must be decontaminated prior to their return, as described below. No returns will be accepted without a return authorization and proper decontamination documentation.

When contacting technical support, please have the following information available:

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- ✓ The serial number of your IS2000R system located on the back of the unit and the camera serial number located on the camera.
 - ✓ The serial number and version number of your KODAK Software.
 - ☞ NOTE: With the software running, select About KODAK MI under the Help menu (Windows) or select About KODAK MI under the Apple menu item (Macintosh).
 - ✓ The type of computer you are using (make, model and operating system version).
 - ☞ NOTE: Check your operating system version by right-clicking on the My Computer icon and then on Properties (Windows) or select About This Mac under the Apple menu item (Macintosh).
 - ✓ The problem you are having and what you were doing when the problem occurred. Please note the exact wording of any error messages, including any error numbers displayed.

When contacting technical support by telephone, you will be assisted best if you are seated in front of your computer with KODAK MI Software running.

Contact Molecular Imaging Systems Technical Support by:

- ✓ Utilizing our World Wide Web support pages at:
<http://www.carestreamhealth.com/go/molecular>
- ✓ Calling Molecular Imaging Systems Technical Support toll free at:
877-747-4357, between the hours of 8:00 a.m. and 6:00 p.m. (Eastern Standard Time) Monday through Friday
- ✓ E-mailing Molecular Imaging Systems Technical Support at:
molecular-support@kodak.com
- ✓ Faxing Molecular Imaging Systems Technical Support at:
203-786-5656

For up-to-date dealer information, visit our WEB site at www.carestreamhealth.com/go/molecular.

General Instructions for Cleaning and Decontamination for Return of the Kodak IS4000R

All returned material must be cleaned and decontaminated prior to shipping. To meet Federal and State Regulatory and Safety standards, please follow the decontamination procedure given here if radioactive materials are used with this product or are used in the vicinity of where this apparatus has been used or stored. You will be required to sign a certificate of decontamination prior to sending the IS4000R for repair. Any instruments received with contamination will be subject to a charge for decontamination.

General Cleaning Procedure

For materials not contaminated with biological or radiological substances, components may be gently washed using water or an ammonia based spray cleaner using soft lint-free cloth or lens paper.



WARNING:Ammonia based spray cleaners may cause eye irritation. Consult the manufacturer's material safety data sheet for additional information prior to use.



WARNING:Do not expose the Platen to strong chemical solvents such as ketones, hexanes, acids and alkalis that may permanently damage the Platen. This type of damage is not covered by the warranty.

Radiological Decontamination Procedure



WARNING:Carestream Health, Inc. cannot accept return of products which are contaminated with any radioactivity.

For beta-or gamma-emitting isotopes such as ^{32}P , use a GM-type survey meter with a thin-end window detector, calibrated in counts per minute (CPM) to determine the background readings for your work area. Wearing latex gloves, survey the unit to be returned with the GM meter. If any part of the unit is found to show readings higher than background, wash the area using Count Off™ (PerkinElmer Life Sciences) or another similar commercially available detergent and paper towels. If none are available, a Formula 409™-like solution or a mild detergent will do. As you clean, discard liquid and solid waste (gloves and paper towels) according to your local and institutional regulations for radioactive material disposal. Continue washing until the GM meter reading for the contaminated area(s) is equal to or below background.

To decontaminate units where a GM-meter is not useful for detection, such as with ^3H , it

will be necessary to perform swipes of the unit and detect using a scintillation counter. The unit should be dry. Wipe surfaces with dry paper circles (these are commercially available or you can make your own). Areas can be charted, so that individual swipes can be done on different surfaces to better isolate areas of contamination. These swipes are then placed into individual scintillation vials with an appropriate fluor and then analyzed on a properly programmed scintillation counter. If contamination above 100 disintegrations per minute dpm/100 cm² (dpm = CPM/efficiency) is found, wash the area as described above for beta-emitting isotope decontamination. After cleaning the area, swipe it a second time to determine the amount of contamination remaining. If the area still has a greater than 100 dpm/cm², continue the cycle of swipes and washing until you achieve a reading of less than 100 dpm/cm.²

Once the unit has been determined to be radiation free (< 100 dpm/cm²), remove all the hazardous and radioactive labels from the unit. If the labels cannot be removed, deface them. Failure to do so may result in significant delay or a refusal of repair.

If your unit has unremovable decontamination (detectable with a GM-meter and not with paper swipes, or detectable with paper swipes but after continued washing the dpm/cm² remains constant above 100) of a short half-life isotope such as ³²P, the unit may be stored for 10 half-lives of isotopic decay and the decontamination procedure repeated.

 NOTE: Units contaminated with unremovable, long half-life isotopes may not be returned.

Repackaging the IS4000R

Damage to the instrument due to inappropriate packing is not covered by the IS4000R warranty. Follow these directions when packing the IS4000R for shipment.

- 1** Decontaminate the IS4000R as described above.
- 2** Close and latch the lid.
- 3** Remove the power cord and IEEE 1394 (FireWire) Camera/ Computer Interface Cable from the IS4000R.
- 4** Remove any filters from the Filter Wheel and cradle the camera using foam padding.
- 5** Secure the Filter Access Panel to the filter housing with masking tape.
- 6** Put the IS4000R in its original plastic bag.
- 7** Pack the IS4000R in its original box. If you have discarded the original packing material, call your dealer or Technical Support to arrange for delivery of new packing material.

Software License Agreement

You should carefully read the following terms and conditions before using this software package. Using the KODAK Molecular Imaging Software indicates your acceptance of these terms and conditions. If you do not agree with them, you should promptly return the disk package unopened, along with its accompanying materials, and your money will be refunded.

Carestream Health, Inc. provides this program and licenses its use. You assume responsibility for selection of the program to achieve your intended results, and for installation, use, and results obtained from the program.

License

You may:

- a. use the program on a single machine, or if you have purchased a network license, on the number of machines authorized by your network license (as indicated on the invoice for that license);
- b. copy the program into any machine-readable or printed form for backup or modification purposes in support of your use of the program;
- c. transfer the program and license to another party if the other party agrees to accept the terms and conditions of this Agreement. If you transfer the program, you must at the same time either transfer all copies whether in printed or machine-readable form to the same party or destroy any copies not transferred; this includes all modifications and portions of the program contained or merged into other programs.

You must reproduce and include the copyright notice on any copy, modification, or portion merged into another program.

You may not use, copy, modify, or transfer the program, or any copy, modification, or merged portion, in whole or in part, except as expressly provided for in this license.

If you transfer possession of any copy, modification, or merged portion of the program to another party, your license is automatically terminated.

Term

The license is effective until terminated. You may terminate it at any time by destroying the program together with all copies, modifications, and merged portions in any form. It will also terminate upon conditions set forth elsewhere in this Agreement or if you fail to comply with any term or condition of this Agreement. You agree upon such termination to destroy the program together with all copies, modifications, and merged portions in any form.

Limitation of Liability and Remedy

Carestream Health, Inc. warrants that it has full power to enter into this Agreement and to grant you the rights provided herein and that the software will substantially conform to Carestream Health, Inc.'s published specifications. Carestream Health, Inc. further warrants that it will defend you against any claim that the software supplied hereunder infringes a patent, copyright, or other third party intellectual property right, and Carestream Health, Inc. will pay any costs and damages that a court finally awards against you as a result of such claim, provided you give Carestream Health, Inc. prompt written notice of such claim and tender to Carestream Health, Inc. the defense and all related settlement negotiations. Carestream Health, Inc. shall have no obligation with respect to any such claim based upon your modification of software or its combination, operation, or use with data or programs not furnished by Carestream Health, Inc. or with other than the specified computer system. At any time during the course of any litigation arising out of such a claim, or if, in Carestream Health, Inc.'s opinion, the software or any part thereof is likely to become the subject of a claim of infringement, Carestream Health, Inc. will, at its option and at its expense, either procure for you the right to continue using the software, replace or modify the same so that it becomes non-infringing, or grant you a credit for the software as depreciated, and accept its return. The depreciation will be an equal amount per year over the lifetime of the software as established by Carestream Health, Inc.

EXCEPT FOR REPLACEMENT OF UNMODIFIED COPIES OF SOFTWARE FOUND BY CARESTREAM HEALTH, INC. TO BE DEFECTIVELY REPRODUCED OR DAMAGED PRIOR TO DELIVERY TO YOU, CARESTREAM HEALTH, INC. EXPRESSLY EXCLUDES ALL OTHER REMEDIES OR WARRANTIES OTHER THAN THE FOREGOING, EXPRESS OR IMPLIED, IN RELATION TO THE SOFTWARE (AND ANY SERVICES RENDERED TO SUPPORT THE SOFTWARE), INCLUDING ANY WARRANTY OF MERCHANTABILITY OR OF FITNESS FOR A PARTICULAR PURPOSE.

In particular, Carestream Health, Inc. does not warrant that the functions contained in the program will meet your requirements or that the operation of the program will be uninterrupted or error free.

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IN NO EVENT WILL CARESTREAM HEALTH, INC.'S LICENSOR(S), THEIR DIRECTORS, OFFICERS, EMPLOYEES OR AGENTS (COLLECTIVELY CARESTREAM HEALTH, INC.'S LICENSOR) BE LIABLE TO YOU FOR ANY CONSEQUENTIAL, INCIDENTAL, OR INDIRECT DAMAGES (INCLUDING DAMAGES FOR LOSS OF BUSINESS PROFITS, BUSINESS INTERRUPTION, LOSS OF BUSINESS INFORMATION AND THE LIKE) ARISING OUT OF THE USE OR INABILITY TO USE THE SOFTWARE EVEN IF CARESTREAM HEALTH, INC.'S LICENSOR HAS BEEN ADVISED OF THE POSSIBILITY OF SUCH DAMAGES. BECAUSE SOME JURISDICTIONS DO NOT ALLOW THE EXCLUSION OR LIMITATION OF LIABILITY FOR CONSEQUENTIAL OR INCIDENTAL DAMAGES; THE ABOVE LIMITATIONS MAY NOT APPLY TO YOU.

General

You may not sublicense, assign, or transfer the license or the program except as expressly provided for in this Agreement. Any attempt otherwise to sublicense, assign, or transfer any of the rights, duties, or obligations hereunder is void.

This Agreement will be governed by the laws of the State of New York.

Should you have any questions concerning this agreement, contact Technical Service, Molecular Imaging Systems, Carestream Health, Inc., 4 Science Park West, New Haven, CT 06511, USA.

Regulatory Information

“This equipment has been tested and found to comply with the limits for a Class A digital device pursuant to Part 15 of the FCC rules. These limits are designed to provide reasonable protection against harmful interference when the equipment is operated in a commercial environment. This equipment generates, uses, and can radiate radio frequency energy and, if not installed and used in accordance with the instruction manual, may cause harmful interference to radio communications. Operation of this equipment in a residential area is likely to cause harmful interference in which case the user will be required to correct the interference at his own expense. Any changes or modifications not expressly approved by the party responsible for compliance could void the user’s authority to operate the equipment. Where shielded interface cables have been provided with the product or specified additional components or accessories elsewhere defined to be used with the installation of the product, they must be used in order to ensure compliance with FCC regulations.

This Class A digital apparatus meets all requirement of the Canadian Interference-Causing Equipment Regulations.

Cet appareil numérique de la classe A respecte toutes les exigences du Règlement sur le matériel brouilleur du Canada.”

For Use in European Union

WARNING

This is a class A product. In a domestic environment this product may cause radio interference in which case the user may be required to take adequate measures.

For Use in Japan

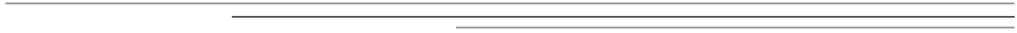
この装置は、情報処理装置等電波障害自主規制協議会（VCCI）の基準に基づくクラスA情報技術装置です。この装置を家庭環境で使用すると電波障害を引き起こすことがあります。この場合には使用者が適切な対策を講ずるよう要求されることがあります。

For Use in Taiwan

警告使用者:

這是甲類的資訊產品，在居住的環境中使用時，可能會造成射頻干擾，在這種情況下，使用者會被要求採取某些適當的對策。

The IS4000R contains small amounts of lead in the lens and circuit boards and mercury in the lamps. Disposal of these materials may be regulated due to environmental considerations. For disposal or recycling information, please contact your local authorities, or in the U.S.A., contact the Electronics Industry Alliance web site at www.eiae.org



Digital Imaging Concepts

Digital imaging principles as they relate to the IS4000R are discussed in this appendix in somewhat greater detail than elsewhere in the manual. Gaining a working knowledge of these concepts will be helpful in making you at home with digital, electronic and quantitative imaging technology. The discussion is presented in an order that starts with the most difficult concept, resolution. The concept of image resolution is central to the understanding of all aspects of digital and quantitative imaging.

Image Resolution

Image resolution generally refers to the capability to distinguish objects of interest. This discussion focuses on image data in an effort to clarify the point that the information in an image data file has a limited capacity to resolve objects, whether or not those objects may be visibly resolved. In digital imaging, resolution can be discussed with greater clarity since the fundamental concepts and terms are well-defined and amenable to measure. The concepts of bit depth, grayscale, CCD, noise, pixels and many other relevant terms are briefly defined in the *Glossary*. To simplify the following discussion, the term “objects” refers to physical items located on a flat surface that emit or modulate light that is captured by an imaging system. The image information resides in an image data file, and the word “image” most often refers to the data file. A subsequent representation of the image may be printed or displayed on screen for viewing; hence, the image data may be visualized as a “perceived image”.

As defined above, an object in a plane is emitting a signal or is otherwise modulating light, creating contrast that is construed as signal. The signal is captured, and an image represents that object within a measurable precision or level of error. The image data may be conceived as having an X,Y plane, and a signal (G) associated with each point on the X,Y plane. See Figure 1. Each of the image variables X, Y, and G have an uncertainty or an error introduced by the imaging system. The capability of the imaging data to resolve two objects depends upon the uncertainty of each of these variables. The uncertainty in the X and Y image coordinates is called the spatial component of resolution, and depends upon the number of CCD pixels and the limitation of all elements in the optical path. The uncertainty in the signal G is called the dynamic range component of resolution and depends upon the CCD and supporting electronics. The smaller any of these uncertainties, the higher the precision and the higher the resolution. While the variables X, Y, and G are not necessarily separable variables in an image, resolution is generally simplified into spatial and dynamic range considerations.

Figure 1.

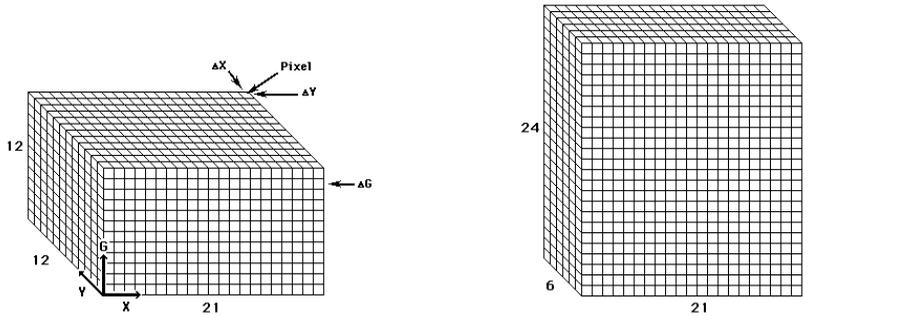


Image resolution is related to the volume of information in a cube with dimensions X and Y (the image plane) and G (the dynamic range).

Binning the adjacent Y -pixels, by a factor of 2 or 4, doubles or quadruples the signal G resolution, but halves or quarters the Y -resolution.

If the coordinate X (a position X in the X, Y plane of the image) has an inherent error or uncertainty ΔX , then the X coordinate may be divided into $X/\Delta X$ increments, each one of which significantly differs from its neighbor by the margin of error ΔX . See Figure 1. Similarly, Y is divided into $Y/\Delta Y$ increments. The fundamental element having a dimension of ΔX and ΔY is defined as the smallest spatial element in the image that may be resolved in the X, Y plane. Similarly, the signal variable G has an uncertainty ΔG , and can be divided into $G/\Delta G$ increments. The signal error is generally called noise; hence, the number of significantly different signal increments ($G/\Delta G$) is the signal/noise ratio, which is the most relevant measure of dynamic range. Thus, the smallest increment of the signal scale that may be resolved within the spatial element $\Delta X \Delta Y$ is ΔG . Both the spatial and dynamic range components of an image are discussed later in this section.

Spatial Resolution

Assuming the spatial resolution is CCD limited, the smallest spatial element in the image is represented by a “pixel”, having the dimension ΔX by ΔY . Depending upon the image magnification, a certain total number of image pixels of a CCD are applied to an object area, limiting the spatial resolution to a certain number of pixels per object area, often referred to as pixel density. To simplify, the pixel limitation of resolution is usually quoted for only one dimension (as in pixels/mm), assuming that X and Y magnification are identical and the pixels are square. The pixel limited spatial resolution of the IS4000R is variable (zoom lens), ranging from a low resolution of about 10 pixels/mm to a high resolution of about 10 pixels/mm.

Other measures or quotations are often associated with spatial resolution. The measure of dots/inch or dots/mm is often used in the printing industry, and is a true measure of printed resolution; it is clearly analogous to the pixels/mm quoted above. Another common measure used in imaging is line-pairs/mm, a number which corresponds to one-half of the above pixels/mm; two pixels are needed to present the same information as one line-pair.

As a general rule of thumb, a minimum of two image pixels or one line-pair must be applied to an object to “resolve” the object in the image. The same applies to the distance between two objects that are to be resolved. For example, objects 0.2 mm apart will require at least 10 pixels/mm or 5 line-pairs/mm to discern the space between them (spatially resolve the objects). While this general rule may be a mathematical necessity for spatial resolution, it is not sufficient. Although similar objects within an image captured at a high signal/noise may be well resolved using a spatial resolution of two or more image pixels per object, objects having extreme differences in signal may not be resolved, or objects captured with a low signal/noise may not be resolved.

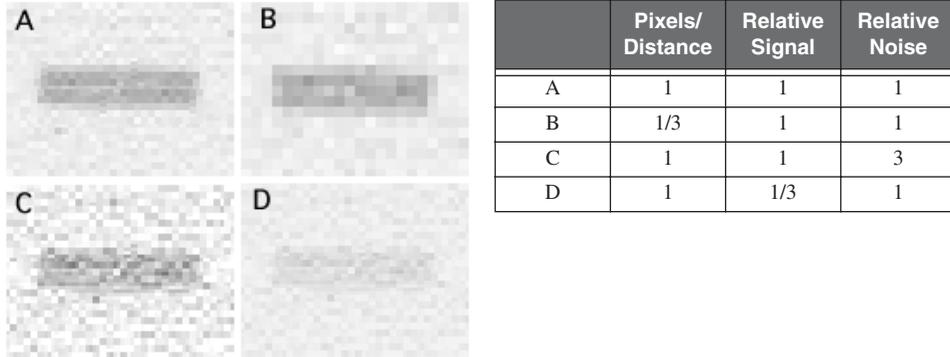


Figure 2. Resolution depends upon spatial and dynamic range variables.

A relevant example of image resolution is displayed in Figure 2, in which two bands are perceived as being nearly “resolved” in the upper left panel (A). If the image is manipulated to reduce the spatial resolution ($1/3$ pixels/distance (B)), or alternatively manipulated to reduce the signal resolution (3X noise) (C) and ($1/3$ signal (D)), the resolution of the bands is obscured. Hence, image resolution depends on both spatial and dynamic range considerations.

Dynamic Range

Dynamic range is the name given to the scale of signal or signal/noise ratio of a digital camera system. The maximum signal/noise that is quoted for an imaging system is the ratio of the maximum signal to the minimum noise. The maximum signal of the IS4000R is 16-bits per capture, or 65,536 signal levels. The minimum noise is approximately 12 signal levels. Therefore, the maximum signal/noise ratio ($65,536/12$) is about 5,460, and is related to the number of significantly different signals $G/\Delta G$ in the above discussion of *Resolution*.

The dynamic range of a camera may be increased by adding different image captures of an object together. The signal scale must be increased to accommodate higher numbers; the IS4000R software provides a 32-bit scale (4.3×10^9), providing more signal levels than needed for the accumulation of multiple images. While signal among accumulated images directly adds, the noise among the images generally does not. If the noise level of a series of independent (but otherwise identical) images is random, then the noise level of the accumulated image will be the product of the noise (of any one image) and the square root of the number of accumulated images. Thus, the signal/noise ratio of an accumulated image is increased by the square root of the number of images participating in the accumulation. The maximum signal/noise ratio of the IS4000R accumulated image is ideally achieved with the addition of 32 images, giving a maximum ratio of $(65,536 \times \sqrt{32})/12$, which is about 30,889.

Noise

Image noise is a random deviation in signal and is identified with the signal error or uncertainty ΔG in the previous discussion on *Resolution*. Noise may be measured among pixel signals within an image (for example, background), or among the signals derived from the same pixel from different but otherwise identical images. Noise is most often represented as a standard deviation of a collection of signals about a mean. Noise arising from a digital camera system has two distinct causes:

- ✓ The supporting electronics contribute “read” noise as the CCD pixel signals are converted to digital signals. The read noise may appear nearly random in a single capture, but usually has subtle non-random components, which become apparent upon adding a large number of images wherein structured features are obvious. For a single exposure (one “read”), the read noise of the IS4000R is about 12 signal levels using a 16-bit scale.
- ✓ The dark (no light) charge accumulation within a CCD pixel increases with exposure time contributing “dark current noise”. Dark current noise is directly related to exposure time, and is significantly reduced with operating temperature (thermoelectric cooling). The “dark” component of noise is generally quoted as a coefficient of time. The IS4000R has a cooled CCD, and the nominal dark noise is very low.

Noise components in imaging generally add according to the square root of the sum of the squares of the components. A summary of the IS4000R dark noise components is shown in the following graph

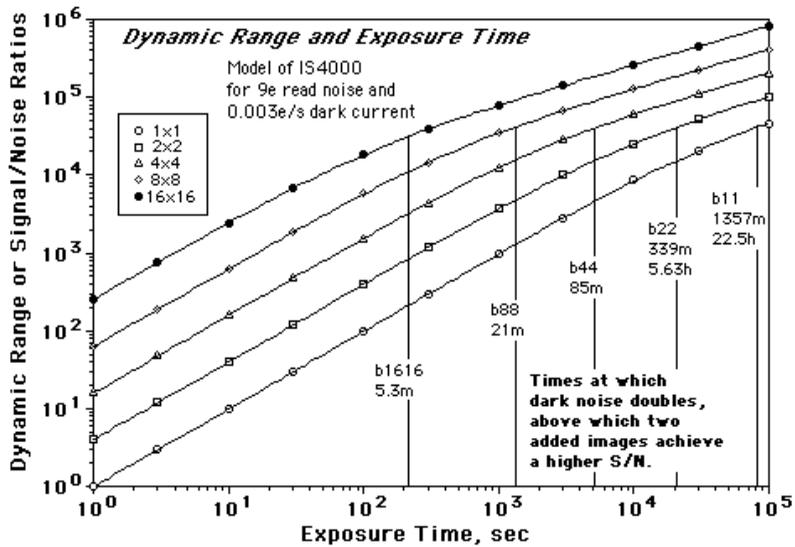


Figure 3. IS4000R noise estimates.

Upon studying the graph, it becomes clear that guidelines relating to the number of captures and exposure time per capture can be established for the IS4000R to minimize noise. There will be some variation among IS4000R cameras, and extreme ambient temperature may influence the dark noise. It is most likely, however, that a single capture will minimize unless extremely low light is limiting the signal. Should multiple capture accumulations be necessary to attain signal significance, a rule of thumb for the above graph suggests that a single exposure of 1360/binning number (in minutes) should be utilized, where binning number is the product of the bin state. For example, for 8 X 8 binning, $1360/64 = 21$ minutes, so an accumulation of 21 minutes captures maximizes the signal/noise ratio for 688.

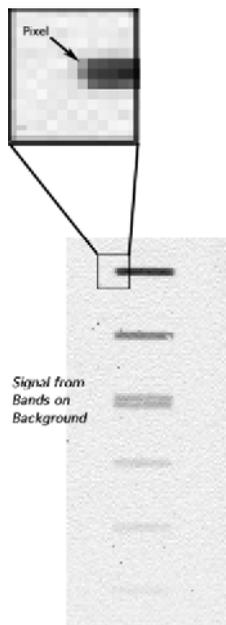
Note that the above guidelines are based upon image noise only, and give no consideration to signal or other aspects of the typical image capture. The above guidelines relate to those captures that are signal-limited, typical luminescent captures, wherein noise minimization is crucial. For those imaging experiments with ample signal to fill the available scales in a relatively short amount of time, the above guidelines do not relate. In most fluorescent images, the experimental background noise will be determined by the materials used in the experimental sample rather than the noise contributed by the electronic system; thus, a large number of “reads” will not significantly detract from the image quality, and may lend an element of experimental convenience as the image display and histogram “grows” in significance as the images accumulate.

Binning Pixels

The total amount of information the image file is capable of containing is the summation of all elements in the X,Y,G volume, or the product $(Y/\Delta Y)(X/\Delta X)(G/\Delta G)$ (See Figure 1). Just as information is the capability to distinguish between alternatives, then the total volume of the cube is related to the image resolution and it is the basis for discerning objects. Note that the information volume may be unchanged if the shape of the cube is changed or distorted (any one dimension may increase with a proportional decrease of another).

The CCD has a special hardware capability that can transform the shape in this information cube. The capability is called binning, and it is the addition of the signal levels of adjacent pixels without the addition of noise. If adjacent pixels are binned in the Y dimension yielding half or a quarter of the Y levels, the number of levels on the signal scale will double or quadruple. Should both the X and Y dimensions be binned, the signal scale will be 1X, 4X, 16X, 64X and 256X corresponding to 1 x 1, 2 x 2, 4 x 4, 8 x 8, and 16 X 16 binning states. Binning in both X and Y directions can be a 2X or 4X bin in each direction. The maximum signal scale is 16X. The volume of the cube remains the same, hence the image resolution has not changed. If an image has a spatial resolution that far exceeds the need to resolve an object, then the apparent resolution of the object is increased by binning. This increase is usually associated with an apparent increase in detection sensitivity when binning is effectively applied. To reduce confusion, asymmetric binning is limited to the horizontal direction of the user perspective. Asymmetric binning is useful in the case where spacial resolution can be sacrificed in one direction only, typically in a separatory experiment (a gel or blot). Hence, the gel is oriented so that the sample wells are horizontal. Upon image capture in the 4 X 4 and X 1 states, the band resolution in the Y direction is conserved, and the signal is enhanced by 4 X at the expense of reduced sample well resolution.

Background Subtraction



The background signal level, shown in Figure 4, is associated with the absence of experimental signal. In an image histogram, background is a peak of signal frequency, having a mean background level and a breadth related to image noise. The low signal side of the peak diminishes to a pixel frequency of zero at a signal level of about 200 on a 65,535 scale for a single image capture of the IS4000R. Below the level of zero pixel frequency (200 in this case) there is no image information, and this level is simply the total electronic background. The electronic background is composed of a dark current (electronic charges emanating from the CCD and amplifier), and an intentional voltage offset to assure that the electronic background is always positive.

The electronic background diminishes the total dynamic range of the IS4000R by a small amount, resulting in a net signal level maximum of about $65,535 - 200 = 65,335$. While the background is small, it may be inconvenient or detrimental to a subsequent analysis and it is best subtracted. Electronic background subtraction is managed by the IS4000R Image Acquire software.

Figure 4

Electronic background subtraction must be executed with sufficient precision to assure that the zero-level of signal equals the actual level of no light. Given an appropriate no-light/zero-signal, lens and illumination correction may be executed with precision. See Appendix C: *Image Correction*. As many as half the pixel signal levels become zero upon subtracting electronic background from a perfectly dark image. Only the upper half of the background noise peak represented by the histogram remains, consequently estimates of the background electronic noise are distorted, but are nevertheless consistent throughout the image. Subsequent lens correction will further distort the background noise with a function that increases in a radial manner (from the middle of the image outward). Should an image of uniform background noise be desirable for analysis, the file must be saved prior to lens correction.

Software management of the electronic background is camera-specific, and the only the software registered to the specific IS4000R camera properly encodes the appropriate background and defective pixel information.

UV Epi-Illumination

In the following section, you will learn about the principles of UV Epi-illumination as they relate to the IS4000R.



WARNING: According to “Recommended Practice for Photobiological Safety for Lamps & Lamp Systems—General Requirements (ANSI/IESNA)”, the UV light emitted from the IS4000R during normal or continuous use is safe for the general population. The safety of the IS4000R relates to both skin and eye exposure. By comparison, the UV exposure is similar to that received from common overhead lighting in laboratories, and is much less than that received by sunlight at sea-level. Hence, no safety warning is issued, nor is any appropriate, for the normal use of the IS4000R. However, as noted in the cited publication, there are certain photosensitive individuals whose tolerance to UV is reduced, but guidelines have not been established.

The KODAK Image Station 4000R supplies UV irradiation to the sample Platen from the inside of the chamber. The light is directed at a 45° angle to the underside of the Platen, which is generally called “epi-illumination”. The illumination system has been designed to optimize the physical features that relate to the fluorescent excitation of samples placed on the Platen, among which are:

- 1** The optical elements that manage the illumination have been designed to provide a reasonably uniform distribution of excitation energy over the Platen area. While it is not possible to design absolutely uniform illumination from light sources at the distance used in this system, the design does achieve a uniformity that is at about the same quality that a camera lens can represent. The KODAK Molecular Imaging Software provides corrections that address both the lens and illumination distortions, thus contributing to a quality analysis of the image data.
- 2** The epi-illumination angle requires that the excitation photons interact with the sample and be re-emitted in order to have an angle appropriate to enter the camera lens. Photons simply reflected by a planar surface at the platen will not enter the lens, obviating the required use of special optical filters that reject the excitation energy. However, reflected light from curved surfaces in a sample can enter the lens and will present artifacts in fluorescent images. There are specialized filters to accommodate these artifacts.
- 3** The energy spectrum of the illumination has been designed to provide a broad range of light (300–400 nm, see Figure) within the range of wavelengths called the UVA (ultraviolet absorbance). The 300–400 nm range of wavelengths effectively excites a very large number of fluorochromes in common usage. Even fluorochromes that are particularly suited to laser-excitation at a much higher wavelength may also be excited by the IS4000R UV epi-illumination.
- 4** Compared to commonly used UV transilluminators or hand-held UV light source, the energy spectrum of the IS4000R UV epi-illumination system emits very little energy below 300 nm. See Figure. Compared to the 300–400 nm range of light, light below 300 nm contributes very little to the excitation of fluorochromes in common usage, but does substantially contribute to the general fluorescence of commonly used materials in laboratory samples. Hence, lower autofluorescent background may be anticipated in the fluorescent image applications supported by the IS4000R.

The absence of energy below 300 nm in the IS4000R contributes to its safe usage. Wavelengths below 300 nm are hazardous. See Figure and Safety Statement.

- 5** The total energy of the IS4000R UV epi-illumination is very low, nominally 50 µW/

cm² at the Platen surface. However, the digital camera is sufficiently sensitive to assure that fluorescence data can be accumulated over a reasonable period of time with UV epi-illumination, provided. Additionally, the low intensity reduces the possibility that a fluorescent response is the result of multi-photon molecular processes that can contribute to the autofluorescence of solid materials commonly used in laboratory samples (such as nylon).

The reduced energy of the IS4000R UV epi-illumination eliminates the possibility of visualizing the fluorescence of a sample, so its usefulness for cutting bands out of gels is precluded, and should never be attempted on an optical surface critical to image quality.

The total energy emanating from the IS4000R Platen is less than 200-fold less than commonly used UV transilluminators, contributing to its safe usage. See Figure and Safety Statement.

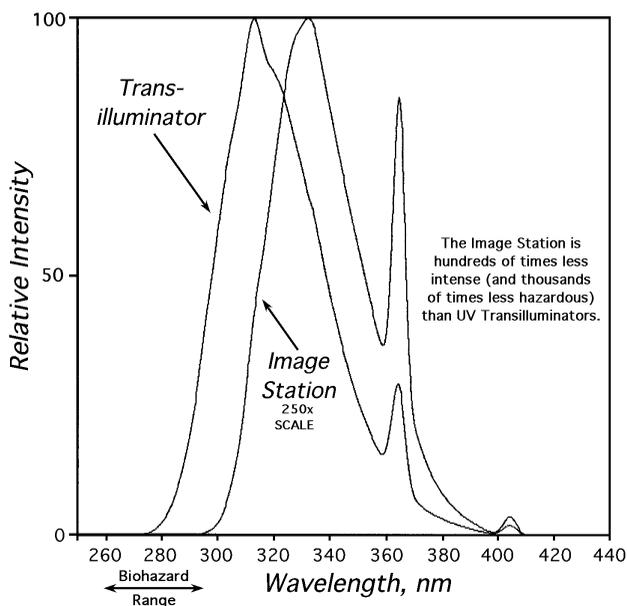


Figure B-1. Emission spectra of UV illumination.

Image Correction

In the following section, you will learn about the principles of image correction as they relate to the IS4000R.

Image capture with a CCD camera has the distinct advantages of speed and nearly live representation of image data. There are associated disadvantages, the most important of which are the image distortions associated with any lens and defects associated with the CCD sensor. Another important image correction is that associated with the non-uniform illumination of the object.

The removal of defective pixel information is automatic. Defective pixels are characteristic of a particular CCD, have elevated signals (dark current), are always present and amount to about 0.1% of all pixels. Defective pixels in an image are corrected by recalculating their signal value by interpolating the values of their nearest neighbor pixels. For long exposures times occasional “bright” (or high-signal) pixels may be observed. Generally, you will find those pixels will not be reproduced in other exposures. The “spontaneous” bright pixel cannot be prevented and is likely the result of a spurious charge on the sensor or other spontaneous event such as background gamma-rays

The lens distorts the image of a flat object in two significant ways: the object intensity is reduced as a function of the distance from the center of the image, and the magnification of the image is slightly increased as a function of the same distance (referred to as spherical aberration or “pin-cushion” distortion). Some distortion is apparent using the widest field of view.

A distinct advantage of the IS4000R system is that a flat object on the Platen is at a fixed distance from the camera. Consequently, the extent of image distortion is a function of the magnification (FOV) only. The IS4000R image distortion has been characterized as a function of FOV, and the appropriate correction may be applied by selecting the Warping option in the IS4000R Acquire window (Preferences). Since the extent of distortion is a function of magnification, care must be taken to accurately record the lens FOV setting in the acquisition interface.

Most important to quantitative imaging is the intensity correction, since any lens will cause a reduction of object intensity as a function of its distance from the center of the image (vignetting). The corresponding correction is a complicated function of both f -stop and FOV. The IS4000R image intensity has been characterized as a function of f -stop and

FOV, and the appropriate correction may be applied by selecting the Lens Correction option in the Preferences section of the IS4000R Acquire window. Since the extent of intensity correction is a function of both f -stop and FOV, care must be taken to accurately record both lens settings in the acquisition interface. Upon accurately providing that information, an appropriate image transform is applied to the image to correct all intensity information throughout the field of view. This correction will apply to any luminescent image and transmission images in which care is taken (proper use of the Light Diffuser) to supply uniform illumination to the Platen. Use of a filter should not significantly influence the correction. Except for the very corner of the field at the widest angle and the lowest f -stop, the representation of the intensity will conform to a nominal 1% accuracy. Application of the lens correction does not change the global mean signal of the image.

For UV illuminated images, field illumination correction should also be applied. The extent of UV illumination non-uniformity will cause approximately the same amount of quantitation error as the lens distortion of the IS4000R. It is critical to capture a reference image (to be used for image correction) of the illumination field using optical parameters (f -stop and FOV) that are identical to an image that is to be corrected. The illumination non-uniformity is highly reproducible and may be corrected by mathematically dividing an image by a reference image captured by the IS4000R, using the protocol outline in the *Generating an Illumination Reference File* in Chapter 5: *Capturing Images*.

The reference image used for correction will be filtered to remove noise and artifacts (such as scratches and particulate dust). The filtered reference image is normalized and divided into the experimental image to be corrected. Regions of the experimental image that experienced a greater-than-average illumination are divided by a number greater than one, while those experiencing a less-than-average illumination are divided by a number less than one. As with the lens correction, the global mean signal of the image is unchanged by illumination correction. Application of illumination correction precludes the need for lens correction.

It is recommended that multiple reference images at varying f -stops and field of view (FOV) be captured and clearly designated as a library of images from which reference images may be selected for use in illumination correction. For the sake of accuracy, create reference files at all f -stop and FOV settings commonly used, and make certain that the settings are made and documented precisely; it is suggested that only f -stop and FOV adjustments that are indexed be chosen for experiments or reference files.

It is likely that such a library could be applied for an extended time period, because the field illumination should be constant for many hundreds of hours. A suggested library of

UV illumination reference files is listed in following table. (18 reference files).

Table 1: Suggested library of UV illumination files

<i>f</i> -stop Setting	FOV Setting
0	20, 30, 40, 60, 100, 150
4.0	20, 30, 40, 60, 100, 150
5.8	20, 30, 40, 60, 100, 150

Glossary

Analog Signal—A signal that is continuous and uninterrupted. Variations in voltage correspond to variations in brightness. This differs from digital, where a signal is represented in discrete steps of digital value (1's, 0's).

Analog to Digital (A/D) Converter—An integrated circuit that transforms an analog (voltage level) to a binary (or digital) value. The IS4000R camera converts the analog signal into a digital signal using an A/D converter.

Aperture—The opening within a lens that limits light passage, usually adjustable and referred to as the *f*-stop.

Autoradiography—A method of capturing radioactive signals by contact-exposure of film to a flat sample consisting of radioactive objects or labeled molecules.

Band-Pass Filters—Clear optical elements that transmit only a limited spectrum of light (limited range of wavelengths) and may be of two general types: colorimetric or interference.

Binning—The process by which adjacent CCD signal values are added together and then each pixel value is replaced with the new added value. For example, if two adjacent pixels have an intensity value of 100 and 150, when binning has been applied, both pixels will be assigned a value of 250. Binning can be applied on X and/or Y axis of an image. While binning decreases the resolution of the image, it increases the sensitivity of the signal.

Bit (Binary Digit)—The smallest amount (unit) of information, measured on a binary scale (integer exponents of 2), where combinations of 0's and 1's are used to code information. See *Bit Depth*. Eight bits of information is called one byte.

Bit Depth (Bits-Per-Pixel or Pixel Depth)—The number of intensity values that can be assigned to each pixel. Images usually fall between 8 and 24 bits. The IS4000R camera produces a 16-bit image in the single capture mode or n-bit in the multiple capture mode. A 12-bit image has 4,096 gray levels, 14-bit image has 16,384 gray levels, while a 16-bit image has 65,536 gray levels.

Black Point—Corresponds to an intensity value in the image that represents pure black in the screen image. The black point can be adjusted using the histogram sliders to help visualize different features in the image. Features with intensity values below the black point can no longer be seen in the screen image. Adjustments to the black point alter the screen image and do not alter the data that is transferred to KODAK Molecular Imaging Software for analysis.

Blooming—If the CCD is overexposed, the CCD pixels do not have the capacity to hold the charge and will “bloom” or spill into the adjacent pixel. Blooming distorts the image. However, excessive signal may often be associated with the property of the object and not the blooming of the CCD.

Blot—A methodology in which molecules/particles of interest are transferred and affixed to a solid support (membrane), usually for the sake of detection and imaging. A liquid sample may be applied to a membrane as a spot, dot, or slot (band), defining a formatted sample array as a Dot or Slot Blot. Electrophoresis gels used to length-resolve DNA, RNA or protein molecules may be similarly transferred to membranes by mechanical extrusion or electrophoresis; such blots are respectively called Southern, Northern, or Western Blots.

Brightness—A relative measure of light associated with a pixel representing its gray level from black and white, through intermediate levels of gray. Perceived brightness increases from dark to bright, or black to white through intermediate levels of gray. However, the convention of quantitative imaging is quite the opposite, wherein the gray scale is increased from white to black.

CCD (Charged-Coupled Device)—An electronic sensor used for imaging light. A silicon crystal, etched to produce micro-electronic circuitry, for the purpose of transforming a real image into an electronic image. A photon impinging upon the silicon is converted to an electron (a negative charge, stable in the crystal matrix), that may be subsequently managed (collected and transported) by the circuitry. The CCD in the IS4000R has 2,048 x 2,048 = 4,194,304 imaging elements (pixels) defined by etchings.

Chemifluorescence—Chemically mediated production of a fluorochrome. Fluorescent molecules may be produced by the chemical (enzymatic) conversion of a non-fluorescent molecule (substrate), upon excitation (laser or UV illumination).

Chemiluminescence—Chemically mediated production of light. Luminescence or light emission may be produced by a chemical reaction or an enzyme operating on a substrate.

Contrast—The contrast of an image represents the perceived differences in intensity between dark and light areas within the image. A low contrast image contains gray levels that are similar in visual intensity, whereas a high contrast image contains extreme differences in visual intensity.

COPI (Closed Optical Path Image)—Chamber design used in the IS4000R. Provides a light-tight environment for imaging laboratory samples, reducing the possibility of contamination of the optical path of the instrument.

Cosmic Ray—a event in nature that generates a photon which can expose a pixel on a CCD.

Dark Current Noise—Electronic noise inherent to the CCD sensor and circuitry, which is increased with extended exposure time. Cooling the CCD helps reduce dark current noise. The IS4000R camera is capable of about 1 hour exposure times with only a doubling of noise.

Digital Camera—A camera that uses an electronic sensor to record an image in a digital binary format.

Dynamic Range—The maximum range of significant digits over which system hardware performs electronic imaging, usually expressed as a signal-to-noise ratio. The IS4000R camera provides a 16-bit (65,536 gray levels) signal range for single captures. These single captures can be accumulated by addition in software into a n-bit file.

Electrophoresis—Separation of molecules on the basis of charge and size.

Epi-illumination—Illumination of an object in a direction other than the optical axis defined by an object and sensor. IS4000R illuminates objects at a 45° acute angle from the optical axis of the lens.

Filter Cassette—A predefined shaped cassette containing UV filters for fluorescent capture mode.

Floating Point—A means of representing a signal as real numbers (fractions or decimals). Floating point calculations are important in maintaining the accuracy of analysis data. The KODAK Molecular Imaging Software uses floating point numbers.

***f*-stop**—The ratio of focal length to the aperture diameter in the lens.

Fluorescence—The emission of light from a substance having been excited by light absorption.

Gamma—A mathematical transformation function that can be used to improve image appearance by decreasing or increasing the contrast of an element of interest in an image. Adjusting the gamma of an image disproportionately skews the gray level distribution, higher gamma values lighten the image and lower gamma values darken the image. Adjusting the gamma does not alter the image data file and is only used to enhance the viewing of the image.

Gel—A separation matrix which is typically agarose or acrylamide and is used for electrophoresis.

Gray Level—The digital signal assigned to a pixel associated with a level of light (from black to white). For example, an 8-bit and a 16-bit system includes gray level values between 0–255 and 0–65,535, respectively.

Histogram—The frequency of the distribution of pixels over the range of signals within an image. The horizontal axis represents the gray level and the vertical axis represents the number of pixels. See Appendix A.

Image Accumulation—The addition of multiple exposures (also referred to as frame captures). The data from all the summed images are used to generate the final image for analysis.

Image Compression—A computational operation upon image data that results in a reduction of data storage volume. Usually results in some loss of image data.

Interpolation—A numerical estimate of a value within a range of empirical data, based on the mathematical trend of data. Contrasts with extrapolation, in which a value outside the range of data is estimated.

Noise—Random and/or persistent deviations in signal. In a digital camera the CCD has noise associated with the collection, reading and amplification of the signal that may affect image quality. The IS4000R camera reduces noise by cooling the CCD using thermoelectrical cooling. Also see *Dark Current Noise* and *Read Noise*.

Photon—The smallest unit of light, or any electromagnetic radiation. Characterized by wavelength.

Pixel—The fundamental element in a digital image. In a digital camera, the pixels represents the light sensitive elements on the CCD.

rad PADD—(radioisotopic phosphor assisted detection device) is an optional cassette accessory that converts beta particles emitting from radioisotope samples into light which can be imaged on the IS4000R.

Read Noise—Electronic distortions in signal introduced when converting the analog signal to a digital signal.

Resolution—The capability to distinguish between objects of interest. It is customary, when describing the characteristics of a digital imaging device or image, to describe the resolution by specifying the number of pixels it captures in the horizontal by vertical direction.

Saturation—The limitation of signal imposed by either the CCD or the digital scale in which the signal is being represented. If the signal becomes too large the individual pixels on the CCD will be filled and no additional signal can be detected.

Thermoelectrical Cooling—A system that uses electron flow through semiconductors to generate temperature differences. The CCD is mounted on the cool side and the heat is dissipated by cooling fans. IS4000R uses a thermoelectrical cooling system to reduce image noise generated by the heat from within the camera's CCD.

TIFF—(Tagged Image File Format) is an industry standard file format for storing images. Images from KODAK Molecular Imaging Software can be saved in TIFF format for use with other computer programs.

Transmission—Mode of image capture, usually absorbance, in which light passes through the illuminated object.

Ultraviolet Light (UV)—Generally refers to the spectrum of electromagnetic energy from about 4 to 400 nm in wavelength. Wavelengths of 200–400 nm are useful in the usual laboratory environment.

Vignetting—An imaging artifact that is associated with the limitation of the lens. Vignetting generally appears as signal reduction around the edges of the image.

White Light—Light that is in the visual part (400–700 nm) of the spectrum (daylight, regular light bulbs).

White Point—Corresponds to an intensity value in the image that represents pure white in the screen image. The white point can be adjusted using the histogram sliders to help visualize different features in the image. Features with intensity values above the white point can no longer be seen in the screen image. Adjustments to the white point alter the screen image and do not alter the data that will be transferred to KODAK Molecular Imaging Software for analysis.

X and/or Y Binning—See *Binning*.

Index

A

- About the User Guide 1-2
- Acquire software
 - launching 4-6—4-7, 8-23
 - Preferences 3-29—3-30
 - Window 3-4—3-17
- Acquire Window 3-4
 - Buttons section 3-4, 3-17
 - Exposure section 3-4, 3-7—3-12
 - Illumination section 3-4, 3-14—3-15
 - Illumination reference 3-13
 - Set Camera To section 3-16
 - User section 3-4, 3-5—3-6
- Analyze button 3-20—3-21, 3-22—3-23, 3-24—3-25, 3-27—3-28
- Annotation bar 3-5—3-6
- Automatic Illumination Reference File 5-23

B

- background subtraction A-10
- binning A-9, G-1
 - applying 3-7—3-8, 3-19, 3-10, 3-11—3-12, 5-3, 5-9, 5-14, 5-19

C

- Cable Connector, Camera 2-3, 8-13, 8-21
- Camera
 - About 2-10
 - computer interface cable 2-3, 8-13, 8-21
 - FOV ring 2-11, 4-11
 - f*-Stop ring 2-11, 4-16, 4-17
 - Focus ring 2-11, 4-11—4-12, 4-18
 - Focusing Tool 4-18

- Lens 2-11, 4-11—4-12
 - noise A-7
- Capture System Chamber 2-10—2-11
- Capture Status windows 3-19—3-28
 - Progressive 3-2, 3-24—3-25, 5-16—5-17
 - rad PADD 3-3, 3-26—3-28, 5-18—5-22
 - Standard, single 3-1, 3-19, 5-5
 - Standard, multiple 3-1—3-2, 3-20—3-21, 5-5—5-7
 - Time Lapse 3-2, 3-22—3-23, 5-11—5-12
- Charged-Coupled-Device (CCD) 2-10, G-2
- Circuit Breaker 2-3
 - cleaning 6-2
- Closed Optical Path Image (COPI) Chamber 2-7—2-9
- Compression Pad 2-4, 2-5
- conventions, User's Guide 1-2
- copy protection device 8-11, 8-18
- corrections C-1—C-3
 - illumination 3-13, 3-29—3-30, 5-23—5-25
 - lens 3-29—3-30
 - warping 3-29—3-30
- custom capture settings 5-26—5-27

D

- digital imaging concepts A-1—A-10
- decontamination 8-30—8-31, 9-3—9-4
- dynamic range A-6, G-3

E

- Exposure
 - Progressive 3-2, 3-10, 5-13—5-17

rad PADD 3-3, 3-11—3-12, 5-18—5-22
Standard 3-1—3-2, 3-7—3-8, 5-2—5-7
Time Lapse 3-2, 3-9, 5-8—5-12
Time text edit box 3-7, 3-9, 3-10—3-11
types 3-1—3-3

F

f-Stop

ring 2-11, 4-16, 4-17
setting 2-11, 3-16, 4-16
slider 3-4, 3-16

file

Illumination Reference 3-13, 3-29—3-30, 5-23—5-25

filter

applying 4-12—4-13
changing 6-3—6-5
choosing 4-12—4-13
Selection Dial 2-11, 4-12

Filter Wheel 2-12

Access Panel 2-12

FireWire - See IEEE 1394

Focusing Tool 4-18

Focus Ring 2-11, 4-10—4-11, 4-18

FOV

ring 2-11
slider 3-4, 3-16, 4-11
setting 4-11

H

hardware

overview 2-1—2-12
setup 8-3
Transilluminator Lid 8-4—8-7

I

IEEE 1394

II

Camera Cable Connector 2-3, 8-13, 8-21

installing 8-3

Illumination

automatic Correction 5-23
choosing 4-12—4-15
correction C-1—C-3
Reference File 3-13, 3-29—3-30, 5-23—5-25
Reference Library 5-24—5-25
UV epi-illumination 4-2, 4-13
UV fluorescence quenching 4-13
UV transillumination 4-2, 4-13
white light transillumination 4-3, 4-14

Image

capture settings 4-17
illumination correction 3-13, 3-29—3-30, C-1—C-3
lens correction 3-29—3-30
orientation 4-8—4-9
previewing 4-10—4-11
saturation 3-29—3-30
warping 3-29—3-30

Imaging Software, KODAK Molecular

installation 8-8—8-22
launching 4-6—4-7, 8-23
package contents 1-4
system requirements 1-5

installation

copy protection device 8-11, 8-18
hardware 8-3
IEEE 1394 (FireWire) card 8-3
Molecular Imaging Software 8-8—8-22

L

launching software 4-6—4-7, 8-23

lens

Focusing Tool 4-18
Focus ring 2-11, 4-10—4-11, 4-18

-
-
- f*-stop 2-11, 3-4, 3-16, 4-16, 4-17
- Lid
- clasp 2-4
 - compression pad 2-4
 - illuminator bulb 6-7
 - transilluminator 2-5, 8-4—8-7
- luminescence 4-2
- M**
- Macintosh
- requirements 1-5
 - software installation 8-14—8-22
- maintenance 6-1—6-7
- N**
- noise, digital imaging systems A-7—A-8
- P**
- package contents
- IS4000R 1-4
 - Molecular Imaging Software 1-4
- Platen 2-5—2-6
- care 2-6, 6-2
- Preferences
- button 3-17
 - dialog box 3-29—3-30
- Preview window 3-18, 4-10—4-11
- Progressive Exposure 3-2, 3-10, 5-13—5-17
- R**
- Radiator 2-2
- radioisotopic 2-13, 4-4
- sample handling 4-9
 - decontamination 6-6, 8-30—8-31
- rad PADD 2-13, 4-4
- cleaning 6-6
 - exposure 3-3, 3-11, 5-18—5-22
 - sample orientation 4-8—4-9
 - using 3-11—3-12
- registering 1-6
- regulatory information 9-9
- repacking 8-30—8-31
- requirements
- electrical 8-2
 - environment 8-2
 - software 1-5
 - space 8-2
- resolution
- image A-2—A-3
 - spatial A-4—A-5
- S**
- Sample
- orienting 4-8—4-9
 - Platen 2-4
 - Work Area 2-4—2-6
- saturation 3-29—3-30, G-5
- saving settings 3-5—3-6
- software
- installation 8-8—8-22
 - launching 4-6—4-7, 8-23
 - license agreement 9-6—9-8
 - package contents 1-4
 - preferences 3-29—3-30
 - previewing 4-10—4-11
 - system requirements 1-5
- Standard Capture 3-1—3-2, 3-7—3-8, 5-2—5-7
- Stop Preview button 3-18, 4-10
- System, IS4000R
- components 1-4
 - front view 2-2
 - hardware 2-1—2-12
 - installation 8-3
 - Lid, standard 2-4
 - Lid, Transillumination 2-5
 - maintaining 6-1—6-7
 - rear view 2-3
 - registering 1-6
 - regulatory information 9-9
 - repacking 8-30—8-31
 - requirements 1-5, 8-2
 - software 3-1—3-30
 - troubleshooting 7-1—7-7

T

Technical Support 1-7, 7-2
Time Lapse Exposure 3-2, 3-9, 5-8—5-12
Troubleshooting 7-1—7-7
 images 7-4—7-6
 instrument 7-3
 software 7-7

U

updating settings 3-4, 3-5—3-6, 5-26—5-27
user name option 3-4, 3-5—3-6
User's Guide
 about the 1-2
 conventions 1-2
 navigating 1-3
 Molecular Imaging Software 1-2
User section 3-4, 3-5—3-6
UV illumination 4-2, B-1—B-3

epi 4-2
trans 4-2

W

warranty 9-1—9-5
 coverage 9-2
 limitations 9-5
 registration 1-6
 service 9-2
 time period 9-2

Windows

 requirements 1-5
 software installation 8-8—8-22
white light transillumination 4-3